ABSTRACT. Objective. The differential diagnosis between rheumatic diseases and infectious conditions is a great challenge in clinical practice. Adult-onset Still’s disease (AOSD) is a rare systemic inflammatory syndrome that shares several clinical and laboratory variables with sepsis. Interleukin (IL)-18 is overexpressed in AOSD, suggesting a possible role as a disease biomarker. The aim of our study was to detect IL-18 serum levels in a cohort of patients with AOSD and sepsis and to address its possible role as a biomarker for differential diagnosis.

Methods. A group of unselected patients with AOSD diagnosed according to the Yamaguchi criteria and consecutive patients with sepsis diagnosed according to the American College of Chest Physicians/Society of Critical Care Medicine Consensus Conference criteria were enrolled. The clinical and laboratory data were collected. In the AOSD group, disease activity was assessed by Pouchot’s and Rau’s criteria. IL-18 serum levels were detected by ELISA.

Results. Thirty-nine patients with AOSD and 18 patients with sepsis were enrolled. Two out of 18 patients with sepsis (11.1%) also fulfilled the Yamaguchi criteria. A significant difference was found in IL-18 serum levels between patients with active and inactive disease (p < 0.001), and it positively correlated with disease activity (p = 0.0003), ferritin serum level (p = 0.016), and erythrocyte sedimentation rate (p = 0.041). IL-18 was significantly increased in patients with AOSD when compared with sepsis (p = 0.014). For a cutoff of 148.9 pg/ml, this test had a specificity of 78.3% and a sensitivity of 88.6%.

Conclusion. We have demonstrated that IL-18 can be a biomarker for differential diagnosis between AOSD and sepsis. (First Release May 1 2014; J Rheumatol 2014;41:1118–23; doi:10.3899/jrheum.130575)

Key Indexing Terms:
ADULT-ONSET STILL’S DISEASE          SEPSIS          INTERLEUKIN 18          FERRITIN
(IL-18), an immunoregulatory and inflammatory cytokine, has attracted interest as a potential therapeutic target in autoimmune/inflammatory disorders. We confirmed the presence of high serum levels of IL-18 in AOSD and demonstrated its possible role as a disease biomarker. This cytokine seems to be overexpressed also in patients with sepsis, where the inflammatory process is driven by a cytokine storm. Thus, the aim of our study was to detect IL-18 serum level in a cohort of patients with AOSD and sepsis and to address its possible role as a biomarker for differential diagnosis.

MATERIALS AND METHODS
A group of unselected patients with AOSD diagnosed according to the Yamaguchi criteria and followed at the Rheumatology Unit of Sapienza University of Rome and consecutive patients with sepsis diagnosed according to the American College of Chest Physicians/Society of Critical Care Medicine Consensus Conference criteria and hospitalized at the Infectious Disease Unit of the same hospital were enrolled in our study. Patients were not matched for age and sex owing to the rarity of the disease. For this reason we evaluated all the possible serum samples collected from patients with AOSD during the last 10 years in our unit. The clinical and laboratory data [erythrocyte sedimentation rate (ESR), C-reactive protein (CRP), white blood cell count (WBC), and ferritin] were collected in both groups. Patients of the sepsis group were also screened to verify whether they fulfilled the Yamaguchi criteria. In the AOSD group, disease activity was assessed either by the criteria proposed by Pouchot in 1991 or by those modified by Rau in 2010. In both sets of criteria, the total score ranges from 0 to 12 and is calculated through the addition of points assigned to each symptom. The Pouchot’s score uses the following items: fever, evanescent rash, pleurisy, pneumonia, pericarditis, hepatomegaly or abnormal liver function tests, splenomegaly, lymphadenopathy, WBC > 15000/mm³, sore throat, myalgias, and abdominal pain. In the score proposed by Rau, “splenomegaly” and “abdominal pain” were changed to “ferritin serum levels ≥ 3000 µg/l”, and “arthritis”. Patients with a Pouchot’s or Rau’s score ≥ 4 were considered to have active disease. Blood samples were collected from patients with active and inactive AOSD and from patients with sepsis during the febrile attack (first 48 h of hospitalization). IL-18 serum level was detected by means of ELISA (Immuno Pharmacol Research) as previously described. All subjects provided their informed consent.

The values were expressed as mean ± SD. For the statistical analysis, the Mann-Whitney U test and Spearman’s rank correlation test were used. Two-tailed p values < 0.05 were considered significant. Area under the receiver-operating characteristic curve (ROC-AUC) analysis was used to evaluate the diagnostic utility of the IL-18 serum level. The evaluation of the laboratory features between patients with active and inactive disease was performed using Rau’s criteria.

RESULTS
Thirty-nine patients with AOSD (F = 25, 64.1%; M = 14, 35.9%; mean age ± SD = 35 ± 14 yrs, mean age at onset ± SD = 32 ± 12 yrs) and 18 patients with sepsis (F = 8, 44.4%; M = 10, 55.6%; mean age ± SD = 54 ± 17.4 yrs) were enrolled. Seventeen out of 39 (43.5%) and 23 out of 39 (58.9%) patients with AOSD had active disease when the serum was taken, according to Pouchot’s criteria and Rau’s criteria, respectively. Two of the 18 patients with sepsis (11.1%) also fulfilled the Yamaguchi criteria. Main clinical features from patients with AOSD and sepsis are reported in Table 1. A significant difference in ferritin serum level was found between patients with active and inactive AOSD (active disease mean ± SD = 6462.4 ± 8427.7, inactive disease mean ± SD = 416.7 ± 466.8; p = 0.039). IL-18 mean value in sera of the whole cohort of patients with AOSD was 1043.9 ± 1831.4 pg/ml. A significant difference was found in IL-18 serum level between patients with active disease and patients with inactive disease (1716.8 ± 2182.6 vs 118.7 ± 159.2 pg/ml, p < 0.001; Figure 1). IL-18 serum level positively correlated with disease activity (p = 0.0003; Figure 2A), ferritin serum level (p = 0.016; Figure 2B), and ESR (p = 0.041; Figure 2C). In patients with sepsis, mean IL-18 serum level was 91.2 ± 73.3 pg/ml, thus significantly lower when compared with the whole AOSD cohort (p = 0.014; Figure 1) as well as only with the patients with inactive disease (p < 0.0001; Figure 1). The area under the ROC-AUC for detection of AOSD by reference to the level of serum IL-18 was 0.864 (Figure 3). For a cutoff of IL-18 of 148.9 pg/ml, this test had a specificity of 78.3% and a sensitivity of 88.6%; likelihood ratio was 2.89. Concerning other laboratory features, there was no significant difference in ESR (p = 0.769), CRP (p = 0.62), WBC (p = 0.14), and ferritin serum level (p = 0.27) between AOSD and sepsis (Table 2). Moreover, in patients with sepsis, IL-18 level did not correlate with any of the serological variables: ESR (p = 0.79), CRP (p = 0.46), WBC (p = 0.98), and ferritin (p = 0.10).

DISCUSSION
To our knowledge, we demonstrated for the first time that IL-18 can be a biomarker for differential diagnosis between AOSD and sepsis. Prompt and correct diagnosis of these life-threatening conditions may allow proper treatment and lifesaving action. Sometimes a clinical and serological overlap between AOSD and sepsis can be observed. Indeed, about 11% of our patients with sepsis also fulfilled the Yamaguchi criteria, and the diagnosis could have been puzzling. In our opinion, this evidence supports the idea that, at least in a small percentage of cases, clinical aspects of the life-threatening conditions may allow proper treatment and lifesaving action. Sometimes a clinical and serological overlap between AOSD and sepsis can be observed. Indeed, about 11% of our patients with sepsis also fulfilled the Yamaguchi criteria, and the diagnosis could have been puzzling. In our opinion, this evidence supports the idea that, at least in a small percentage of cases, clinical aspects of the life-threatening conditions may allow proper treatment and lifesaving action.

Table 1. Clinical features in 39 patients with adult-onset Still’s disease (AOSD) and 18 with sepsis.

<table>
<thead>
<tr>
<th>Signs and Symptoms</th>
<th>AOSD (% of patients)</th>
<th>Sepsis (% of patients)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fever</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>Rash</td>
<td>76.9</td>
<td>9</td>
</tr>
<tr>
<td>Arthritis</td>
<td>64.1</td>
<td>9</td>
</tr>
<tr>
<td>Sore throat</td>
<td>64.1</td>
<td>9</td>
</tr>
<tr>
<td>Lymphadenopathy</td>
<td>58.9</td>
<td>27.2</td>
</tr>
<tr>
<td>Hepatomegaly/increased liver enzymes</td>
<td>48.7</td>
<td>45.4</td>
</tr>
<tr>
<td>Myalgia</td>
<td>38.4</td>
<td>18.1</td>
</tr>
<tr>
<td>Splenomegaly</td>
<td>15.3</td>
<td>36.3</td>
</tr>
<tr>
<td>Pericarditis</td>
<td>10.2</td>
<td>0</td>
</tr>
<tr>
<td>Pleurisy</td>
<td>7.6</td>
<td>0</td>
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have a pathogenic role in several autoinflammatory condi-
tions and its production is driven by the “inflammasome”
activation. Indeed, the inflammasome induction can be
mediated by different innate immune receptors (Toll-like
receptors or NOD-like receptors) previously activated by
external pathogens (pathogen-associated molecular
patterns) or by endogenous molecules (damage-associated
molecular patterns). Such structures mediate the innate
immune system response, promoting the production of
inflammatory caspases and activating proinflammatory
cytokines such as IL-1β and IL-18. Because of the
promising results obtained by the use of anti-IL-1 agents in
AOSD as well as in other autoinflammatory conditions,
modulation of these pathways is now the main therapeutic
target. Following its production, IL-18 interacts with a
complex receptor (IL-18R), which is widely expressed on
cells implicated in innate and acquired immunity. IL-18
was initially identified as a major inducer of interferon
(IFN)-γ and is a critical regulator of Th1 cell maturation.
Otherwise, depending on the context of stimulation,
IL-18 may promote either Th1 or Th2 response.

It has been demonstrated that not only classic anti-
gen-presenting cells, but also nonimmune cells such as
synovial fibroblasts, intestinal epithelial cells, and keratino-
cyes, contribute in a proinflammatory manner as
IL-18-producing cells. Thus, IL-18 provides a critical link
between innate and acquired immune responses with the
involvement of both nonprofessional and professional
immune cell lineages. This is why an increasing interest in
the role of this cytokine has been emerged in connection
with different kinds of autoimmune/inflammatory disorders
such as AOSD.

In previous studies, we have shown that IL-18 is
over-expressed in sera as well as in lymph nodes and liver
of patients with AOSD. Otherwise, it has been
suggested that IL-18 plays an important role in the patho-
physiology of sepsis. Infections, this cytokine has
a key function because of its ability to regulate innate and
acquired immune response. Further, it has been demon-
strated that it participates in the defense against intracellular
bacteria including Listeria, Shigella, Salmonella, and
demonstrated that stimulation of blood monocytes with
Staphylococcus aureus causes a superinduction of IL-18
compared with lipopolysaccharide. Thereafter, a compar-
ison of plasma IL-18 concentrations between patients with
gram-positive and gram-negative sepsis showed signifi-
cantly higher levels of this cytokine in the group with
gram-positive infection.

For this reason, to identify a possible biomarker in
patients with AOSD, several studies have been undertaken.
Kawashima, et al. were the first to demonstrate the
markedly increased expression of IL-18 in systemic circu-
ation of patients with AOSD. Later on, Kawaguchi, et al.,
proposed IL-18 not only as a diagnostic marker for AOSD
but also as an indicator of disease severity. IL-18 is
considered a member of the IL-1 superfamily because of its
structural homology and biological effects. IL-1 seems to
have a pathogenic role in several autoinflammatory condi-
P<0.014

Figure 1. Comparison between IL-18 serum levels in the whole cohort of patients with AOSD (n = 39), patients with active disease (n = 17), and sepsis (n = 18). AOSD: adult-onset Still’s disease; IL: interleukin.
and evolution of organ dysfunction in sepsis\textsuperscript{27}. Within this group of cytokines, IL-18 seems to make the major contribution as central initiator and regulator of the inflammatory cascade\textsuperscript{6}. Nonetheless, Rau, et al\textsuperscript{9} compared a panel of serum inflammatory molecules (including IL-1\(\beta\), IL-6, IL-8, IL-10, IL-12, IFN-\(\gamma\), TNF, and calprotectin, as well as IL-18) without finding any significant difference between patients with sepsis and those with AOSD. The major limitation of that study was the low number of patients in the study cohort (18 patients with AOSD and 14 sepsis). Further, as the authors underlined, the observed broad range of IL-18 levels in all groups of patients may have biased the statistical analysis.

Interestingly, our results may integrate these findings. IL-18 serum level was significantly increased in the AOSD group when compared with sepsis, considering the whole cohort of patients and only the patients with active disease. Apparently, in our study, patients with sepsis presented IL-18 serum levels lower than previously reported\textsuperscript{21}. We cannot exclude that those levels may depend on the duration of septic disease and treatment. Indeed, although all the serum samples were taken within 48 h of hospitalization, some patients had already been treated with antibiotics at home. Moreover, such a finding is not surprising in light of the known variability described for IL-18 in the course of sepsis\textsuperscript{28}. A wide range of serum values has been

\textbf{Figure 2}. Comparison between IL-18 serum levels in patients with AOSD and (A) disease activity; (B) ferritin serum levels; and (C) erythrocyte sedimentation rate (ESR). AOSD: adult-onset Still’s disease; IL: interleukin.
documented\textsuperscript{29,30}, suggesting how such variability may be ascribed to different aspects, including the methodology used for cytokine detection\textsuperscript{31} or the different infectious agents responsible for sepsis\textsuperscript{6}. Rau and colleagues\textsuperscript{9} agreed, and their study did not show the data on IL-18 because of a too-broad range of values. This may lead us to hypothesize very low levels of IL-18 in the patients with sepsis in their study too. Further, in our study serum IL-18 detection was performed using an ELISA test, while in Rau’s a multiplex analysis was undertaken. As reported by Leng,\textsuperscript{31} experience with multiplex arrays remains limited, particularly in the context of human aging studies, and although good correlations between ELISA and multiplex have been reported, careful side-by-side comparisons are rare. Moreover, because there are no World Health Organization-accepted standards for normal age-adjusted cytokine levels, comparing results obtained using single ELISA kits from different manufacturers or comparing singleplex to multiplex results can be problematic\textsuperscript{31}.

Yet another consideration has to be taken in account. In 2001, Novick,\textit{et al}\textsuperscript{30} measured serum levels of both IL-18 and IL-18 binding protein (IL-18BP) in a large cohort of patients with sepsis. The authors found that total IL-18 and its binding protein were elevated, and most of the IL-18 was bound to IL-18BP. The authors concluded that the binding protein considerably inhibits the free circulating form of IL-18, thus giving rise to lower detectable levels.

In our study, IL-18 was the only serological variable that distinguished the 2 groups. No significant differences between AOSD and sepsis were found in ESR, CRP, WBC, and ferritin levels. Moreover, IL-18 serum level correlated with other serological markers of inflammation only in patients with AOSD. Because we did not observe such correlation in sepsis, we may suggest a marginal role of this cytokine in monitoring the inflammatory status of these patients. Further, we can confirm that in patients with AOSD, IL-18 serum level correlates with disease activity, and its levels were significantly different between patients with active and inactive disease.

Such correlation of IL-18 with AOSD disease activity was reported by Kim,\textit{et al}\textsuperscript{32}, although in that study a better correlation was observed of another molecule, S100A8/A9, with serological and clinical variables. Currently, in the evaluation of AOSD disease activity, both Pouchot’s and Rau’s criteria have been used. We found that Rau’s criteria were more sensitive in the detection of patients with active disease compared with Pouchot’s criteria. This finding is further supported by the evidence of a significant difference between patients with active and inactive AOSD in ferritin serum level, a variable included in Rau’s but not in Pouchot’s criteria.

Our results confirm the key role of IL-18 in AOSD and we suggest the detection of this cytokine as a useful tool for differential diagnosis between AOSD and sepsis. Moreover, we suggest that Rau’s criteria are more sensitive in defining AOSD disease activity.

**REFERENCES**


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**Figure 3.** Area under the receiver-operating characteristic curve (ROC) for detection of AOSD by reference to the level of serum IL-18. AOSD: adult-onset Still’s disease.

**Table 2.** Laboratory features in patients with AOSD and sepsis. Except for p value, data are mean (± SD).

<table>
<thead>
<tr>
<th>Features</th>
<th>AOSD</th>
<th>Sepsis</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>ESR, mm/h</td>
<td>60.4 (34.2)</td>
<td>61.3 (33.9)</td>
<td>0.769</td>
</tr>
<tr>
<td>CRP, mg/dl</td>
<td>73.2 (84.4)</td>
<td>116.4 (98.2)</td>
<td>0.62</td>
</tr>
<tr>
<td>WBC, cells/µl</td>
<td>15,822 (8038)</td>
<td>11,192.5 (7120.5)</td>
<td>0.14</td>
</tr>
<tr>
<td>Ferritin, ng/ml</td>
<td>3540.3 (6701.9)</td>
<td>1720 (3882)</td>
<td>0.27</td>
</tr>
<tr>
<td>IL-18, pg/ml</td>
<td>1043.9 (1831.4)</td>
<td>91.2 (73.3)</td>
<td>0.014</td>
</tr>
</tbody>
</table>

AOSD: adult-onset Still’s disease; ESR: erythrocyte sedimentation rate; CRP: C-reactive protein; WBC: white blood cells; IL: interleukin.
Priori, et al: IL-18 in AOSD and sepsis

2012;2012:156890.


