

# Biomarkers of Bone Metabolism in Ankylosing Spondylitis in Relation to Osteoproliferation and Osteoporosis

Eva Klingberg, Merja Nurkkala, Hans Carlsten, and Helena Forsblad-d'Elia

**ABSTRACT.** *Objective.* To identify biomarkers for bone metabolism in patients with ankylosing spondylitis (AS) and to determine the relationship between these biomarkers and disease activity, back mobility, osteoproliferation, and bone mineral density (BMD).

*Methods.* Serum levels of Wingless protein (Wnt-3a), Dickkopf-1 (DKK-1), sclerostin, soluble receptor activator of nuclear factor- $\kappa$ B ligand (sRANKL), and osteoprotegerin were assessed using ELISA. Ankylosing Spondylitis Disease Activity Score-C reactive protein, Bath Ankylosing Spondylitis Disease Activity Index, Bath Ankylosing Spondylitis patient global score, and C-reactive protein (CRP) were used as disease activity measures, and Bath Ankylosing Spondylitis Metrology Index (BASMI) as a measure of spinal mobility. Lateral spine radiographs were scored for chronic AS-related changes (mSASSS). BMD was measured with dual-energy x-ray absorptiometry.

*Results.* Two hundred four patients with AS (NY criteria; 57% men), with a mean age of  $50 \pm 13$  years and disease duration  $15 \pm 11$  years, and 80 age and sex-matched controls were included. The patients with AS had significantly higher serum levels of Wnt-3a ( $p < 0.001$ ) and lower levels of sclerostin ( $p = 0.014$ ) and sRANKL ( $p = 0.047$ ) compared with the controls. High CRP was associated with low sclerostin ( $r_s = -0.21$ ,  $p = 0.003$ ) and DKK-1 ( $r_s = -0.14$ ,  $p = 0.045$ ). In multiple linear regression analyses, increasing BASMI and mSASSS were independently associated with older age, male sex, high CRP, and elevated serum levels of Wnt-3a. In addition, mSASSS remained associated with a high number of smoking pack-years after adjusting for age. Low BMD of femoral neck was associated with high mSASSS after adjusting for age.

*Conclusion.* Serum levels of Wnt-3a are elevated in AS and associated with increased BASMI and mSASSS, independent of age, indicating that Wnt-3a could be a biomarker for the osteoproliferative process. (First Release June 15 2014; J Rheumatol 2014;41:1349–56; doi:10.3899/jrheum.131199)

## Key Indexing Terms:

ANKYLOSING SPONDYLITIS  
SCLEROSTIN

BIOMARKERS  
DICKKOPF-1

WNT  
OSTEOPOROSIS

Ankylosing spondylitis (AS) is a chronic rheumatic disease characterized by sacroiliac and spinal inflammation and the growth of bony spurs (syndesmophytes) between the vertebrae. Syndesmophyte formation is associated with restriction of spinal mobility. Pathologic new bone is mainly formed by endochondral ossification, but membranous ossification also contributes<sup>1,2</sup>. Inflammation and osteoproliferation are at least partially uncoupled events in AS, because osteoproliferation can occur even if the inflam-

matory activity is low. Although inflammation can be treated successfully with blockers of tumor necrosis factor (TNF- $\alpha$ ), several 2-year followup studies have shown a lack of effect on syndesmophyte formation<sup>3,4,5</sup>. In one observational study, however, patients with AS taking TNF inhibitors were followed for up to 5 years, and TNF inhibition appeared to reduce radiographic progression<sup>6</sup>. Daily or high use of nonsteroidal antiinflammatory drugs (NSAID) has been associated with less radiographic progression, but a truly effective treatment against new bone formation in AS has yet to be found<sup>7,8,9</sup>.

Parallel to the osteoproliferation, patients with AS also have an increased loss of trabecular bone, resulting in an elevated risk for vertebral fractures, sometimes complicated by neurological injuries<sup>10,11</sup>. Prevalences of osteoporosis of 19% to 62% and vertebral fractures of 9% to 42% have been reported from different AS cohorts<sup>10,12,13,14</sup>. These 2 enhanced but opposite bone processes create diagnostic and therapeutic problems. Osteoproliferative changes cause an artifactual increase in lumbar bone mineral density (BMD). Levels of bone biomarkers can be difficult to interpret if they mirror both processes. Treating and preventing osteoproliferation could potentially worsen osteoporosis and vice versa, and the knowledge is missing about which time

From the Department of Rheumatology and Inflammation Research, Sahlgrenska Academy, University of Gothenburg, Sweden.

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E. Klingberg, MD, PhD; M. Nurkkala, PhD; H. Carlsten, MD, PhD, Professor; H. Forsblad-d'Elia, MD, PhD, Associate professor, Department of Rheumatology and Inflammation Research, Sahlgrenska Academy at the University of Gothenburg.

Address correspondence to Dr. E. Klingberg, Department of Rheumatology at Sahlgrenska/SU, Gröna stråket 14; S-413 45 Gothenburg, Sweden. E-mail: Eva.Klingberg@vgregion.se

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window to use to treat these 2 conditions. There is a need for biomarkers for both the osteoproliferation and the osteoporosis to guide physicians in the decision of who to treat, when to treat, and how to follow up.

The bone morphogenic proteins, hedgehogs, and wingless proteins (Wnt) are important regulators of bone formation by intracellular signaling pathways that affect gene transcription. Wnt can signal by at least 4 different pathways, of which the “canonical” Wnt/ $\beta$ -catenin pathway is best described. Wnt/ $\beta$ -catenin signaling stimulates osteoblast differentiation, proliferation, and survival resulting in increased bone mass, whereas suppressed signaling leads to bone loss<sup>15,16,17</sup>. Wnt signaling also triggers the formation of osteophytes<sup>18</sup>. The Wnt signaling is regulated by several secreted receptor antagonists. Dickkopf-1 (DKK-1), expressed mainly by osteoblasts and osteocytes, and sclerostin, expressed exclusively by osteocytes, are negative regulators of bone formation by binding to Wnt coreceptors<sup>19,20</sup>. The expression of sclerostin in the osteocytes is reduced by mechanical loading<sup>21,22</sup>. Insufficient or excess activity of DKK-1 or sclerostin in bone results in increased or decreased bone density, respectively<sup>16,23</sup>.

Osteoclast maturation requires macrophage colony stimulating factor (M-CSF) and receptor activator of nuclear factor- $\kappa$ B ligand (RANKL)<sup>24,25</sup>. RANKL binds to RANK on osteoclast precursors, inducing their transformation into mature osteoclasts and thus has a resorptive effect on bone. The RANKL expression is enhanced by proinflammatory cytokines. The RANK-RANKL interaction is inhibited by osteoprotegerin (OPG), a decoy receptor for RANKL that has an antiresorptive effect on bone.

The goal of our study was to identify biomarkers of importance for both osteoporosis and osteoproliferation in AS. The specific aims of the study were to compare the serum concentrations of a panel of biomarkers reflecting bone metabolism in patients with AS versus in healthy controls and to analyze the relationship between these biomarkers and disease activity, back mobility, osteoproliferation, and BMD. The biomarkers studied were Wnt-3a, DKK-1, sclerostin, soluble RANKL (sRANKL), and OPG.

## MATERIALS AND METHODS

**Patients.** All patients with AS (modified New York criteria) who were registered at the departments of rheumatology at the Sahlgrenska University Hospital and Hospitals of Alingsås and Borås were invited to participate. Exclusion criteria were psoriasis, inflammatory bowel disease, dementia, other concomitant rheumatic disease, and difficulties in understanding Swedish. The inclusion of the patients has been described<sup>10,26</sup>. The patients all gave written informed consent according to the Declaration of Helsinki and the study was approved by the Regional Ethics committee in Gothenburg.

The patients answered questionnaires regarding medical history and medication including use of NSAID (daily or on-demand) and TNF- $\alpha$  blockers. Disease activity was assessed using the Bath Ankylosing Spondylitis Disease Activity Index (BASDAI), the Ankylosing Spondylitis Disease Activity Score with C-reactive protein (ASDAS-CRP), and the

Bath Ankylosing Spondylitis patient global score (BAS-G). Spinal mobility was measured for calculation of the Bath Ankylosing Spondylitis Metrology Index (BASMI)<sup>27,28</sup>.

The patients were asked about current and previous smoking, age at onset and disruption of smoking, and average daily cigarette consumption. Smoking pack-years were subsequently calculated.

**Healthy controls.** Blood samples from healthy sex and age-matched blood donors were collected to use as controls to the AS patients. The blood donors were recruited while giving blood at Sahlgrenska University Hospital. The blood donors all gave their written informed consent and answered questionnaires stating that they were in full health.

**Blood samples.** Blood samples were drawn in a non-fasting state. Levels of hemoglobin, erythrocyte sedimentation rate (ESR), and CRP were analyzed using standard laboratory techniques at the hospitals. Serum was stored at  $-20^{\circ}\text{C}$  until needed for analysis.

**Biomarkers of bone metabolism.** Serum samples were assayed for markers of bone metabolism using specific ELISA kits according to manufacturers' instructions. The biomarkers measured were Wnt-3a (Usen Life Science Inc.), total DKK-1 (R&D Systems), sclerostin (Biomedica Gruppe), sRANKL (Biovendor), and OPG (Immunodiagnostic Systems). The following values of sensitivity were reported by the manufacturers: Wnt-3a lower limits of detection (LLD) = 0.055 ng/ml; DKK-1 LLD = 4.2 pg/ml; sclerostin LLD = 2.6 pmol/l; sRANKL LLD = 0.4 pmol/l; OPG LLD = 14 pmol/l.

Absorbance was read at 450 nm in SpectraMax 340PC384 spectrophotometer (Molecular Devices). The software SoftMax Pro 5.2 was used to calculate the biomarker concentrations.

**Radiography and dual-energy x-ray absorptiometry (DEXA).** Lateral radiographs of the cervical and lumbar spine were scored for chronic AS-related changes using the modified Stoke Ankylosing Spondylitis Spine Score (mSASSS)<sup>29</sup>. The mSASSS scoring was done by an experienced radiologist, specialized in rheumatology and AS in particular. BMD was measured with DEXA (Hologic Discovery A, Hologic Inc.) in the femoral neck.

**Statistical analyses.** Statistical analyses were performed using PASW Statistics 18.0 (SPSS Inc.). Descriptive statistics are presented as median and range and/or mean and SD. Mann-Whitney U test or chi-square tests were used for comparisons between groups (patients/controls, men/women, ever-smokers/never-smokers, anti-TNF treated/non-treated). All correlations were calculated using Spearman's correlation ( $r_s$ ). All tests were 2-tailed and  $p < 0.05$  was considered statistically significant. Multiple linear regressions were run using a stepwise method with BASMI, mSASSS, and BMD of femoral neck as outcome. The linear regressions with mSASSS as outcome was run in 2 models. In the first model, conventional mSASSS score was used. In the second model, mSASSS was log-transformed and patients with only sacroiliitis (mSASSS = 0) were excluded, leaving data from 148 patients ready to analyze. This was done to make mSASSS normally distributed. Covariates in all multiple linear regressions were age, sex, smoking pack-years, CRP, and the biomarkers of bone metabolism, which were significantly associated with the outcome in the univariate analyses. In the model where BMD of femoral neck was the outcome, mSASSS was also included as a covariate. Age, sex, smoking, and CRP were chosen as covariates because they are known risk factors for osteoproliferation and osteoporosis.

## RESULTS

**Patients and healthy controls.** In total, 204 patients with AS (87 women and 117 men) with a median age of 49 years (range 17–78), symptom duration of 24 years (2–55), BASDAI 3.5 (0–9.6), and ASDAS-CRP 2.3 (0.8–5.9) were included (Table 1).

The healthy control group consisted of 80 blood donors

Table 1. Characteristics of 204 patients with ankylosing spondylitis in western Sweden.

Characteristics	No. Patients (%) or Median (range)
Sex	
Women	87 (43)
Men	117 (57)
Age, yrs	49 (17, 78)
Current smokers	24 (12)
Ever smokers (current + prior smokers during $\geq 6$ mos)	93 (46)
Years since symptom onset	24 (2, 55)
Years since diagnosis	12 (1, 47)
Present or past anterior uveitis	102 (50)
Present or past peripheral arthritis	120 (59)
Present or past coxitis	17 (8)
BASMI, score	3.0 (0.6, 7.4)
BASDAI, score	3.5 (0, 9.6)
BAS-G, score	2.9 (0, 10)
ASDAS-CRP, score	2.3 (0.8, 5.9)
mSASSS, score	5.5 (0, 72)
ESR, mm/h	11 (2, 105)
CRP, mg/l	5 (3, 80)
Hemoglobin, g/l	139 (105, 166)
HLA-B27 positive	178 (87)
Patients taking NSAID	158 (77)
Patients taking DMARD	62 (30)
Patients taking TNF- $\alpha$ blocker	42 (21)
Patients taking bisphosphonate	8 (4)
Patients taking glucocorticoid	7 (3)

ASDAS-CRP: Ankylosing Spondylitis Disease Activity Score using CRP; BASDAI: Bath Ankylosing Spondylitis Disease Activity Index; BAS-G: Bath Ankylosing Spondylitis global patient score; BASMI: Bath Ankylosing Spondylitis Metrology Index; CRP: C-reactive protein; DMARD: disease-modifying antirheumatic drug; ESR: erythrocyte sedimentation rate; mSASSS: modified Stoke Ankylosing Spondylitis Spine Score; NSAID: nonsteroidal antiinflammatory drug; TNF: tumor necrosis factor.

(26 women and 54 men) with a median age of 48 years (19–71). The age- and sex-distributions were not significantly different between the patients and the controls (Table 2).

*Levels of biomarkers in AS compared with healthy controls.*

The patients with AS had significantly higher serum levels of Wnt-3a ( $p < 0.001$ ), lower serum levels of sclerostin ( $p = 0.014$ ), lower serum levels of sRANKL ( $p = 0.047$ ), and lower sRANKL/OPG ratio ( $p = 0.022$ ) compared with the healthy controls (Table 2 and Figure 1). The serum levels of DKK-1 showed a tendency to be higher in the patients with AS ( $p = 0.058$ ).

*Correlation between the different biomarkers in the patients with AS.* Sclerostin was positively correlated with DKK-1 and OPG. No significant correlation was found between Wnt-3a and the other biomarkers (Table 3).

*Age and biomarkers.* Age was positively correlated with sclerostin and OPG, but negatively correlated with sRANKL in the patients with AS (Table 4).

*Disease activity and back mobility and biomarkers.* CRP was negatively correlated with both sclerostin and DKK-1. BASMI was positively correlated with Wnt-3a, sclerostin, and OPG (Table 4). No significant correlations were found between the biomarkers analyzed and BASDAI, ASDAS-CRP, BAS-G, ESR, or hemoglobin.

In multiple linear regression, increasing BASMI was independently associated with older age, male sex, high CRP, and elevated serum levels of Wnt-3a (Table 5).

*Chronic AS-related changes in the spine, smoking, and biomarkers.* mSASSS was significantly higher in the male patients compared with the female ( $20 \pm 22$  vs  $6 \pm 9$ ;  $p < 0.001$ ). mSASSS was positively correlated with Wnt-3a, sclerostin, CRP, age, and smoking pack-years (Table 4). Comparing the first (mSASSS = 0) and fourth (mSASSS  $\geq 20$ ) quartiles of mSASSS we found significantly higher Wnt-3a ( $3.98 \pm 1.00$  vs  $3.49 \pm 1.02$  ng/ml,  $p = 0.005$ ) and sclerostin ( $40.90 \pm 23.07$  vs  $31.4 \pm 17.65$  pmol/l,  $p = 0.01$ ), but lower DKK-1 ( $2.78 \pm 0.94$  vs  $3.27 \pm 1.58$  ng/ml,  $p = 0.003$ ) in the fourth quartile.

The smoking habits of the patients are displayed in Table 1. Smoking pack-years were positively correlated with mSASSS, BASMI, and age (Table 4). In comparison with never-smokers, the patients who smoked for at least 6

Table 2. Demographics and serum levels of biomarkers in patients with ankylosing spondylitis (AS) and in the controls.

	Patients with AS, n = 204, Median (range)	Controls, n = 80, Median (range)	p
Sex, female/male	87/117	26/54	0.100
Age, yrs	49 (17, 78)	48 (19, 71)	0.202
Wnt-3a, ng/ml	3.66 (0.99, 7.98)	2.78 (1.50, 5.10)	$< 0.001$
DKK-1, ng/ml	2.89 (1.14, 7.11)	2.66 (0.85, 6.10)	0.058
Sclerostin, pmol/l	31.88 (0, 173.48)	36.96 (0, 85.52)	0.014
sRANKL, pmol/l	165.23 (14.96, 2875.39)	210.80 (21.49, 1671.53)	0.047
OPG, pmol/l	3.62 (1.62, 7.63)	3.46 (0, 6.32)	0.264
sRANKL/OPG, ratio	45.55 (4.44, 1020.00)	68.49 (4.50, 671.01)	0.022

DKK-1: Dickkopf-1; OPG: osteoprotegerin; sRANKL: soluble receptor activator of nuclear factor- $\kappa$ B ligand; Wnt: wingless protein.

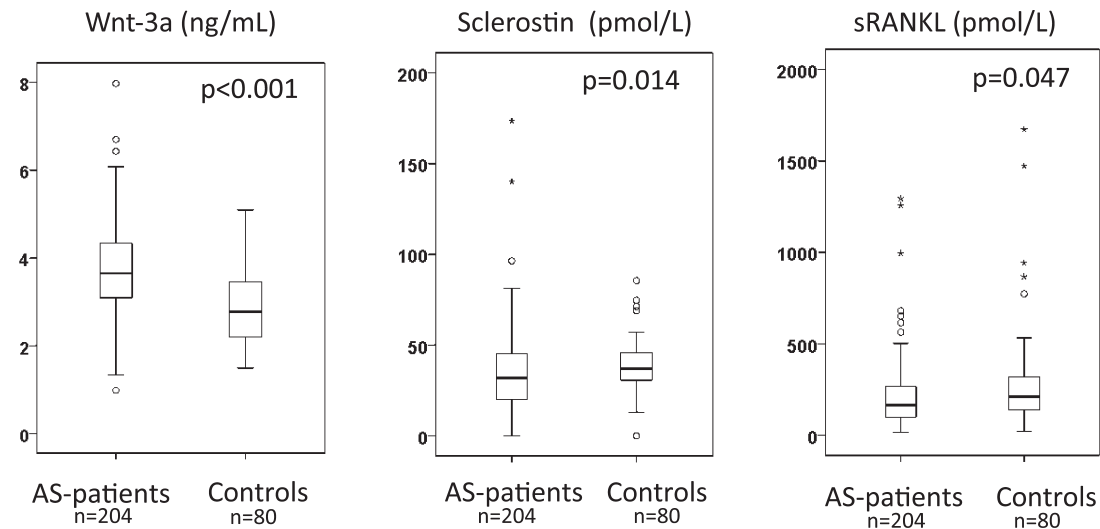


Figure 1. Box plot showing serum levels of Wnt-3a, sclerostin, and sRANKL in the patients with AS in comparison with the healthy controls. Values are the medians (horizontal line), interquartile ranges (box), and ranges of values (whiskers). O (outliers) shows cases with values between 1.5-3.0 box lengths and stars (extremes) values more than 3 box lengths from the upper or lower edge of the box. Wnt: wingless proteins; AS: ankylosing spondylitis; sRANKL: soluble receptor activator of nuclear factor- $\kappa$ B ligand.

Table 3. Univariate analyses. Correlation matrix showing significant correlations (Spearman's rho) between the biomarkers.

	Wnt-3a	DKK-1	Sclerostin	sRANKL	OPG
Wnt-3a		$r_s = -0.044$ $p = 0.533$	$r_s = -0.054$ $p = 0.455$	$r_s = 0.062$ $p = 0.382$	$r_s = -0.103$ $p = 0.142$
DKK-1	$r_s = -0.044$ $p = 0.533$		$r_s = 0.137$ $p = 0.050$	$r_s = -0.025$ $p = 0.720$	$r_s = -0.011$ $p = 0.878$
Sclerostin	$r_s = -0.054$ $p = 0.455$	$r_s = 0.137$ $p = 0.050$		$r_s = -0.029$ $p = 0.681$	$r_s = 0.354$ $p < 0.001$
sRANKL	$r_s = 0.062$ $p = 0.382$	$r_s = -0.025$ $p = 0.720$	$r_s = -0.029$ $p = 0.681$		$r_s = -0.220$ $p = 0.002$
OPG	$r_s = -0.103$ $p = 0.142$	$r_s = -0.011$ $p = 0.878$	$r_s = 0.354$ $p < 0.001$	$r_s = -0.220$ $p = 0.002$	

DKK-1: Dickkopf-1; OPG: osteoprotegerin; sRANKL: soluble receptor activator of nuclear factor- $\kappa$ B ligand; Wnt: wingless protein;  $r_s$ : Spearman's correlation coefficient.

Table 4. Univariate analyses. Significant correlations (Spearman's rho) between the biomarkers of bone metabolism and measures of disease activity, back mobility, chronic ankylosing spondylitis (AS)-related changes in the spine and bone mineral density (BMD). Data are  $r_s$ : Spearman's correlation coefficient, and p value.

	Age, yrs	Sex	Smoking, Pack-years	CRP, mg/l	BASMI Score	mSASSS Score	Femoral Neck BMD, g/cm <sup>2</sup>
Age, yrs		$-0.057$ , $p = 0.421$	$0.265$ , $p < 0.001^*$	$-0.018$ , $p = 0.794$	$0.594$ , $p < 0.001^*$	$0.389$ , $p < 0.001^*$	$-0.322$ , $p < 0.001^*$
Sex	$-0.057$ , $p = 0.421$		$0.096$ , $p = 0.173$	$0.075$ , $p = 0.284$	$0.053$ , $p = 0.452$	$0.377$ , $p < 0.001^*$	$0.228$ , $p = 0.001^*$
Smoking, pack-years	$0.265$ , $p < 0.001^*$	$0.096$ , $p = 0.173$		$0.095$ , $p = 0.178$	$0.304$ , $p < 0.001^*$	$0.300$ , $p < 0.001^*$	$-0.162$ , $p = 0.021^*$
CRP, mg/l	$-0.018$ , $p = 0.794$	$0.075$ , $p = 0.284$	$0.095$ , $p = 0.178$		$0.214$ , $p = 0.002^*$	$0.241$ , $p = 0.001^*$	$-0.072$ , $p = 0.307$
Wnt-3a, ng/ml	$0.042$ , $p = 0.549$	$-0.081$ , $p = 0.248$	$0.080$ , $p = 0.257$	$0.050$ , $p = 0.481$	$0.219$ , $p = 0.002^*$	$0.196$ , $p = 0.005^*$	$-0.160$ , $p = 0.023^*$
DKK-1, ng/ml	$-0.005$ , $p = 0.941$	$0.032$ , $p = 0.646$	$-0.045$ , $p = 0.525$	$-0.140$ , $p = 0.045^*$	$-0.080$ , $p = 0.256$	$-0.121$ , $p = 0.086$	$0.015$ , $p = 0.829$
Sclerostin, pmol/l	$0.496$ , $p < 0.001^*$	$0.121$ , $p = 0.084$	$0.099$ , $p = 0.159$	$-0.208$ , $p = 0.003^*$	$0.244$ , $p < 0.001^*$	$0.185$ , $p = 0.008^*$	$0.065$ , $p = 0.356$
sRANKL, pmol/l	$-0.242$ , $p = 0.001^*$	$-0.012$ , $p = 0.867$	$-0.025$ , $p = 0.720$	$0.096$ , $p = 0.174$	$-0.082$ , $p = 0.242$	$-0.044$ , $p = 0.532$	$0.152$ , $p = 0.031^*$
OPG, pmol/l	$0.421$ , $p < 0.001^*$	$-0.141$ , $p = 0.044^*$	$0.028$ , $p = 0.691$	$-0.008$ , $p = 0.907$	$0.201$ , $p = 0.004^*$	$0.072$ , $p = 0.307$	$-0.151$ , $p = 0.031^*$

\*Statistically significant if  $p < 0.05$ . Coding for sex: 1 = woman, 2 = man. BASMI: Bath Ankylosing Spondylitis Metrology Index; CRP: C-reactive protein; DKK-1: Dickkopf-1; OPG: osteoprotegerin; sRANKL: soluble receptor activator of nuclear factor- $\kappa$ B ligand; mSASSS: modified Stoke Ankylosing Spondylitis Spine Score; vBMD: volumetric bone mineral density; Wnt: wingless protein.



Table 5. Multivariate analyses. The results of the stepwise multiple linear regressions with BASMI, mSASSS, Log10 mSASSS, and femoral neck BMD as outcome.

	BASMI Score as Outcome, R <sup>2</sup> = 0.405		mSASSS Score as Outcome, R <sup>2</sup> = 0.328		Log10 mSASSS Score as Outcome, R <sup>2</sup> = 0.380		Femoral Neck BMD, g/cm <sup>2</sup> , as outcome, R <sup>2</sup> = 0.178	
	B	p	B	p	B	p	B	p
Age, yrs	0.068	< 0.001	0.539	< 0.001	0.022	< 0.001	-0.003	0.001
Sex	0.440	0.012	14.509	< 0.001	0.306	< 0.001	0.070	< 0.001
Smoking, pack-years	n.s.	n.s.	0.253	0.033	n.s.	n.s.	n.s.	n.s.
CRP, mg/l	0.023	0.006	n.s.	n.s.	0.006	0.035	n.s.	n.s.
mSASSS score							-0.001	0.013
Wnt-3a, ng/ml	0.305	< 0.001	2.532	0.027	0.085	0.019	n.s.	n.s.
DKK-1, ng/ml								
Sclerostin, pmol/l	n.s.	n.s.	n.s.	n.s.	n.s.	n.s.		
sRANKL, pmol/l							n.s.	n.s.
OPG, pmol/l	n.s.	n.s.					n.s.	n.s.

Covariates in all of the models of multiple linear regression were age, sex, smoking pack-years, CRP, and the biomarkers significantly associated with the outcome in the univariate analyses. Beta values are unstandardized regression coefficients. Empty boxes = the variable was not included in the linear regression model. Coding for sex: 1 = woman, 2 = man. n.s.: the variable was included in analysis, but not significantly associated with the outcome in the multiple linear regression; BASMI: Bath Ankylosing Spondylitis Metrology Index; BMD: bone mineral density; CRP: C-reactive protein; DKK-1: Dickkopf-1; OPG: osteoprotegerin; sRANKL: soluble receptor activator of nuclear factor- $\kappa$ B ligand; mSASSS: modified Stoke Ankylosing Spondylitis Spine Score; vBMD: volumetric BMD; Wnt: wingless protein.

months of their lives (ever-smokers) had significantly higher mSASSS (mean  $20 \pm 22$  vs  $9 \pm 15$ ;  $p < 0.001$ ), BASMI (mean  $3.6 \pm 1.6$  vs  $2.6 \pm 1.4$ ;  $p < 0.001$ ), and age (mean  $53 \pm 12$  vs  $47 \pm 13$ ;  $p = 0.001$ ). The distributions of sex, CRP, and the biomarkers of bone metabolism were equal between current smokers, ever-smokers, and never-smokers.

In multiple linear regression, high mSASSS was independently associated with age, male sex, smoking pack-years, and elevated levels of serum Wnt-3a. High log10 mSASSS was associated with age, male sex, high CRP, and elevated levels of serum Wnt-3a (Table 5).

**BMD and biomarkers.** In total, 5% of the patients had osteoporosis in the femoral neck and 10% osteoporosis in the lumbar spine. BMD results for this AS cohort are presented in more detail in a previous publication<sup>26</sup>. BMD of the femoral neck is not affected by the lumbar osteoproliferation in AS and was thus selected as the BMD measure to be compared with the levels of the biomarkers. BMD of the femoral neck was positively correlated with sRANKL, but negatively correlated with OPG and Wnt-3a (Table 4).

In multiple linear regression, low BMD of the femoral neck was independently associated with older age, female sex, and high mSASSS (Table 5).

**Medication and biomarkers.** A total of 20.6% (42/204) of the patients were treated with TNF- $\alpha$  blockers ( $n = 30$  infliximab,  $n = 8$  etanercept,  $n = 4$  adalimumab). The patients treated with TNF- $\alpha$  blockers had significantly higher serum levels of Wnt-3a ( $4.02 \pm 1.20$  ng/ml vs  $3.64 \pm 0.92$ ;  $p = 0.049$ ) and sRANKL ( $410.54 \pm 473.89$  pmol/l vs  $175.61 \pm 112.88$ ;  $p < 0.001$ ), but lower serum levels of OPG ( $3.25 \pm 0.87$  pmol/l vs  $3.84 \pm 1.14$ ;  $p = 0.002$ ) compared

with patients without a TNF- $\alpha$  blocker. There was no significant difference in age, BASMI, mSASSS, femoral neck BMD, or CRP between users and non-users of TNF- $\alpha$  blockers. The correlations between the biomarkers and BASMI, mSASSS, and BMD of femoral neck did not change in any significant way when the patients taking TNF- $\alpha$  blockers were excluded from the analysis. No significant difference in serum levels of the biomarkers was found between users (daily or on-demand) and nonusers of NSAID.

## DISCUSSION

In our study the serum levels of Wnt-3a were significantly elevated in the patients with AS and associated with higher BASMI and mSASSS, but not with age. In addition, serum Wnt-3a remained independently associated with BASMI and mSASSS after adjusting for age, sex, smoking, and CRP in multiple linear regressions. The results indicate that serum Wnt-3a may be a marker for the osteoproliferative process in AS.

Wnt-3a was the first of the Wnt to be isolated from cell cultures in its active form and characterized<sup>30</sup>. It is reasonable to assume that Wnt-3a has osteoproliferative effects because it has been shown to activate both canonical and noncanonical Wnt pathways in mesenchymal stem cells and to promote osteoblast differentiation, proliferation, and survival<sup>31,32,33</sup>. Total knockout (KO) of Wnt-3a causes embryonic lethality in mice, but heterozygote KO mice display bone loss and low BMD and trabecular number<sup>34</sup>. Animal models give support to the importance of Wnt signaling in the osteoproliferative process of spondyloarthritides. Increased Wnt signaling caused by DKK-1-blocking

antibodies has been shown to induce fusion of sacroiliac joints in mice transgenic for TNF<sup>35</sup>. In 1 mouse model, the DKK-1 blockage reversed the RA-like bone-destructive pattern to a more osteoarthritis (OA)-like bone-forming pattern with growth of osteophytes in the inflamed joints<sup>18</sup>.

Extensive syndesmophyte formation is a risk factor for osteoporosis and these conditions often coexist. Low BMD of the femoral neck was also correlated with higher Wnt-3a, but only in the univariate analysis.

In our study, high CRP levels were correlated with lower levels of both sclerostin and DKK-1, suggesting a possible connection between inflammation and reduced inhibition of Wnt signaling. Elevated CRP has been associated with increased syndesmophyte formation in AS and in early axial spondyloarthritis (SpA) in previous studies<sup>8,36</sup>. A recent study on patients with AS treated with TNF blockers also reported an association between persistently elevated CRP and low sclerostin<sup>37</sup>.

We found that DKK-1 was positively correlated with sclerostin, which is supported by a 2-year followup study on early AS that also reported significantly lower levels of DKK-1 measured with functional ELISA in patients with syndesmophyte growth compared with patients without syndesmophyte growth<sup>38</sup>. We found significantly lower DKK-1 in the patients with the highest mSASSS score. In our study, however, we measured total DKK-1 with sandwich ELISA.

Also supporting the role of Wnt signaling, we found significantly lower serum sclerostin in the patients with AS compared with healthy controls, a finding consistent with those of 2 previous studies<sup>37,39</sup>. We found that increasing levels of sclerostin were associated with increasing BASMI and mSASSS, which may be explained by sclerostin being positively correlated with age in both the patients with AS and the controls. In an earlier prospective study over 2 years, low serum sclerostin was associated with radiologic progression<sup>39</sup>. Decreased expression of sclerostin was also found in osteocytes in immunohistochemical analyses of zygapophyseal joints from patients with AS and in OA<sup>40</sup>.

Interestingly, we found that patients treated with TNF inhibition had higher serum levels of Wnt-3a, but no difference in serum levels of total DKK-1 was detected between the groups. Resolution of inflammation following anti-TNF treatment has been associated with new bone formation in AS<sup>41</sup>. It has been hypothesized that this could be caused by increased Wnt signaling and that TNF- $\alpha$  acts as a brake on bone formation by stimulating the expression of the Wnt antagonist DKK-1<sup>42</sup>. Our findings give some support to the theory that anti-TNF treatment is associated with increased Wnt activity, but caution should be taken in the interpretation, because only 21% of the patients were treated with TNF- $\alpha$  blockers and the study was not designed to evaluate the effects of treatment.

Anti-TNF- $\alpha$  treatment has been shown to improve BMD

in SpA in earlier studies<sup>43,44</sup>. In our study, anti-TNF treatment was associated with increased serum levels of sRANKL and lower levels of OPG. Studies on TNF inhibition and levels of sRANKL and OPG in rheumatoid arthritis (RA) have shown both unchanged or lowered levels of sRANKL and unchanged or decreased levels of OPG<sup>45</sup>.

We found the levels of serum sRANKL and the sRANKL/OPG ratio to be lower in patients with AS compared with the healthy controls. In contrast to our findings, 2 earlier studies with younger cohorts (mean age 32 and 34 yrs) of 60 and 42 patients showed higher levels of sRANKL in AS compared with controls, and in another investigation of 21 patients (mean age 51 yrs), no difference was shown<sup>46,47,48</sup>. Previous studies on serum RANKL in relation to osteoporosis in men and postmenopausal women have also yielded conflicting results<sup>49</sup>.

In our study, serum levels of OPG were positively correlated with BASMI and aging. The data are corroborated by an earlier study showing similar association between elevated levels of OPG and poorer spinal mobility and functional outcome<sup>47</sup>. Serum concentrations of OPG are increased in aging healthy women and men and in patients with osteoporosis<sup>50,51</sup>. This is viewed as a compensatory mechanism to prevent further bone loss. An earlier study on OPG in AS demonstrated a lack of age-related OPG increase in patients with AS, but this was not confirmed in 2 other studies including the present<sup>48,52</sup>. We found a negative correlation between OPG and BMD of the femoral neck. Diverging results regarding correlation between OPG and BMD have been reported before<sup>48,52</sup>.

Cigarette smoking has been identified as a risk factor for radiologic progression, impairment of back mobility, and functional outcome in AS and early axial SpA<sup>36,53,54</sup>. Our study supports the prior findings. We found that smoking was strongly associated with higher mSASSS, BASMI, age, and lower femoral neck BMD. In addition, high number of smoking pack-years was an independent covariate for mSASSS in the multivariate analyses after adjusting for age.

A limitation of our study was the cross-sectional design. A longitudinal study is needed to show whether the elevated levels of Wnt-3a just reflect presence of chronic AS related changes in the spine or whether Wnt-3a is a predictor for radiologic progression. The validity of the Wnt-3a analysis also needs to be confirmed in other cohorts of patients with AS and in early SpA. It would also be of interest to investigate the levels in other diseases, such as RA and osteoporosis, and to study other members of the Wnt family with respect to disease activity, osteoproliferation, and osteoporosis in AS. A strength of our study was that the biomarkers were studied both in relation to osteoproliferation and BMD, and to the best of our knowledge, was the first to do so.

We analyzed serum levels of biomarkers for bone metabolism in a cohort of 204 well-characterized patients with

AS. Most importantly, we found that serum Wnt-3a was significantly higher in the patients with AS and positively correlated with mSASSS and BASMI. After adjusting for sex, age, smoking, and CRP using multiple linear regression, serum Wnt-3a remained independently associated with mSASSS and BASMI, indicating that Wnt-3a could be a marker for the osteoproliferative process in AS.

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## REFERENCES

1. Francois RJ, Gardner DL, Degraive EJ, Bywaters EG. Histopathologic evidence that sacroiliitis in ankylosing spondylitis is not merely enthesitis. *Arthritis Rheum* 2000;43:2011-24.
2. Lories RJ, Schett G. Pathophysiology of new bone formation and ankylosis in spondyloarthritis. *Rheum Dis Clin North Am* 2012;38:555-67.
3. van der Heijde D, Landewe R, Baraliakos X, Houben H, van Tubergen A, Williamson P, et al. Radiographic findings following two years of infliximab therapy in patients with ankylosing spondylitis. *Arthritis Rheum* 2008;58:3063-70.
4. van der Heijde D, Landewe R, Einstein S, Ory P, Vosse D, Ni L, et al. Radiographic progression of ankylosing spondylitis after up to two years of treatment with etanercept. *Arthritis Rheum* 2008;58:1324-31.
5. Marzo-Ortega H, Emery P, McGonagle D. The concept of disease modification in spondyloarthropathy. *J Rheumatol* 2002;29:1583-5.
6. Haroon N, Inman RD, Learch TJ, Weisman MH, Lee M, Rahbar MH, et al. The impact of tumor necrosis factor alpha inhibitors on radiographic progression in ankylosing spondylitis. *Arthritis Rheum* 2013;65:2645-54.
7. Poddubnyy D, Rudwaleit M, Haibel H, Listing J, Märker-Hermann E, Zeidler H, et al. Effect of non-steroidal anti-inflammatory drugs on radiographic spinal progression in patients with axial spondyloarthritis: results from the German Spondyloarthritis Inception Cohort. *Ann Rheum Dis* 2012;71:1616-22.
8. Kroon F, Landewe R, Dougados M, van der Heijde D. Continuous NSAID use reverts the effects of inflammation on radiographic progression in patients with ankylosing spondylitis. *Ann Rheum Dis* 2012;71:1623-9.
9. Wanders A, Heijde D, Landewe R, Béhier JM, Calin A, Olivieri I, et al. Nonsteroidal antiinflammatory drugs reduce radiographic progression in patients with ankylosing spondylitis: a randomized clinical trial. *Arthritis Rheum* 2005;52:1756-65.
10. Klingberg E, Geijer M, Göthlin J, Mellström D, Lorentzon M, Hilme E, et al. Vertebral fractures in ankylosing spondylitis are associated with lower bone mineral density in both central and peripheral skeleton. *J Rheumatol* 2012;39:1987-95.
11. Westerveld LA, Verlaan JJ, Oner FC. Spinal fractures in patients with ankylosing spinal disorders: a systematic review of the literature on treatment, neurological status and complications. *Eur Spine J* 2009;18:145-56.
12. Donnelly S, Doyle DV, Denton A, Rolfe I, McCloskey EV, Spector TD. Bone mineral density and vertebral compression fracture rates in ankylosing spondylitis. *Ann Rheum Dis* 1994;53:117-21.
13. El Maghraoui A, Borderie D, Cherruau B, Edouard R, Dougados M, Roux C. Osteoporosis, body composition, and bone turnover in ankylosing spondylitis. *J Rheumatol* 1999;26:2205-9.
14. Geusens P, Vosse D, van der Linden S. Osteoporosis and vertebral fractures in ankylosing spondylitis. *Curr Opin Rheumatol* 2007;19:335-9.
15. Krishnan V, Bryant HU, Macdougald OA. Regulation of bone mass by Wnt signaling. *J Clin Invest* 2006;116:1202-9.
16. Monroe DG, McGee-Lawrence ME, Oursler MJ, Westendorf JJ. Update on Wnt signaling in bone cell biology and bone disease. *Gene* 2012;492:1-18.
17. Westendorf JJ, Kahler RA, Schroeder TM. Wnt signaling in osteoblasts and bone diseases. *Gene* 2004;341:19-39.
18. Diarra D, Stolina M, Polzer K, Zwerina J, Ominsky MS, Dwyer D, et al. Dickkopf-1 is a master regulator of joint remodeling. *Nat Med* 2007;13:156-63.
19. Johnson ML, Harnish K, Nusse R, Van Hul W. LRP5 and Wnt signaling: a union made for bone. *J Bone Miner Res* 2004;19:1749-57.
20. Neve A, Corrado A, Cantatore FP. Osteocytes: central conductors of bone biology in normal and pathological conditions. *Acta Physiol* 2012;204:317-30.
21. Bonewald LF, Johnson ML. Osteocytes, mechanosensing and Wnt signaling. *Bone* 2008;42:606-15.
22. Robling AG, Niziolek PJ, Baldridge LA, Condon KW, Allen MR, Alam I, et al. Mechanical stimulation of bone in vivo reduces osteocyte expression of Sost/sclerostin. *J Biol Chem* 2008;283:5866-75.
23. Balemans W, Ebeling M, Patel N, Van Hul E, Olson P, Dioszegi M, et al. Increased bone density in sclerosteosis is due to the deficiency of a novel secreted protein (SOST). *Hum Mol Genet* 2001;10:537-43.
24. Boyce BF, Xing L. Biology of RANK, RANKL, and osteoprotegerin. *Arthritis Res Therapy* 2007;9 Suppl 1:S1.
25. Boyle WJ, Simonet WS, Lacey DL. Osteoclast differentiation and activation. *Nature* 2003;423:337-42.
26. Klingberg E, Lorentzon M, Mellström D, Geijer M, Göthlin J, Hilme E, et al. Osteoporosis in ankylosing spondylitis — prevalence, risk factors and methods of assessment. *Arthritis Res Ther* 2012;14:R108.
27. Lukas C, Landewe R, Sieper J, Dougados M, Davis J, Braun J, et al. Development of an ASAS-endorsed disease activity score (ASDAS) in patients with ankylosing spondylitis. *Ann Rheum Dis* 2009;68:18-24.
28. Sieper J, Rudwaleit M, Baraliakos X, Brandt J, Braun J, Burgos-Vargas R, et al. The Assessment of SpondyloArthritis international Society (ASAS) handbook: a guide to assess spondyloarthritis. *Ann Rheum Dis* 2009;68 Suppl 2:ii1-44.
29. Creemers MC, Franssen MJ, van't Hof MA, Gribnau FW, van de Putte LB, van Riel PL. Assessment of outcome in ankylosing spondylitis: an extended radiographic scoring system. *Ann Rheum Dis* 2005;64:127-9.
30. Willert K, Brown JD, Danenberg E, Duncan AW, Weissman IL, Reya T, et al. Wnt proteins are lipid-modified and can act as stem cell growth factors. *Nature* 2003;423:448-52.
31. Qiu W, Chen L, Kassem M. Activation of non-canonical Wnt/JNK pathway by Wnt3a is associated with differentiation fate determination of human bone marrow stromal (mesenchymal) stem cells. *Biochem Biophys Res Commun* 2011;413:98-104.
32. Almeida M, Han L, Bellido T, Manolagas SC, Kousteni S. Wnt proteins prevent apoptosis of both uncommitted osteoblast progenitors and differentiated osteoblasts by beta-catenin-dependent and -independent signaling cascades involving Src/ERK and phosphatidylinositol 3-kinase/AKT. *J Biol Chem* 2005;280:41342-51.
33. Boland GM, Perkins G, Hall DJ, Tuan RS. Wnt 3a promotes proliferation and suppresses osteogenic differentiation of adult human mesenchymal stem cells. *J Cell Biochem* 2004;93:1210-30.
34. Takada I, Mihara M, Suzawa M, Ohtake F, Kobayashi S, Igarashi M, et al. A histone lysine methyltransferase activated by non-canonical Wnt signaling suppresses PPAR-gamma

- transactivation. *Nat Cell Biol* 2007;9:1273-85.
35. Uderhardt S, Diarra D, Katzenbeisser J, David JP, Zwerina J, Richards W, et al. Blockade of Dickkopf (DKK)-1 induces fusion of sacroiliac joints. *Ann Rheum Dis* 2010;69:592-7.
  36. Poddubnyy D, Haibel H, Listing J, Märker-Hermann E, Zeidler H, Braun J, et al. Baseline radiographic damage, elevated acute-phase reactant levels, and cigarette smoking status predict spinal radiographic progression in early axial spondylarthritis. *Arthritis Rheum* 2012;64:1388-98.
  37. Saad CG, Ribeiro AC, Moraes JC, Takayama L, Goncalves CR, Rodrigues MB, et al. Low sclerostin levels: a predictive marker of persistent inflammation in ankylosing spondylitis during anti-tumor necrosis factor therapy? *Arthritis Res Ther* 2012;14:R216.
  38. Heiland GR, Appel H, Poddubnyy D, Zwerina J, Hueber A, Haibel H, et al. High level of functional dickkopf-1 predicts protection from syndesmophyte formation in patients with ankylosing spondylitis. *Ann Rheum Dis* 2012;71:572-4.
  39. Appel H, Ruiz-Heiland G, Listing J, Zwerina J, Herrmann M, Mueller R, et al. Altered skeletal expression of sclerostin and its link to radiographic progression in ankylosing spondylitis. *Arthritis Rheum* 2009;60:3257-62.
  40. Power J, Poole KE, van Bezooijen R, Doube M, Caballero-Alías AM, Lowik C, et al. Sclerostin and the regulation of bone formation: Effects in hip osteoarthritis and femoral neck fracture. *J Bone Miner Res* 2010;25:1867-76.
  41. Pedersen SJ, Chiowchanwisawakit P, Lambert RG, Ostergaard M, Maksymowych WP. Resolution of inflammation following treatment of ankylosing spondylitis is associated with new bone formation. *J Rheumatol* 2011;38:1349-54.
  42. Maksymowych WP. Disease modification in ankylosing spondylitis. *Nat Rev Rheumatol* 2010;6:75-81.
  43. Briot K, Gossec L, Kolta S, Dougados M, Roux C. Prospective assessment of body weight, body composition, and bone density changes in patients with spondyloarthritis receiving anti-tumor necrosis factor-alpha treatment. *J Rheumatol* 2008;35:855-61.
  44. Marzo-Ortega H, McGonagle D, Haugeberg G, Green MJ, Stewart SP, Emery P. Bone mineral density improvement in spondyloarthritis after treatment with etanercept. *Ann Rheum Dis* 2003;62:1020-1.
  45. Barnabe C, Hanley DA. Effect of tumor necrosis factor alpha inhibition on bone density and turnover markers in patients with rheumatoid arthritis and spondyloarthritis. *Sem Arthritis Rheum* 2009;39:116-22.
  46. Kim HR, Kim HY, Lee SH. Elevated serum levels of soluble receptor activator of nuclear factors-kappaB ligand (sRANKL) and reduced bone mineral density in patients with ankylosing spondylitis (AS). *Rheumatology* 2006;45:1197-200.
  47. Chen CH, Chen HA, Liao HT, Liu CH, Tsai CY, Chou CT. Soluble receptor activator of nuclear factor-kappaB ligand (RANKL) and osteoprotegerin in ankylosing spondylitis: OPG is associated with poor physical mobility and reflects systemic inflammation. *Clin Rheumatol* 2010;29:1155-61.
  48. Stuppahann D, Rauner M, Krenbek D, Patsch J, Pirker T, Muschitz C, et al. Intracellular and surface RANKL are differentially regulated in patients with ankylosing spondylitis. *Rheumatol Int* 2008;28:987-93.
  49. Kearns AE, Khosla S, Kostenuik PJ. Receptor activator of nuclear factor kappaB ligand and osteoprotegerin regulation of bone remodeling in health and disease. *Endocr Rev* 2008;29:155-92.
  50. Yano K, Tsuda E, Washida N, Kobayashi F, Goto M, Harada A, et al. Immunological characterization of circulating osteoprotegerin/osteoclastogenesis inhibitory factor: increased serum concentrations in postmenopausal women with osteoporosis. *J Bone Miner Res* 1999;14:518-27.
  51. Szulc P, Hofbauer LC, Heufelder AE, Roth S, Delmas PD. Osteoprotegerin serum levels in men: correlation with age, estrogen, and testosterone status. *J Clin Endocrinol Metab* 2001;86:3162-5.
  52. Franck H, Meurer T, Hofbauer LC. Evaluation of bone mineral density, hormones, biochemical markers of bone metabolism, and osteoprotegerin serum levels in patients with ankylosing spondylitis. *J Rheumatol* 2004;31:2236-41.
  53. Avers HL, Oxtoby J, Taylor HG, Jones PW, Dziedzic K, Dawes PT. Smoking and outcome in ankylosing spondylitis. *Scand J Rheumatol* 1996;25:138-42.
  54. Doran MF, Brophy S, MacKay K, Taylor G, Calin A. Predictors of longterm outcome in ankylosing spondylitis. *J Rheumatol* 2003;30:316-20.