

Short Title: TMA: role of complement

**The Role of Complement in Autoimmune Disease-Associated Thrombotic
Microangiopathy and the Potential for Therapeutics**

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Conflict of Interest:

AJ serves on the scientific advisory boards of Alexion, AstraZeneca Rare Disease, and Novartis International AG, and serves as a consultant for Chinook Therapeutics. She is also a Principal Investigator for Apellis Pharmaceuticals.

AK reports personal fees from Alexion, AstraZeneca Rare Disease, AstraZeneca, Aurinia Pharmaceuticals Inc., Exagen Diagnostics Inc., GlaxoSmithKline (now GSK plc), Kypha Inc., and Pfizer, and research grant support to Washington University School of Medicine from Foghorn Therapeutics and GSK plc.

This article has been accepted for publication in The Journal of Rheumatology following full peer review. This version has not gone through proper copyediting, proofreading and typesetting, and therefore will not be identical to the final published version. Reprints and permissions are not available for this version. Please cite this article as doi: 10.3899/jrheum.220752. This accepted article is protected by copyright. All rights reserved.

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Key Indexing Terms (max. 6, must be MeSH terms): thrombotic microangiopathy, complement, systemic lupus erythematosus, antiphospholipid syndrome, neutrophil activation, systemic scleroderma

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ABSTRACT

The complement system is a tightly regulated, cascading protein network representing a key component linking the innate and humoral immune systems. However, if misdirected or dysregulated, it can be similarly damaging to host-tissue. The role of complement dysregulation on vascular endothelial cells has been well established in atypical hemolytic uremic syndrome (aHUS), a thrombotic microangiopathy (TMA) characterized by microangiopathic hemolytic anemia, thrombocytopenia, and target organ injury. Yet a great deal of complexity exists around the role of complement in TMA associated with other diseases. A further complicating factor is the cross-talk between complement, neutrophils, and coagulation pathways in the pathophysiology of TMA. Advancements in the understanding of the etiopathogenesis of aHUS paved the way for the successful development of anti-complement therapies (complement C5 inhibitors), which have revolutionized the treatment of aHUS. Therefore, a clearer understanding of the role of the complement system in TMA associated with other conditions will help to identify patients who would benefit from these therapies. This review aims to provide an assessment of the nature and extent of complement involvement in TMA associated with autoimmune diseases such as systemic lupus erythematosus, antiphospholipid syndrome, and scleroderma renal crisis. Defining the role of complement in TMA in these conditions will help to guide timely diagnosis and management.

INTRODUCTION

Thrombotic microangiopathy (TMA) is a well-known clinicopathologic entity characterized by microangiopathic hemolytic anemia, thrombocytopenia, and organ injury.¹⁻³ The pathological features of TMA are endothelial cell damage and microthrombi formation in small blood vessels, leading to a partial or complete obstruction of the vessel lumina.¹⁻³ Acute kidney injury is a common prominent feature of the disease, owing to the susceptibility of the glomerular circulation to endothelial damage.^{2,4,5} Extrarenal manifestations (occurring in up to 38% of patients) may include strokes, seizures, and/or involvement of the cardiovascular, pulmonary, or gastrointestinal systems.^{2,4-6} Early recognition is important because TMA is associated with significant mortality and morbidity, including end-stage kidney disease, although prompt initiation of supportive and specific management can transform disease outcomes.^{2,7} There are several underlying pathophysiological mechanisms associated with TMA.⁷⁻⁹ A TMA is called ‘primary’ when a genetic or acquired defect in a complement protein is identified (as in atypical hemolytic uremic syndrome [aHUS]) or ‘secondary’ when occurring in the context of another disease process such as infection, autoimmune disease, malignancy, or drugs.¹⁰ This distinction is not absolute because genetic defects in complement proteins have been identified in secondary TMA.¹⁰ Differentiating between a primary complement-mediated process and one triggered by secondary factors is critical since the former is non-responsive to supportive therapy and has a high risk of recurrence.¹⁰ This review aims to provide an assessment of the nature and extent of complement involvement in the underlying pathophysiology of TMA associated with autoimmune diseases that will help to stratify patients for targeted therapy.

COMPLEMENT PATHWAYS

The complement system is a tightly regulated, cascading protein network that performs multiple roles in homeostasis and disease prevention and is a key component of both the innate and the humoral immune systems.^{8,11-14} Numerous stimuli can drive the activation of the complement system, including apoptotic debris, pathogens, and antibody–antigen complexes, in addition to ischemia–reperfusion injuries associated with organ transplantation.^{4,13} Complement plays a crucial role in host defense against foreign bodies by promoting phagocyte-mediated clearance of cell debris through activation of an inflammatory response, opsonization of pathogens, and lysis of susceptible bacteria and cells.^{11,12,15}

Activation of the complement system occurs via the classical (CP), lectin (LP), or alternative (AP) pathways. The CP is initiated by the binding of antibody to antigen, the LP is initiated by the binding of lectin to an oligosaccharide, and the AP is initiated by the binding of one of its components to a pathogen, without need for prior contact/exposure.¹⁶ The AP is thus constitutively active and turns over continuously, generating small amounts of auto-activated C3 (the so-called C3 tick-over that represents a thioester-hydrolyzed form of C3, termed C3[H₂O]).¹⁶ Central to all three pathways is the formation of C3 convertase, which cleaves complement component C3 to C3a and C3b. If C3b deposits on a microbe or foreign debris, the system can be rapidly amplified by engaging two proteases, factors B and D, along with a stabilizing protein properdin (P) to create the powerful AP C3 convertase. Contained within the AP is an efficient feedback or amplification loop for generating large amounts of C3b for opsonization of pathogens, leading them to be recognized, engulfed, and destroyed by phagocytes. The addition of C3b to C3 convertase in all three pathways generates C5 convertase, which leads to the activation of the terminal complement pathway. In the terminal pathway, two types of C5 convertase (C4bC2C3b and C3bBbC3b) can cleave the C5 protein into anaphylatoxin C5a and the larger C5b component, resulting in the formation of the

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membrane attack complex (MAC, C5b-9; Figure 1).¹¹ This complex penetrates membrane bilayers to form pores that disrupt the osmotic barrier, leading to swelling and cell lysis.^{11,12}

The misdirection or dysregulation of the complement system can lead to indiscriminate host inflammation and tissue injury.^{4,15,17,18} Thus, nearly half of the components of the complement system are responsible for the stringent regulation of the arsenal.¹⁹ Disruption of this delicate balance, for example by inherited or acquired deficiencies in its activating or control proteins, is increasingly implicated in disease pathology.¹⁰ Insights gained from genetic studies over the past decade have established that modulation of complement inhibitory activity predisposes to TMA.¹⁰ Although understanding of the complement system is increasing, a lack of clarity remains around the etiology, triggers, and scope of complement activation, as well as the relationship of these factors in the various types of TMA.

Determining the underlying cause can be a challenge but is important for directed therapy.

MECHANISMS OF COMPLEMENT ACTIVATION IN TMA

Complement dysregulation

Complement dysregulation stems from intrinsic genetic factors, due to abnormalities in genes encoding complement components and regulatory proteins, or due to acquired defects in the complement system (such as autoantibodies). A classic example of a ‘primary’ TMA resulting from complement dysregulation is aHUS. In this disease, a loss-of-function mutation in a regulator (such as factor H, factor I, or membrane cofactor protein) or a gain-of-function mutation in a complement component (such as C3 or factor B) predispose patients to endothelial damage (Figure 2). Sometimes, a genetic defect, namely a homozygous deletion in factor H-related proteins 1 and 3, is associated with the development of acquired factors, such as factor H autoantibodies.² These autoantibodies can occur in patients with aHUS and may inhibit the regulatory function of factor H. The presence of factor H autoantibodies

varies between different ethnic populations and has been reported in 11% of patients in the USA, 9% of European patients, and 56% of Indian patients with aHUS.²⁰⁻²²

However, TMA can also occur in response to or following certain conditions or ‘triggers’, including systemic infections, pregnancy, solid organ or hematopoietic stem cell transplantation, certain metabolic disorders, or cancer, as well as in autoimmune diseases such as systemic lupus erythematosus (SLE).^{2,4,23} TMA occurring in the setting of any of the above conditions is often thought to be due to ‘short-lived’ complement overactivation secondary to the associated condition, and is therefore called a ‘secondary’ TMA. However, genetic abnormalities of the complement system have been reported in approximately 10–60% of patients with a ‘secondary’ TMA.^{2,10} In such situations, the complement system may be initially activated in response to the associated condition (such as infection or SLE), but may evolve into dysfunctional control owing to an underlying, and potentially unrecognized, genetic abnormality. Such patients in whom a complement mutation is identified in the presence of concomitant autoimmune disease can be considered to essentially have aHUS or ‘primary’ TMA, with the autoimmune condition functioning as a trigger.

TMA occurring because of an underlying genetic etiology progress despite removal of the precipitating cause, with poor long-term outcomes in the absence of timely treatment.^{24,25}

Therefore, making the distinction between a primary TMA resulting from underlying genetic complement dysregulation leading to overactivation and a secondary TMA involving transient complement overactivation is critical, but can be challenging for clinicians.² Patients in whom the clinical course of an assumed secondary TMA is unusually aggressive and unresponsive to conventional treatment should be considered for treatment with anti-complement therapy. Additionally, genetic testing and/or complement biomarker assessments should be pursued to establish complement involvement in the disease in greater detail.

Several real-world cases have been published that illustrate how sequential and systematic analyses can help to determine disease etiology, pathogenic mechanisms, prognosis, and duration of therapy based on individual risk assessment.^{26,27}

Neutrophil activation

The pro-inflammatory complement cleavage products C3a and C5a activate a range of immune cells, including neutrophils, mast cells, monocytes/macrophages, basophils, eosinophils, and T and B cells (Figure 3).²⁸ This pro-inflammatory response is very potent, and leads to increased expression of inflammatory cytokines such as tumor necrosis factor- α and interleukin (IL)-1 β and IL-6.²⁸ Activated neutrophils create large chromatin structures known as neutrophil extracellular traps (NETs), formed via a type of programmed cell death known as NETosis, which is further stimulated by a number of inflammatory cytokines.^{29,30} NETs incorporate complement components such as C3 cleavage products, properdin, and complement factor B, which can form a C3 convertase leading to exuberant AP complement activation (Figure 3).^{28,31,32} C3a and C5a anaphylatoxins released during neutrophil-driven complement activation can act to further amplify pro-inflammatory neutrophil responses, such as additional NET formation, enhanced CD11b expression, and oxidative burst, again illustrating the multifaceted interaction between neutrophils and the complement system.³¹⁻³⁵ NETs also play a role in non-AP mediated complement activation and the generation of the MAC.³²

Importantly, NETs have been shown to be directly cytotoxic and act as a scaffold for thrombus formation, activating coagulation and eventually forming part of the resulting thrombi.³²⁻³⁴ NETs have been implicated in the development of thrombophilic conditions such as antiphospholipid syndrome (APS), suggesting that NET formation may exert secondary effects such as cell injury and death, in addition to triggering complement

activation.^{32,33} Furthermore, autoantibodies targeting NETs, identified in conditions such as APS and COVID-19, may potentially impair NET clearance and enhance ongoing complement activation.^{33,34} Anti-NET antibodies may act to shield NETs from degradation by deoxyribonuclease (DNase), and may themselves be associated with venous thrombosis in patients with APS.³³ NETs may also be stabilized by complement components such as C1q, further preventing DNase-mediated clearance.³⁵ This complex interplay between the complement components, inflammatory mediators, and coagulation proteins results in endothelial injury and a hypercoagulable state leading to TMA.^{29,33}

AUTOIMMUNE DISEASE-ASSOCIATED TMA

Systemic lupus erythematosus

SLE is a multisystem complex disorder distinguished by the presence of autoantibodies to multiple nuclear antigens, including DNA and ribonucleoproteins, leading to complement activation, inflammation, and tissue injury.³⁶ Virtually any organ may be affected, leading to diverse clinical presentations.^{36,37} TMA in SLE has been reported in <1–9% of affected patients, with multiple mechanisms leading to the pathogenesis and autoimmunity observed in the disease.³⁸⁻⁴⁰

Deficiencies or low gene copy number of the early CP components are the strongest genetic risk factors for SLE,^{36,41} because severe forms of the disease are observed in patients with defects in C1q, C2, or C4.⁴¹ Notably, homozygous mutations of C1q, C1r, or C1s leading to a complete absence of the protein demonstrate the highest genetic penetrance for SLE, with 88% of patients displaying SLE or lupus-like disease.⁴² C4 deficiency due to low copy numbers of the gene has also been identified as a risk factor for the development of SLE.⁴³

These factors are also potential mechanisms for the generation of autoantibodies that bind to host proteins or deposit within tissues as a component of immune complexes,³⁶ and further

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trigger activation of the complement system.^{36,44} Autoantibodies against C1, C5, and factor H have been reported in SLE.^{41,45} Factor H autoantibodies can also arise in patients with SLE, in association with genetic deletions in factor H-related protein 1 (similar to aHUS).⁴⁵

Furthermore, increased levels of complement split products, C3dg, iC3b, and C4d have been observed in SLE and may function as biomarkers.⁴⁶ The ratios of C3dg:C3 and iC3b:C3 also appear to correlate with active disease.⁴⁶ B-cell-bound C4d and erythrocyte-bound C4d may further indicate active lupus, while affected patients display low levels of C3 and/or C4 complement proteins.⁴⁶ Complement-related gene variations, including mutations in factors H, I, and B, CD46, and factor H-related proteins 1–3, have been reported in patients with lupus nephritis-associated TMA.⁴⁷ This suggests that the presence of an underlying genetic predisposition may lead to TMA development in the setting of ongoing complement activation in SLE.^{45,47}

It may seem paradoxical that both a deficiency of the complement system and its excessive activation are both associated with the pathophysiology of SLE.^{36,41} However, this can be reconciled when considering that the excess apoptotic debris following a CP component deficiency serves as a source of nuclear autoantigens for autoantibody formation and binding.⁴⁸⁻⁵⁰ The resultant immune complexes cannot be cleared, leading to pro-inflammatory cytokine production and interferon- α (IFN- α) secretion by plasmacytoid dendritic cells.^{51,52} IFN- α is a hallmark cytokine in SLE, the level of which correlates with disease activity.^{51,52}

NETs have also been under scrutiny because of their role in disease pathogenesis. IFN- α stimulates NET production, with NETs in turn triggering further IFN- α secretion by plasmacytoid dendritic cells.⁵³ Notably, neutrophils in patients with SLE undergo accelerated NETosis, possibly owing to elevated IFN- α .^{54,55} Some patients with SLE have a DNase

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deficiency that leads to impaired degradation of NETs, with the ensuing defect in clearance correlating with the development of lupus nephritis and associated TMA.^{37,56}

The presence of TMA on renal biopsy portends a poor prognosis when associated with lupus nephritis.⁵⁷ It has been reported to be an independent risk factor for poor long-term renal outcomes with 80% of patients with lupus nephritis and TMA developing end-stage kidney disease within 5 years of diagnosis.⁵⁸ Therefore, although long-term immunosuppressant therapy remains the mainstay of SLE management, complement inhibition represents a promising therapy for a subset of patients with TMA.

Antiphospholipid syndrome.

APS is characterized by the development of antiphospholipid antibodies, which bind to endothelial cells and trigger thrombosis. The condition is associated with substantial morbidity and may involve virtually all organ systems,^{36,59,60} resulting in stroke, skin ulcerations, nephropathy, seizures, and cognitive decline.⁶¹ Primary APS is diagnosed when other associated conditions are absent, occurring in over 50% of patients.⁶⁰ APS may also occur in the context of autoimmune diseases such as SLE.⁶⁰ Among patients with SLE, TMA was found in 67% of those with concurrent APS, but only in 32% of those without APS.⁶² Approximately 1% of patients with APS may develop catastrophic APS (CAPS), which is fatal in more than half of cases, manifesting as small vessel thrombosis in three or more organs within the span of a week.^{63,64}

Mounting evidence suggests that the complement system plays a critical role in the pathogenesis of thrombosis in APS, with activation of the complement cascade due to antiphospholipid antibodies causing cellular injury and promoting coagulation via multiple mechanisms.⁵⁹ In murine models of thrombotic APS, C9 deposition has been reported on the vascular endothelium, indicating the presence of the MAC.⁵⁹ Furthermore, antiphospholipid

antibody-induced thrombosis was markedly attenuated in C6-deficient rats or animals treated with a C5 inhibitor.⁶⁵ Investigators have also reported lower plasma C3 and C4 levels and elevated levels of complement activation fragments, particularly Bb and C3a, in patients with APS compared with control individuals.^{66,67} Increased serum MAC has been detected in patients with CAPS, with clinical improvement after treatment with eculizumab correlating with a reduction in serum MAC and normalization of serum C3 and C4.^{68,69} Complement activation is more commonly observed near to the occurrence of a thrombotic event, with 68.5% of samples from patients with APS collected within one year of thrombosis showing evidence of complement activation.⁵⁹ Although these findings and the benefits of complement inhibition support a role for complement in APS,^{70,71} the exact mechanisms of complement activation in the disease and its correlation with vascular events remains incompletely understood.

NETs have been implicated in the pathogenesis of APS, and may aggravate thrombosis through activation of the coagulation cascade and inhibition of anticoagulant factors.⁶¹ Antiphospholipid antibodies themselves promote NET formation, and compared with control individuals, patients with APS have been reported to harbor increased levels of low-density granulocytes, which are prone to exaggerated NETosis.⁶¹ Moreover, NETs formed in APS appear resistant to degradation.^{33,61} Anti-NET antibodies (particularly immunoglobulin M), which may act by stabilizing NETs, are also markedly increased in patients with APS.³³

Although specific biomarkers of APS disease activity are lacking, numerous potential candidates exist for examination, such as NET load, complement activation products, and, in some patients, anti-NET antibodies. When considering disease therapy, the current standard of care for APS is limited to anticoagulation and immunosuppression.⁶¹ The variability of

treatment effectiveness in APS and reports of recurrent thrombosis despite standard treatment suggest that a subset of patients might benefit from treatment beyond anticoagulation.⁶¹

Scleroderma renal crisis. Scleroderma renal crisis (SRC) represents a life-threatening complication of systemic sclerosis and is characterized by the sudden onset of hypertension, TMA, and acute kidney injury.⁷² The underlying pathological mechanisms of SRC include vascular injury, autoantibody production, and fibroblast dysfunction, although the complement system has also been implicated as a factor in development of the disease.⁷³

Whole exome sequencing in patients with SRC identified an association with complement genetic variants, similar to those observed in other forms of TMA such as aHUS, or TMA associated with other conditions (e.g. 35–66% of patients with malignant hypertension-associated TMA have been shown to have mutations in complement genes that produce C3, factor I, CD46, or factor H proteins).^{74,75} Renal deposition of C1q, C4d, and C3b has been observed in SRC, suggesting involvement of the CP, which may be driven by autoantibodies.^{72,76,77} In addition, reduced levels of C3 and factor B have also been noted, suggesting AP activation.^{72,77} These data suggest that complement activation plays a role in SRC and the associated TMA.^{74,77,78}

Similar to APS and SLE, emerging evidence suggests that microparticles released from activated platelets stimulate neutrophils to induce NETosis, leading to abundant NET burden in patients with scleroderma.⁷⁹ Neutrophils have previously been implicated in extrarenal manifestations of scleroderma such as pulmonary arterial hypertension and digital ulcers.^{79,80} Currently, angiotensin-converting enzyme inhibition represents a highly effective therapy for SRC;⁸¹ however, complement inhibitors may represent a potential treatment option for refractory cases of TMA in SRC owing to the fact that they more directly address the primary

cause of SRC.^{72,81,82} Early identification and aggressive treatment is critical to avoid loss of kidney function and other complications.⁷⁷

DIAGNOSIS AND TREATMENT OF AUTOIMMUNE DISEASE-ASSOCIATED TMA

Complement dysregulation and activation play a role in the development of TMA in autoimmune diseases, although defining the extent of complement involvement may be challenging. For some patients, highly complex, multifactorial, and incompletely understood interactions occur across complement activation, coagulation pathways, and neutrophil processes, adding to the challenge. However, a thorough systematic work up will help to delineate the underlying disease etiology and determine treatment.

Diagnostic work-up

Prompt identification of TMA is vital to minimize organ damage and improve patient outcomes (Figure 4). To begin with, thrombotic thrombocytopenia purpura (TTP) should be ruled out through the assessment of ADAMTS13 (a disintegrin and metalloproteinase with a thrombospondin type 1 motif, member 13) activity. Laboratory tests (complete blood count, comprehensive metabolic profile, reticulocyte count, lactate dehydrogenase, haptoglobin, direct Coombs test, and coagulation profile) and/or biopsy (when feasible) should be performed to evaluate organ damage and confirm the presence of TMA.⁸³ Functional hemolytic assays, such as CH₅₀ and AH₅₀, as well as testing for antigenic levels of complement proteins should be used to evaluate the presence of a quantitative defect.^{10,84} Furthermore, evaluation for underlying genetic variants in complement proteins should be conducted. The clinically validated aHUS next-generation sequencing-based panel consists of 15 genes (*ADAMTS13*, *C3*, *CD46*, *CFB*, *CFH*, *CFHR1*, *CFHR2*, *CFHR3*, *CFHR4*, *CFHR5*, *CFI*, *DGKE*, *THBD*, *MMACHC*, and *PLG*). Variants are reported according to Human

Genome Variation Society nomenclature and classified based on the guidelines established by the joint consensus of the American College of Medical Genetics and Genomics, and the Association for Molecular Pathology. If identified, interpretation of variants may require additional functional assays, biomarker testing, or in select cases need for recombinant protein production followed by structure–function assessment of the variants.⁸³ As the technologies allowing examination of molecular phenotypes at the tissue-level improve, future understanding of the impact of complement activation in specific organs will provide valuable insights into both homeostatic and pathogenic mechanisms. Combined genetic, antigenic, functional, and structural evaluations will help to define the role and extent of complement involvement in TMA in autoimmune diseases and will further facilitate the stratification of patients for targeted therapy.^{10,85}

Treatment

In TMA associated with autoimmune diseases, it is a reasonable first step to treat the underlying condition or precipitating factor.⁸⁶ Patients who do not respond to conventional therapy or who have unusually aggressive TMA should be considered for treatment with a complement inhibitor. There are two C5 inhibitors that are currently available: eculizumab and ravulizumab.³¹ Both eculizumab and ravulizumab have received approval from the US Food and Drug Administration and European Medicines Agency for use in patients with aHUS.⁸⁷⁻⁹⁰ Both therapies offer meaningful patient benefit and improved disease outcomes in aHUS as demonstrated in clinical trials, with extensive real-world evidence also illustrating the efficacy and safety profiles of these therapies in this condition (Figure 5).⁹¹⁻⁹⁹ Several other drugs primarily targeting the AP are also in development. These novel complement inhibitors include crovalimab (anti-C5), pegcetacoplan (anti-C3), iptacopan (anti-factor B), danicopan (anti-factor D), and avacopan (anti-complement C5a receptor 1).^{2,100} Blockade of the LP through inhibition of mannose-binding lectin-associated serine protease 2 is also being

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investigated (e.g. narsoplimab)¹⁵ and inhibitors of the initial stages of the CP and LP are also available or undergoing clinical trials.¹⁰⁰ The role of C5 fragments in amplifying neutrophil responses further suggests that C5 inhibitors may be a potential benefit in both complement- and neutrophil-mediated inflammatory diseases. Prospective, randomized controlled trials are required to investigate the utility of anti-complement therapies in TMA associated with autoimmune diseases in more detail, to provide insights on which patients to treat and for how long in this complex and challenging area.^{101,102} Further improvements in the understanding and awareness of the mechanisms underlying complement involvement in different diseases will be of great benefit to clinicians, both in identifying affected patients and in selecting appropriate therapies.

ACKNOWLEDGMENT

Authors fulfilled International Committee of Medical Journal Editors (ICMJE) authorship criteria, and all drafts (including the final draft) were critically reviewed, extensively edited, and approved by the authors. Article preparation was sponsored by Alexion, AstraZeneca Rare Disease. Editorial support was provided by Alexandra Kisbey-Ascott, of Oxford PharmaGenesis, Oxford, UK and was funded by Alexion, AstraZeneca Rare Disease. Radha Narayan, PhD, of Alexion, AstraZeneca Rare Disease reviewed early drafts of this publication for scientific/medical accuracy.

REFERENCES

1. Gavriilaki E, Anagnostopoulos A, Mastellos DC. Complement in thrombotic microangiopathies: unraveling Ariadne's thread into the labyrinth of complement therapeutics. *Front Immunol* 2019;10:337.
2. Java A. Chapter 10 – Thrombotic microangiopathy. In: *Current Progress in Nephrology*, 3rd Edition. Mumbai: TreeLife Media; 2022:181–96.
3. Laurence J, Haller H, Mannucci PM, Nangaku M, Praga M, Rodriguez de Cordoba S. Atypical hemolytic uremic syndrome (aHUS): essential aspects of an accurate diagnosis. *Clin Adv Hematol Oncol* 2016;14 Suppl 11:2–15.
4. Haller H. The role, use and pathophysiology of the complement system. Hot Topics in Nephrology session presented at ERA-EDTA 2021. Available at <https://www.era-edta.org/en/virtual-meeting/#!resources/the-role-use-and-pathophysiology-of-the-complement-system> (accessed August 2021).
5. Hofer J, Rosales A, Fischer C, Giner T. Extra-renal manifestations of complement-mediated thrombotic microangiopathies. *Front Pediatr* 2014;2:97.
6. Schaefer F, Ardissino G, Ariceta G, et al. Clinical and genetic predictors of atypical hemolytic uremic syndrome phenotype and outcome. *Kidney international* 2018;94:408-18.
7. Bayer G, von Tokarski F, Thoreau B, et al. Etiology and outcomes of thrombotic microangiopathies. *Clin J Am Soc Nephrol* 2019;14:557–66.
8. Lemaire M, Noone D, Lapeyraque A-L, Licht C, Frémeaux-Bacchi V. Inherited kidney complement diseases. *Clin J Am Soc Nephrol* 2021;16:942–56.
9. George JN, Nester CM. Syndromes of thrombotic microangiopathy. *N Engl J Med* 2014;371:654–66.
10. Palma LMP, Sridharan M, Sethi S. Complement in secondary thrombotic microangiopathy. *Kidney Int Rep* 2020;6:11–23.
11. Sarma JV, Ward PA. The complement system. *Cell Tissue Res* 2011;343:227–35.
12. Rus H, Cudrici C, Niculescu F. The role of the complement system in innate immunity. *Immunol Res* 2005;33:103–12.
13. Scharz ND, Tenner AJ. The good, the bad, and the opportunities of the complement system in neurodegenerative disease. *J Neuroinflammation* 2020;17:1–25.
14. Ricklin D, Hajishengallis G, Yang K, Lambris JD. Complement: a key system for immune surveillance and homeostasis. *Nat Immunol* 2010;11:785–97.
15. Mastellos DC, Ricklin D, Lambris JD. Clinical promise of next-generation complement therapeutics. *Nat Rev Drug Discov* 2019;18:707–29.
16. Nesargikar PN, Spiller B, Chavez R. The complement system: history, pathways, cascade and inhibitors. *Eur J Microbiol Immunol (Bp)* 2012;2:103-11.
17. Thurman JM, Holers VM. The central role of the alternative complement pathway in human disease. *J Immunol* 2006;176:1305–10.
18. Holers VM. The spectrum of complement alternative pathway-mediated diseases. *Immunol Rev* 2008;223:300–16.
19. Noris M, Remuzzi G. Overview of complement activation and regulation. *Semin Nephrol* 2013;33:479-92.
20. Sinha A, Gulati A, Saini S, et al. Prompt plasma exchanges and immunosuppressive treatment improves the outcomes of anti-factor H autoantibody-associated hemolytic uremic syndrome in children. *Kidney international* 2014;85:1151-60.
21. Moore I, Strain L, Pappworth I, et al. Association of factor H autoantibodies with deletions of CFHR1, CFHR3, CFHR4, and with mutations in CFH, CFI, CD46, and C3 in patients with atypical hemolytic uremic syndrome. *Blood* 2010;115:379-87.

22. Zhang Y, Ghiringhelli Borsa N, Shao D, et al. Factor H Autoantibodies and Complement-Mediated Diseases. *Front Immunol* 2020;11:607211.
23. Goodship TH, Cook HT, Fakhouri F, et al. Atypical hemolytic uremic syndrome and C3 glomerulopathy: conclusions from a “Kidney Disease: Improving Global Outcomes” (KDIGO) Controversies Conference. *Kidney Int* 2017;91:539–51.
24. Praga M, de Córdoba SR. Secondary atypical hemolytic uremic syndromes in the era of complement blockade. *Kidney Int* 2019;95:1298–300.
25. Cavero T, Rabasco C, López A, et al. Eculizumab in secondary atypical haemolytic uraemic syndrome. *Nephrol Dial Transplant* 2017;32:466–74.
26. Ren Z, Perkins SJ, Love-Gregory L, Atkinson JP, Java A. Clinicopathologic Implications of Complement Genetic Variants in Kidney Transplantation. *Front Med (Lausanne)* 2021;8:775280.
27. Java A, Pozzi N, Love-Gregory LD, et al. A Multimodality Approach to Assessing Factor I Genetic Variants in Atypical Hemolytic Uremic Syndrome. *Kidney Int Rep* 2019;4:1007-17.
28. Java A, Apicelli AJ, Liszewski MK, et al. The complement system in COVID-19: friend and foe? *JCI Insight* 2020;5:e140711.
29. Zuo Y, Yalavarthi S, Shi H, et al. Neutrophil extracellular traps in COVID-19. *JCI Insight* 2020;5:e138999.
30. Lee KH, Kronbichler A, Park DD-Y, et al. Neutrophil extracellular traps (NETs) in autoimmune diseases: a comprehensive review. *Autoimmun Rev* 2017;16:1160–73.
31. Camous L, Roumenina L, Bigot S, et al. Complement alternative pathway acts as a positive feedback amplification of neutrophil activation. *Blood* 2011;117:1340–9.
32. Yuen J, Pluthero FG, Doua DN, et al. NETosing neutrophils activate complement both on their own NETs and bacteria via alternative and non-alternative pathways. *Front Immunol* 2016;7:137.
33. Zuo Y, Yalavarthi S, Gockman K, et al. Anti-neutrophil extracellular trap antibodies and impaired neutrophil extracellular trap degradation in antiphospholipid syndrome. *Arthritis Rheumatol* 2020;72:2130–5.
34. Zuo Y, Kanthi Y, Knight JS, Kim AH. The interplay between neutrophils, complement, and microthrombi in COVID-19. *Best Pract Res Clin Rheumatol* 2021;35:101661.
35. Leffler J, Martin M, Gullstrand B, et al. Neutrophil extracellular traps that are not degraded in systemic lupus erythematosus activate complement exacerbating the disease. *J Immunol* 2012;188:3522–31.
36. Thurman JM, Yapa R. Complement therapeutics in autoimmune disease. *Front Immunol* 2019;10:672.
37. Knight JS, Kaplan MJ. Lupus neutrophils: ‘NET’ gain in understanding lupus pathogenesis. *Curr Opin Rheumatol* 2012;24:441–50.
38. Wright RD, Bannerman F, Beresford MW, Oni L. A systematic review of the role of eculizumab in systemic lupus erythematosus-associated thrombotic microangiopathy. *BMC Nephrol* 2020;21:1–8.
39. Pivovarova AI, Thongprayoon C, Hansrivijit P, et al. Thrombotic microangiopathy among hospitalized patients with systemic lupus erythematosus in the United States. *Diseases* 2020;9:3.
40. Kello N, El Khoury L, Marder G, Furie R, Zapantis E, Horowitz DL. Secondary thrombotic microangiopathy in systemic lupus erythematosus and antiphospholipid syndrome, the role of complement and use of eculizumab: case series and review of literature. *Seminars in Arthritis and Rheumatism* 2019:74–83.

41. Sharma M, Vignesh P, Tiewsoh K, Rawat A. Revisiting the complement system in systemic lupus erythematosus. *Expert Rev Clin Immunol* 2020;16:397–408.
42. Schejbel L, Skattum L, Hagelberg S, et al. Molecular basis of hereditary C1q deficiency—revisited: identification of several novel disease-causing mutations. *Genes Immun* 2011;12:626–34.
43. Yang Y, Chung EK, Wu YL, et al. Gene copy-number variation and associated polymorphisms of complement component C4 in human systemic lupus erythematosus (SLE): low copy number is a risk factor for and high copy number is a protective factor against SLE susceptibility in European Americans. *Am J Hum Genet* 2007;80:1037–54.
44. Chen M, Daha MR, Kallenberg CG. The complement system in systemic autoimmune disease. *J Autoimmun* 2010;34:276–86.
45. Foltyn Zadura A, Zipfel PF, Bokarewa MI, et al. Factor H autoantibodies and deletion of Complement Factor H-Related protein-1 in rheumatic diseases in comparison to atypical hemolytic uremic syndrome. *Arthritis research & therapy* 2012;14:1-11.
46. Weinstein A, Alexander RV, Zack DJ. A review of complement activation in SLE. *Current rheumatology reports* 2021;23:1-8.
47. Park MH, Caselman N, Ulmer S, Weitz IC. Complement-mediated thrombotic microangiopathy associated with lupus nephritis. *Blood Advances* 2018;2:2090-4.
48. Lintner KE, Wu YL, Yang Y, et al. Early components of the complement classical activation pathway in human systemic autoimmune diseases. *Front Immunol* 2016;7:36.
49. Casciola-Rosen LA, Anhalt G, Rosen A. Autoantigens targeted in systemic lupus erythematosus are clustered in two populations of surface structures on apoptotic keratinocytes. *J Exp Med* 1994;179:1317–30.
50. Navratil JS, Watkins SC, Wisnieski JJ, Ahearn JM. The globular heads of C1q specifically recognize surface blebs of apoptotic vascular endothelial cells. *J Immunol* 2001;166:3231–9.
51. Lovgren T, Eloranta ML, Kastner B, Wahren-Herlenius M, Alm GV, Ronnblom L. Induction of interferon-alpha by immune complexes or liposomes containing systemic lupus erythematosus autoantigen- and Sjogren's syndrome autoantigen-associated RNA. *Arthritis Rheum* 2006;54:1917–27.
52. Vallin H, Blomberg S, Alm GV, Cederblad B, Ronnblom L. Patients with systemic lupus erythematosus (SLE) have a circulating inducer of interferon-alpha (IFN-alpha) production acting on leucocytes resembling immature dendritic cells. *Clin Exp Immunol* 1999;115:196–202.
53. Lindau D, Mussard J, Rabsteyn A, et al. TLR9 independent interferon α production by neutrophils on NETosis in response to circulating chromatin, a key lupus autoantigen. *Ann Rheum Dis* 2014;73:2199–207.
54. Lande R, Ganguly D, Facchinetti V, et al. Neutrophils activate plasmacytoid dendritic cells by releasing self-DNA-peptide complexes in systemic lupus erythematosus. *Sci Transl Med* 2011;3:73ra19.
55. Villanueva E, Yalavarthi S, Berthier CC, et al. Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. *J Immunol* 2011;187:538–52.
56. Leffler J, Gullstrand B, Jonsen A, et al. Degradation of neutrophil extracellular traps co-varies with disease activity in patients with systemic lupus erythematosus. *Arthritis Res Ther* 2013;15:R84.

57. Banfi G, Bertani T, Boeri V, et al. Renal vascular lesions as a marker of poor prognosis in patients with lupus nephritis. Gruppo Italiano per lo Studio della Nefrite Lupica (GISNEL). *Am J Kidney Dis* 1991;18:240-8.
58. Song D, Wu L-h, Wang F-m, et al. The spectrum of renal thrombotic microangiopathy in lupus nephritis. *Arthritis Research & Therapy* 2013;15:R12.
59. Chaturvedi S, Braunstein EM, Brodsky RA. Antiphospholipid syndrome: complement activation, complement gene mutations, and therapeutic implications. *J Thromb Haemost* 2021;19:607–16.
60. Cohen D, Berger SP, Steup-Beekman GM, Bloemenkamp KW, Bajema IM. Diagnosis and management of the antiphospholipid syndrome. *BMJ* 2010;340:c2541.
61. Meng H, Yalavarthi S, Kanthi Y, et al. In vivo role of neutrophil extracellular traps in antiphospholipid antibody-mediated venous thrombosis. *Arthritis Rheumatol* 2017;69:655–67.
62. Tektonidou MG, Sotsiou F, Nakopoulou L, Vlachoyiannopoulos PG, Moutsopoulos HM. Antiphospholipid syndrome nephropathy in patients with systemic lupus erythematosus and antiphospholipid antibodies: prevalence, clinical associations, and long - term outcome. *Arthritis Rheum* 2004;50:2569 - 79.
63. Asherson R, Cervera R, De Groot P, et al. Catastrophic antiphospholipid syndrome: international consensus statement on classification criteria and treatment guidelines. *Lupus* 2003;12:530–4.
64. Cervera R, Serrano R, Pons-Estel G, et al. Morbidity and mortality in the antiphospholipid syndrome during a 10-year period: a multicentre prospective study of 1000 patients. *Ann Rheum Dis* 2015;74:1011–8.
65. Fischetti F, Durigutto P, Pellis V, et al. Thrombus formation induced by antibodies to β 2-glycoprotein I is complement dependent and requires a priming factor. *Blood* 2005;106:2340–6.
66. Oku K, Atsumi T, Bohgaki M, et al. Complement activation in patients with primary antiphospholipid syndrome. *Ann Rheum Dis* 2009;68:1030–5.
67. Breen KA, Seed P, Parmar K, Moore GW, Stuart-Smith SE, Hunt BJ. Complement activation in patients with isolated antiphospholipid antibodies or primary antiphospholipid syndrome. *Thromb Haemost* 2012;107:423–9.
68. Barratt-Due A, Fløisand Y, Orrem HL, et al. Complement activation is a crucial pathogenic factor in catastrophic antiphospholipid syndrome. *Rheumatology* 2016;55:1337–9.
69. Shapira I, Andrade D, Allen SL, Salmon JE. Brief report: induction of sustained remission in recurrent catastrophic antiphospholipid syndrome via inhibition of terminal complement with eculizumab. *Arthritis Rheum* 2012;64:2719–23.
70. Skoczynska M, Crowther MA, Chowaniec M, Ponikowska M, Chaturvedi S, Legault K. Thrombotic microangiopathy in the course of catastrophic antiphospholipid syndrome successfully treated with eculizumab: case report and systematic review of the literature. *Lupus* 2020;29:631–9.
71. López-Benjume B, Rodríguez-Pintó I, Amigo MC, et al. Eculizumab use in catastrophic antiphospholipid syndrome (CAPS): descriptive analysis from the “CAPS Registry”. *Autoimmun Rev* 2022;21:103055.
72. Devresse A, Aydin S, Le Quintrec M, et al. Complement activation and effect of eculizumab in scleroderma renal crisis. *Medicine* 2016;95:e4459.
73. Zuckerman R, Asif A, Costanzo EJ, Vachharajani T. Complement activation in atypical hemolytic uremic syndrome and scleroderma renal crisis: a critical analysis of pathophysiology. *Braz J Nephrol* 2018;40:77–81.

74. Ghossein C, Varga J, Fenves AZ. Recent developments in the classification, evaluation, pathophysiology, and management of scleroderma renal crisis. *Curr Rheumatol Rep* 2016;18:1–6.
75. Timmermans SA, Abdul-Hamid MA, Vanderlocht J, et al. Patients with hypertension-associated thrombotic microangiopathy may present with complement abnormalities. *Kidney International* 2017;91:1420-5.
76. Batal I, Domsic RT, Shafer A, et al. Renal biopsy findings predicting outcome in scleroderma renal crisis. *Hum Pathol* 2009;40:332–40.
77. Okrój M, Johansson M, Saxne T, Blom AM, Hesselstrand R. Analysis of complement biomarkers in systemic sclerosis indicates a distinct pattern in scleroderma renal crisis. *Arthritis Res Ther* 2016;18:1–10.
78. Zuckerman JE, Chang A. Complement and renal thrombotic microangiopathy associated with hypertension and scleroderma. *Adv Chronic Kidney Dis* 2020;27:149–54.
79. Maugeri N, Capobianco A, Rovere-Querini P, et al. Platelet microparticles sustain autophagy-associated activation of neutrophils in systemic sclerosis. *Sci Transl Med* 2018;10:3089.
80. Didier K, Giusti D, Le Jan S, et al. Neutrophil extracellular traps generation relates with early stage and vascular complications in systemic sclerosis. *J Clin Med* 2020;9:2136.
81. Gouin A, Ribes D, Colombat M, et al. Role of C5 inhibition in idiopathic inflammatory myopathies and scleroderma renal crisis-induced thrombotic microangiopathies. *Kidney Int Rep* 2021;6:1015–21.
82. Thomas CP, Nester CM, Phan AC, Sharma M, Steele AL, Lenert PS. Eculizumab for rescue of thrombotic microangiopathy in PM-Scl antibody-positive autoimmune overlap syndrome. *Clin Kidney J* 2015;8:698–701.
83. McFarlane PA, Bitzan M, Broome C, et al. Making the correct diagnosis in thrombotic microangiopathy: A narrative review. *Canadian Journal of Kidney Health and Disease* 2021;8:20543581211008707.
84. Nilsson B, Ekdahl KN. Complement diagnostics: concepts, indications, and practical guidelines. *Clin Dev Immunol* 2012;2012.
85. Palomo M, Blasco M, Molina P, et al. Complement activation and thrombotic microangiopathies. *Clinical Journal of the American Society of Nephrology* 2019;14:1719–32.
86. Thompson GL, Kavanagh D. Diagnosis and treatment of thrombotic microangiopathy. *International Journal of Laboratory Hematology* 2022;44:101-13.
87. United States Food and Drug Administration. Eculizumab – Highlights of Prescribing Information. [Internet. Accessed March 21, 2022]. Available from: https://www.accessdata.fda.gov/drugsatfda_docs/label/2020/125166s4341bl.pdf.
88. United States Food and Drug Administration. Ravulizumab – Highlights of Prescribing Information. [Internet. Accessed March 21, 2022]. Available from: https://www.accessdata.fda.gov/drugsatfda_docs/label/2022/761108s0201bl.pdf.
89. European Medicines Agency. Eculizumab – Summary of Product Characteristics. [Internet Accessed July 7, 2022] Available from: <https://www.medicines.org.uk/emc/medicine/19966/SPC/soliris/#gref>.
90. European Medicines Agency. Ravulizumab – Summary of Product Characteristics. [Internet Accessed July 7, 2022] Available from: <https://www.medicines.org.uk/emc/product/11945/smpc#gref>.

91. Rondeau E, Scully M, Ariceta G, et al. The long-acting C5 inhibitor, Ravulizumab, is effective and safe in adult patients with atypical hemolytic uremic syndrome naïve to complement inhibitor treatment. *Kidney Int* 2020;97:1287–96.
92. Radhakrishnan J. Anticomplement therapies in “secondary thrombotic microangiopathies”: ready for prime time? *Kidney Int* 2019;96:833–5.
93. Fakhouri F, Hourmant M, Campistol JM, et al. Terminal complement inhibitor eculizumab in adult patients with atypical hemolytic uremic syndrome: a single-arm, open-label trial. *Am J Kidney Dis* 2016;68:84–93.
94. Licht C, Greenbaum LA, Muus P, et al. Efficacy and safety of eculizumab in atypical hemolytic uremic syndrome from 2-year extensions of phase 2 studies. *Kidney Int* 2015;87:1061–73.
95. Legendre C, Licht C, Muus P, et al. Terminal complement inhibitor eculizumab in atypical hemolytic–uremic syndrome. *N Engl J Med* 2013;368:2169–81.
96. Greenbaum LA, Fila M, Ardissino G, et al. Eculizumab is a safe and effective treatment in pediatric patients with atypical hemolytic uremic syndrome. *Kidney Int* 2016;89:701–11.
97. Rondeau E, Cataland SR, Al-Dakkak I, Miller B, Webb NJ, Landau D. Eculizumab safety: five-year experience from the global atypical hemolytic uremic syndrome registry. *Kidney Int Rep* 2019;4:1568–76.
98. Barbour T, Scully M, Ariceta G, et al. Long-term efficacy and safety of the long-acting complement C5 inhibitor ravulizumab for the treatment of atypical hemolytic uremic syndrome in adults. *Kidney Int Rep* 2021;6:1603–13.
99. Ariceta G, Dixon BP, Kim SH, et al. The long-acting C5 inhibitor, ravulizumab, is effective and safe in pediatric patients with atypical hemolytic uremic syndrome naïve to complement inhibitor treatment. *Kidney Int* 2021;100:225–37.
100. Fakhouri F, Schwotzer N, Golshayan D, Frémeaux-Bacchi V. The rational use of complement inhibitors in kidney diseases. *Kidney International Reports* 2022.
101. Werion A, Rondeau E. C5 inhibition in secondary thrombotic microangiopathies: A yet unresolved question. *Kidney Int Rep* 2021;6:878.
102. Brocklebank V, Kavanagh D. Complement C5-inhibiting therapy for the thrombotic microangiopathies: accumulating evidence, but not a panacea. *Clin Kidney J* 2017;10:600–24.
103. Haydock L, Garneau AP, Tremblay L, et al. Genetic abnormalities in biopsy-proven, adult-onset hemolytic uremic syndrome and C3 glomerulopathy. *Journal of Molecular Medicine* 2022;100:269-84.

FIGURE LEGENDS

Figure 1. Activation of the complement system.

This adapted figure was originally published in Lemaire, Mathieu, et al. Inherited kidney complement diseases. *Clinical Journal of the American Society of Nephrology* 16.6 (2021): 942–956. C1 complex consists of C1q, C1r, and C1s. FB: factor B; FD: factor D; MAC: membrane attack complex; MBL: mannose-binding lectin.

Figure 2. Etiopathogenesis of aHUS.

Downloaded on April 19, 2024 from www.jrheum.org

This figure was published in *Advances in Chronic Kidney Disease*, Volume 27 (Issue 2), Java, A., Peri- and post-operative evaluation and management of atypical hemolytic uremic syndrome (aHUS) in kidney transplantation, pg. 128–137, Copyright Elsevier, 2020. The relationship between aHUS and CR1 is hypothetical owing to CR1 being a well-known regulator of CP and AP.¹⁰³ aHUS: atypical hemolytic uremic syndrome; AP: alternative pathway; CFHR: complement factor H-related; CP: classical pathway; CR1: complement receptor 1; FB, factor B; FD, factor D; FH: factor H; FI: factor I; MCP: membrane cofactor protein; P, properdin.

Figure 3. Mechanism of the interplay between complement activation and immune hyperinflammatory reactions.

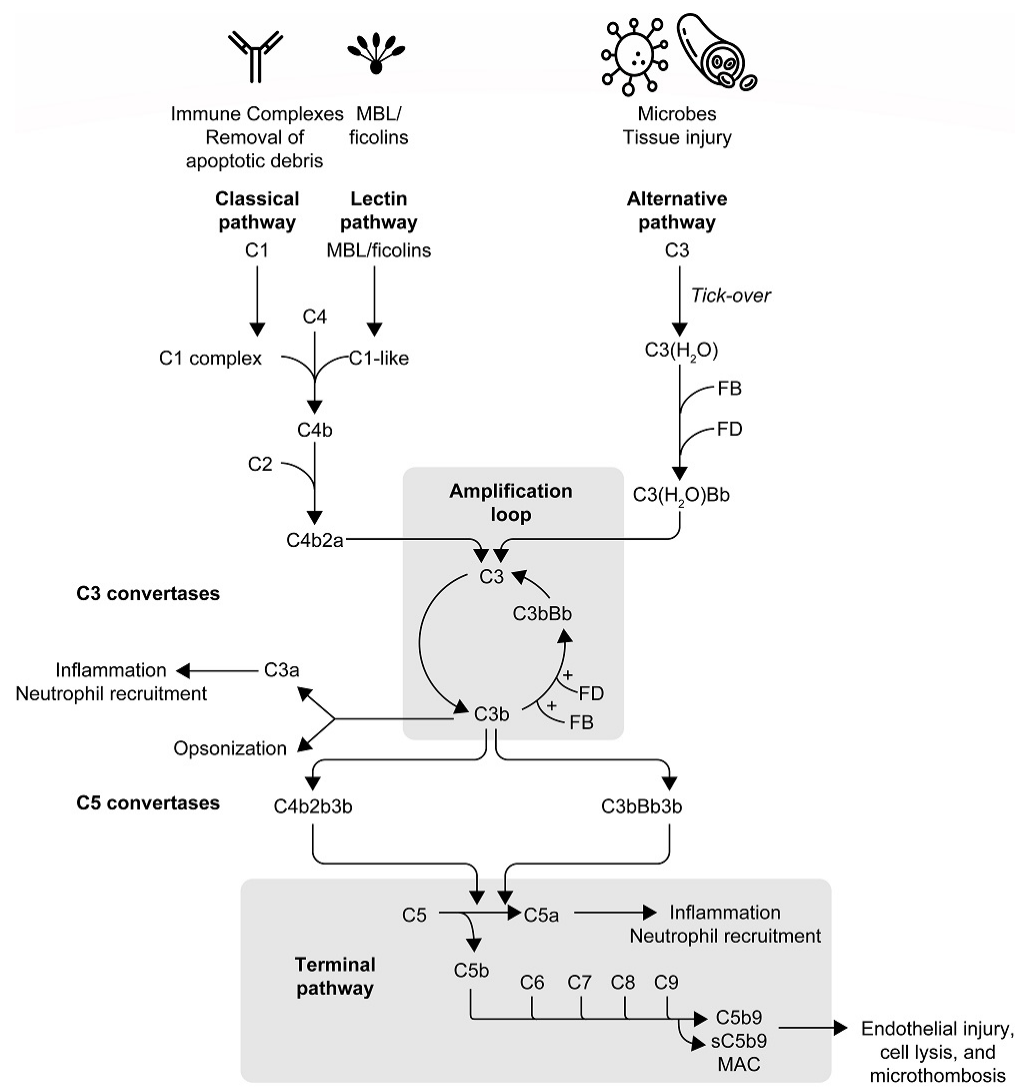
Sourced from: *JCI Insight*, Volume 5 (Issue 15), Java, A. et al., The complement system in COVID-19: friend and foe?, e140711, 2020. Complement activation generates the anaphylatoxins C3a and C5a and leads to neutrophil recruitment. Activated neutrophils generate NET lattices in a type of cell death known as NETosis, which contain components that activate the alternative pathway and generate an inflammatory feedback loop. Additionally, the MAC causes endothelial damage, with injured tissue releasing inflammatory cytokines that further stimulate NETosis. Injury to the endothelial tissues results in the creation of prothrombotic factors, compounded by C5a-mediated release of proteins that promote a hypercoagulable state. AP, alternative pathway; MAC: membrane attack complex; NET: neutrophil extracellular traps; TMA: thrombotic microangiopathy.

Figure 4. Diagnostic work-up

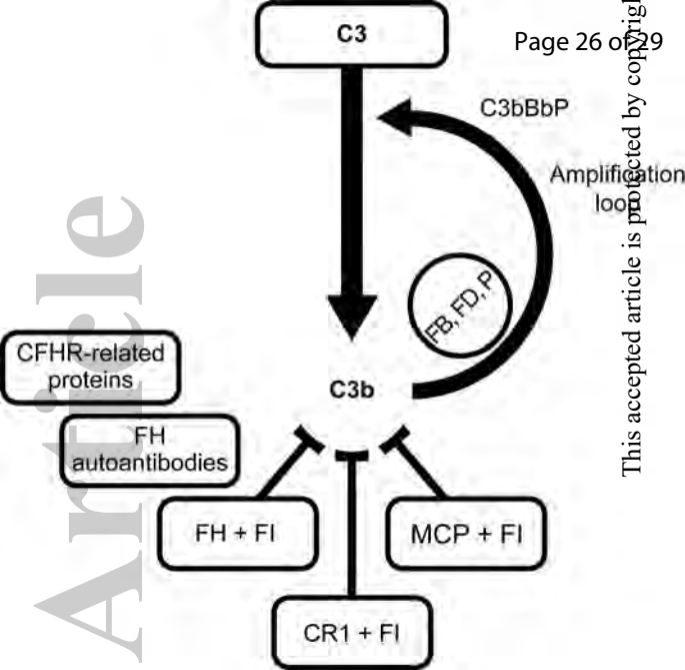
ADAMTS13: a disintegrin and metalloproteinase with a thrombospondin type 1 motif, member 13; LDH: lactate dehydrogenase; TMA: thrombotic microangiopathy; TTP: thrombotic thrombocytopenic purpura; VUS: variant of uncertain significance.

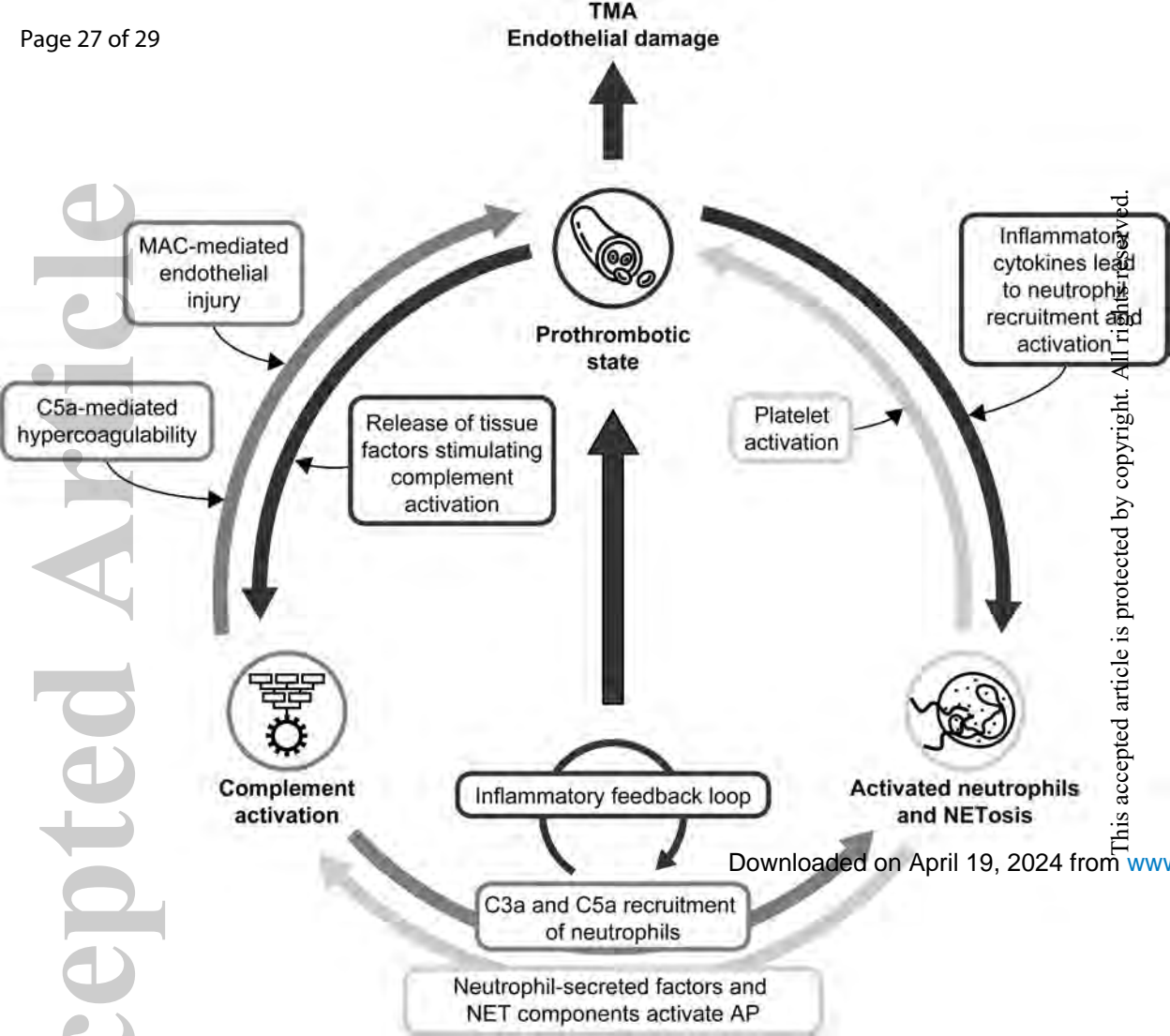
Figure 5. Effects of autoimmune diseases SLE, APS, and SRC, and action of the C5 inhibitors eculizumab and ravulizumab on the complement system.

This adapted figure was originally published in Lemaire, Mathieu, et al. Inherited kidney complement diseases. *Clinical Journal of the American Society of Nephrology* 16.6 (2021): 942–956. Black bar indicates the point of action of C5 inhibitors, which specifically inhibit C5 cleavage to C5a and C5b, preventing C5a-mediated inflammation and the formation of the MAC. APS: antiphospholipid syndrome; FB: factor B; FD: factor D; FH: factor H; MAC: membrane attack complex; MBL: mannose-binding lectins; P, properdin; SLE: systemic lupus erythematosus; SRC: scleroderma renal crisis.



264x290mm (96 x 96 DPI)





Recognize TMA in the setting of autoimmune diseases

ADAMTS13 activity < 10%;
plasma exchange for TTP

Treat the underlying
autoimmune disease

Persistent TMA

Complement studies

Whole plasma/serum testing

- Components (C3, C4)
- Regulators (factor H, factor I, membrane cofactor protein by flow cytometry)
- Autoantibodies (factor H)
- Functional assays (CH₅₀, AH₅₀)
- Split products (C3c, C3d, Ba, Bb, C5b-9)

Genetic testing for complement variants

Variant identified

Benign

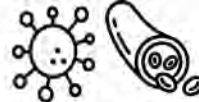
VUS

Pathogenic

Recombinant production of the variant or purified from serum

Combination of antigenic, genetic, and structure–function data to define the significance of the variant and determine the risk of recurrence in the allograft

- Page 28 of 29
- Anemia
 - Thrombocytopenia
 - Elevated LDH
 - Low haptoglobin
 - Coombs test
 - Coagulation profile
 - Schistocytes
 - Absence of improvement or worsening of kidney function
 - Kidney biopsy shows TMA

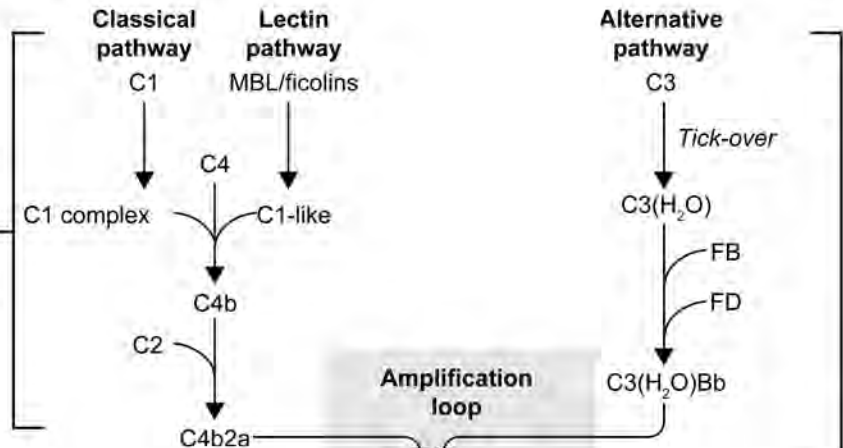


Immune Complexes
Removal of
apoptotic debris

MBL/
ficolins

Microbes
Tissue injury

SLE
Deficiencies or low copy number variations of early classical pathway proteins. Autoantibodies may also be generated against C1, C5, and FH.



SRC
Renal deposition of C1q, C4d, and C3b observed, suggesting classical pathway involvement. Reduced levels of C3 and FH also noted, suggesting alternative pathway activation. Renal involvement associated with reduced C4d and reduced C3bBbP and MAC, reflecting complement consumption.

APS
Lower plasma C3 and C4 levels, and increased Bb, C3a, and serum MAC. C9 deposition on vascular endothelium also observed, indicating presence of MAC.

C3 convertases

Inflammation ← C3a
Neutrophil recruitment

Opsonization

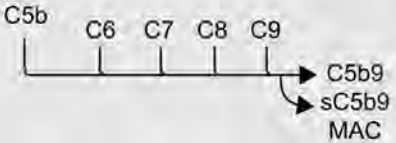
C5 convertases

C4b2b3b

C3bBb3b

C5 → C5a (C5 inhibition) + C5b

Terminal pathway



Accepted Article

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