Endothelial Activation in Ankylosing Spondylitis

Blood Monocyte Heterogeneity and Markers of Endothelial Activation in Ankylosing Spondylitis

Andrzej Surdacki, Joanna Sulicka, Mariusz Korkosz, Tomasz Mikołajczyk, Dorota Telesińska-Jasiówka, Ewa Klimek, Izabella Kierzkowska, Tomasz Guzik, and Tomasz K. Grodzicki

ABSTRACT. Objective. Ankylosing spondylitis (AS) is associated with excessive cardiovascular (CV) morbidity. Interactions between activated endothelium and monocytes precede atherosclerotic plaques. Our aim was to quantify blood monocyte subsets in relation to endothelial activation and inflammatory activity in subjects with AS who were free of clinical atherosclerotic CV disease. Methods. Markers of inflammation and endothelial activation were measured in 47 patients with AS receiving no disease-modifying antirheumatic drugs, and 22 healthy controls. Exclusion criteria included atherosclerotic CV disease and traditional risk factors. Flow cytometry was used to identify monocyte subsets: classical CD14++CD16−, intermediate CD14++CD16+, and nonclassical CD14+CD16++ monocytes and to evaluate their expression of CD11b and CD11c. Results. Traditional risk factors were comparable among the groups, except for lower high-density lipoprotein cholesterol in AS (p = 0.007). Relative to controls, in subjects with AS counts of classical monocytes were higher (84.3 ± 5.4 vs 78.9 ± 5.3% of blood monocytes, p < 0.001) and nonclassical monocytes lower (2.9 ± 2.2 vs 5.5 ± 2.3%, p < 0.001). In AS we observed increased soluble intercellular adhesion molecule-1 (102 CCK3–293) vs 202 (187–230) ng/ml, p = 0.002), an endothelial ligand for monocyte beta-2-integrin CD11b/CD18. CD11b expression on all 3 monocyte subsets was elevated in 21 AS subjects with a Bath Ankylosing Spondylitis Disease Activity Index score ≥ 4 versus the remaining patients (p = 0.005–0.03). C-reactive protein, interleukin 6 (IL-6), and pentraxin-3 were increased in AS, in contrast to tumor necrosis factor-alpha and IL-18. IL-6 correlated with classical monocytes numbers in AS (r = 0.56, p < 0.0001) but not in the controls (r = 0.10, p = 0.65). Conclusion. Our findings suggest a contribution of immune dysregulation to enhanced monocyte-endothelial interactions in AS, especially in patients with active disease, which possibly can accelerate atherogenesis on a longerterm basis. (First Release Feb 1 2014; J Rheumatol 2014;41:481–9; doi:10.3899/jrheum.130803)

Key Indexing Terms: ANKYLOSING SPONDYLITIS MONOCYTES ENDOTHELIUM

Rheumatoid arthritis (RA), ankylosing spondylitis (AS), and psoriatic arthritis are associated with increased cardiovascular CV) mortality and morbidity because of accelerated atherogenesis and disease-specific CV manifestations1,2,3. Features of endothelial dysfunction, a predecessor of atherosclerotic plaques, have been observed in patients with each of these inflammatory arthropathies and free of clinical atherosclerotic CV disease or classical risk factors4,5,6,7,8,9.

Interactions between activated endothelium and blood monocytes precede development of atherosclerotic plaques10. It has been recognized that not only endothelial dysfunction10,11,12 but also a shift in blood monocyte subsets can predict adverse CV outcome in various clinical...
settings\textsuperscript{13,14,15,16}. Elevated counts of intermediate CD14\textsuperscript{++}CD16\textsuperscript{+} monocytes (a minor monocyte subpopulation presumably representing a transitory stage of differentiation of predominant classical CD14\textsuperscript{++}CD16\textsuperscript{−} monocytes toward nonclassical CD14\textsuperscript{+}CD16\textsuperscript{++} macrophage-like monocytes\textsuperscript{15}) were associated with increased risk of ischemic CV events in endstage renal disease\textsuperscript{13} and nondialysis chronic kidney disease (CKD)\textsuperscript{14}, and excessive risk of major CV events in patients referred for coronary angiography\textsuperscript{16}. An increased frequency of CD14/CD16–double-positive monocytes or intermediate CD14\textsuperscript{+}CD16\textsuperscript{+} monocytes was also reported in RA\textsuperscript{18,19,20} and psoriatic arthritis\textsuperscript{21}. We have recently described elevated counts of intermediate monocytes in young adult survivors of childhood acute lymphoblastic leukemia\textsuperscript{22}, also known to exhibit endothelial dysfunction\textsuperscript{23} and to be prone to late CV morbidity\textsuperscript{24}.

To our knowledge, no study has focused on blood monocyte heterogeneity in AS to date. Thus, our aim was to estimate frequencies of circulating monocyte subsets and their relations with markers of endothelial dysfunction and inflammatory activity in patients naïve to disease-modifying antirheumatic drugs (DMARD) and having AS free of clinical atherosclerotic CV disease.

**MATERIALS AND METHODS**

**Subjects.** We studied 47 patients aged 20–47 years (mean ± SD, 33 ± 6 yrs; 41 men and 6 women) with AS diagnosed according to the modified New York criteria\textsuperscript{25}. All patients had predominantly axial disease and were HLA-B27–positive. The patients were receiving only nonsteroidal anti-inflammatory drugs and none of them had been treated with nonbiologic DMARD, anti-tumor necrosis factor (anti-TNF) agents, or steroids. The mean disease duration was 8 ± 5 years. Bath Ankylosing Spondylitis Disease Activity Index (BASDAI) scores\textsuperscript{26} averaged 4.1 ± 2.0 (range: 1.4–9.5). We also recruited a control group of 22 healthy volunteers aged 22–42 years (mean 30 ± 7 yrs; 12 men and 10 women).

Exclusion criteria were common for both groups and included clinical evidence of atherosclerotic CV disease, diabetes, hypertension, an estimated glomerular filtration rate (eGFR) below 90 ml/min per 1.73 m\textsuperscript{2} of body surface area, a history of malignancy, acute infections within previous 4 weeks, and any other chronic coexistent diseases. In agreement with the Declaration of Helsinki, the protocol had been approved by the ethics committee of our university, and written informed consent was obtained from each subject before inclusion in the study.

**Study protocol.** The procedure was carried out in the morning in the out-patient clinic of our center. The subjects had been asked to refrain from eating, smoking, and alcohol or caffeine consumption for at least 12 h prior to the study. The personnel involved in the protocol recorded demographic and clinical characteristics, anthropometric measurements were taken, and blood pressure was measured on the left arm after 5 min of rest in a sitting position. Then all participants underwent blood sampling for biochemical assays and flow cytometry.

**Biochemical assays.** A sample of blood was drawn from a left antecubital vein for measurements of serum concentrations of high-sensitivity C-reactive protein (hs-CRP) by immunonephelometry (Dade Behring II Nephelometer, GMi Inc.). Glucose, lipids, and creatinine were measured with a Hitachi 917 analyzer (Roche Diagnostics) using standardized laboratory techniques. An eGFR was calculated by the Chronic Kidney Disease Epidemiology Collaboration formula, which has proven more accurate than the Modification of Diet in Renal Disease study equation, especially at normal renal function\textsuperscript{27}.

Plasma for extended biochemical analyses was collected from EDTA-anticoagulated blood and frozen at –80°C until assayed. Commercially available ELISA were used to measure plasma levels of TNF-\(\alpha\) and interleukin 6 (IL)-6 (Quantikine high-sensitivity kits), monocyte chemotactic protein-1 (MCP-1), pentraxin-3 (PTX-3), soluble forms of vascular cell adhesion molecule-1 (sVCAM-1) and intercellular adhesion molecule-1 (sICAM-1; all R&D Systems), and IL-18 (MBL).

**Flow cytometry.** As described\textsuperscript{22}, peripheral blood mononuclear cells were isolated from EDTA-anticoagulated blood and collected on the same day as remaining samples, using density gradient centrifugation and LSM 1077 (Lymphocyte Separation Medium, PAA Laboratories GmbH). One hundred thousand cells were stained for 20 min with fluorochrome-conjugated monoclonal antibodies (anti-CD14-APC-H7, anti-CD16-PE, anti-HLA-DR-PE-Cy7, anti-CD11b-Pacific Blue; and anti-CD11c-APC; Becton Dickinson Biosciences — Pharmingen) and then washed with phosphate buffered saline containing 1% fetal bovine serum (GIBCO). The cells were processed in the FACScanto II flow cytometer (BD Biosciences) and then analyzed with FACSDiva software (BD Biosciences) and FlowJo (TreeStar Inc.). The cells were gated in a side scatter/forward scatter plot with the scatter gate for monocytes partially extending into lymphocytes\textsuperscript{28}. The cells containing all monocytes and a part of the lymphocyte population were then gated in an HLA-DR/CD14 plot to exclude HLA-DR–negative natural killer cells (which would otherwise contaminate the CD14\textsuperscript{+}CD16\textsuperscript{++} subpopulation) and finally analyzed for CD14 and CD16 expression\textsuperscript{29,29}. Monocyte subsets were defined according to the expression of CD14 (Fc receptor type III) and CD14 (lipopolysaccharide receptor) as classical monocytes (CD14\textsuperscript{++}CD16\textsuperscript{+}), intermediate monocytes (CD14\textsuperscript{+}CD16\textsuperscript{+}), and nonclassical monocytes (CD14\textsuperscript{+}CD16\textsuperscript{+})\textsuperscript{17}. The expression of CD11b and CD11c on the 3 monocyte subsets was quantified. The results have been presented as either the percentage of cells expressing individual markers, absolute cell counts (per \(\mu\)l of blood), or mean fluorescence intensity (MFI). Total numbers of monocytes were calculated using peripheral white blood cell counts. In separate experiments BD Trucount beads were used, confirming that both methods yielded similar absolute values of cell counts.

All experiments were performed under standardized experimental conditions, including the procedure of cell isolation and reagents for sample preparation. The same number of cells and the same amounts of antibodies were used for samples. Simultaneously, in each case we checked the proper operation of the flow cytometer using BD Cytometer Setup and Tracking beads. Also, the operational stability was verified, so that the individual measurements did not differ significantly one from the other. Flow cytometry compensation was verified regularly using cells studied in the experiment and CompBeads. One operator performed the sample collection and the data analysis.

**Statistical analysis.** Data are presented as mean ± SD unless stated otherwise or medians (interquartile range) for continuous variables, and numbers (percentages) for categorical variables. The accordance with a normal distribution was tested by the Kolmogorov-Smirnov test, and uniformity of variance (homoscedasticity) by Levene’s test. Intergroup differences were assessed by the unpaired 2-sided Student’s t test for continuous variables and Fisher’s exact test for proportions. Logarithmic transformation (In, natural logarithm) was applied, when necessary, to obtain a normal distribution (hs-CRP, TNF-\(\alpha\), IL-6, sVCAM-1, PTX-3, percentage of intermediate monocytes) or to eliminate the nonuniformity of variances. The 2-sided Welch’s t test was used in case of the lack of homogeneity of variances even in In-transformed data. A p-value below 0.05 was considered significant. Bivariate relations between continuous variables were estimated by Pearson’s correlation coefficients (r). Owing to multiple testing, exclusive correlations with p ≤ 0.001 were inferred as significant. Analyses were performed by means of the Statistica data analysis software system (version 10.0.1011.0; StatSoft Inc.).
RESULTS

Clinical and biochemical characteristics of the patients with AS and healthy controls. Traditional CV risk factors were similar in patients with AS and healthy controls except for lower high-density lipoprotein (HDL) cholesterol, in agreement with a previous report30, and a higher ratio of total cholesterol to HDL cholesterol in AS (Table 1). Additionally, levels of fibrinogen, hs-CRP, IL-6, sICAM-1, and PTX-3 were significantly increased in patients with AS, whereas TNF-α, sVCAM-1, IL-18, and MCP-1 were similar between the groups (Table 2).

Blood monocyte subsets and their expression of CD11b and CD11c in patients with AS and healthy controls. Compared to the controls, patients with AS exhibited significantly higher counts of classical CD14++CD16– monocytes and lower numbers of nonclassical CD14+CD16+ monocytes (Table 3). There were no differences in the expression of CD11b and CD11c on all 3 monocyte subsets between AS subjects and the controls (p > 0.1).

Pooling both groups as a whole, men tended to have higher total monocyte counts than women (536 ± 208 vs 436 ± 115 cells/μl, p = 0.09). Because the percentage of men was higher in patients with AS (87% vs 55%), we repeated the comparisons after limitation of the analysis to male subjects, which revealed similar directions and levels of significance for intergroup differences in classical (p = 0.001) and nonclassical (p < 0.001) monocytes.

There was a correlation between IL-6 and the count of

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**Table 1.** Clinical and conventional biochemical characteristics of patients with ankylosing spondylitis (AS) and control subjects. Data are shown as medians (interquartile range) or n (%). P values in boldface are statistically significant.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Patients with AS, n = 47</th>
<th>Controls, n = 22</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, yrs</td>
<td>33 (28–36)</td>
<td>29 (22–38)</td>
<td>0.10</td>
</tr>
<tr>
<td>BASDAI</td>
<td>3.7 (2.6–4.7)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Disease duration, yrs</td>
<td>6 (4–10)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Male sex</td>
<td>41 (87)</td>
<td>13 (59)</td>
<td>0.008</td>
</tr>
<tr>
<td>Body mass index, kg/m²</td>
<td>24.3 (22.5–26.6)</td>
<td>23.3 (22.6–24.6)</td>
<td>0.15</td>
</tr>
<tr>
<td>Smoking</td>
<td>14 (30)</td>
<td>6 (27)</td>
<td>0.83</td>
</tr>
<tr>
<td>Mean blood pressure, mmHg</td>
<td>92 (87–97)</td>
<td>90 (87–92)</td>
<td>0.13</td>
</tr>
<tr>
<td>Glucose, mmol/l</td>
<td>4.7 (4.5–5.0)</td>
<td>4.7 (4.4–4.9)</td>
<td>0.72</td>
</tr>
<tr>
<td>eGFR, ml/min per 1.73 m²</td>
<td>121 (110–132)</td>
<td>120 (99–141)</td>
<td>0.15</td>
</tr>
<tr>
<td>Total cholesterol, mmol/l</td>
<td>5.0 (4.2–5.8)</td>
<td>4.8 (4.5–5.6)</td>
<td>0.66</td>
</tr>
<tr>
<td>LDL cholesterol, mmol/l</td>
<td>3.1 (2.4–3.8)</td>
<td>2.8 (2.3–3.5)</td>
<td>0.11</td>
</tr>
<tr>
<td>HDL cholesterol, mmol/l</td>
<td>1.3 (1.2–1.5)</td>
<td>1.7 (1.4–2.0)</td>
<td>0.007*</td>
</tr>
<tr>
<td>Total/HDL cholesterol</td>
<td>3.7 (3.2–4.3)</td>
<td>3.1 (2.5–3.7)</td>
<td>0.01</td>
</tr>
<tr>
<td>Triglycerides, mmol/l</td>
<td>0.9 (0.6–1.2)</td>
<td>0.9 (0.7–1.2)</td>
<td>0.48</td>
</tr>
<tr>
<td>hs-CRP, mg/l</td>
<td>5.05 (2.57–16.50)</td>
<td>0.88 (0.39–1.48)</td>
<td>&lt; 0.001†</td>
</tr>
<tr>
<td>Fibrinogen, g/l</td>
<td>4.17 (3.40–5.23)</td>
<td>3.09 (2.30–3.40)</td>
<td>&lt; 0.001*</td>
</tr>
</tbody>
</table>

* By Welch’s t test for untransformed data because of the non-uniformity of variances. † By Welch’s t test for ln-transformed data because of the lack of normal distribution and the non-uniformity of variances. BASDAI: Bath Ankylosing Spondylitis Disease Activity Index; NA: not applicable; eGFR: estimated glomerular filtration rate; LDL: low-density lipoprotein; HDL: high-density lipoprotein; hs-CRP: high-sensitivity C-reactive protein.

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**Table 2.** Extended inflammatory markers in ankylosing spondylitis (AS) patients and controls. Data are shown as medians (interquartile range). P values in boldface are statistically significant.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Patients with AS, n = 47</th>
<th>Controls, n = 22</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>TNF-α, pg/ml</td>
<td>1.76 (1.57–2.13)</td>
<td>1.49 (1.34–1.96)</td>
<td>0.17*</td>
</tr>
<tr>
<td>IL-6, pg/ml</td>
<td>3.51 (1.79–6.19)</td>
<td>0.74 (0.56–1.15)</td>
<td>&lt; 0.001†</td>
</tr>
<tr>
<td>IL-18, pg/ml</td>
<td>407 (322–493)</td>
<td>417 (285–472)</td>
<td>0.50</td>
</tr>
<tr>
<td>sICAM-1, ng/ml</td>
<td>251 (224–293)</td>
<td>202 (187–230)</td>
<td>0.002</td>
</tr>
<tr>
<td>sVCAM-1, ng/ml</td>
<td>585 (496–644)</td>
<td>551 (507–635)</td>
<td>0.69*</td>
</tr>
<tr>
<td>MCP-1, pg/ml</td>
<td>256 (211–307)</td>
<td>256 (218–293)</td>
<td>0.62</td>
</tr>
<tr>
<td>Pentraxin-3, pg/ml</td>
<td>589 (483–772)</td>
<td>420 (364–483)</td>
<td>0.002*</td>
</tr>
</tbody>
</table>

* By Student’s t test for natural logarithm (ln)–transformed data to obtain a normal distribution. † By Welch’s t test for ln-transformed data because of the lack of normal distribution and the nonuniformity of variances. TFN-α: tumor necrosis factor-α; IL: interleukin; sICAM-1: soluble intercellular adhesion molecule-1; sVCAM-1: soluble vascular cell adhesion molecule-1; MCP-1: monocyte chemotactic protein-1.

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classical monocytes in patients with AS ($r = 0.56$, $p < 0.0001$; (Table 4 and Figure 1A) but not in the controls ($r = 0.10$, $p = 0.65$; Figure 1B). Neither HDL cholesterol nor the ratio of total to HDL cholesterol correlated to any of the variables derived from flow cytometry ($p \geq 0.1$).

**Clinical and biochemical characteristics, monocyte subset size, and monocytic expression of CD11b and CD11c in relation to disease activity.** BASDAI correlated to the expression of CD11b on classical ($r = 0.47$, $p = 0.001$), intermediate ($r = 0.49$, $p < 0.001$), and nonclassical monocytes ($r = 0.52$, $p < 0.001$; Table 4). Accordingly, compared to AS patients with a BASDAI score $< 4$ ($n = 26$), AS patients with BASDAI $\geq 4$ ($n = 21$) had higher monocytic expression of CD11b on classical ($1529 \pm 1006$ vs $998 \pm 591 \text{ MFI}$, $p = 0.03$; Figure 2A), intermediate ($1591 \pm 899$ vs $1009 \pm 579 \text{ MFI}$, $p = 0.01$; Figure 2B), and nonclassical monocytes ($607 \pm 185$ vs $461 \pm 143 \text{ MFI}$, $p = 0.005$; Figure 2C). Relative increases in CD11b expression on classical, intermediate, and nonclassical monocytes associated with a high disease activity averaged $53\%$, $58\%$, and $32\%$, respectively, with the reference to the remaining subjects with AS. The expression of CD11b was reduced by about one-third in the patients with AS who had a low disease activity, relative to controls on all 3 monocyte subpopulations: classical ($998 \pm 591$ vs $1539 \pm 629 \text{ MFI}$, $p = 0.003$), intermediate ($1009 \pm 579$ vs $1523 \pm 584 \text{ MFI}$, $p = 0.004$), and nonclassical monocytes ($461 \pm 143$ vs $729 \pm 566 \text{ MFI}$, $p = 0.02$).

<table>
<thead>
<tr>
<th>Subset of Blood Monocytes</th>
<th>Patients with AS, $n = 47$</th>
<th>Healthy Subjects, $n = 22$</th>
<th>$p$</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD14**CD16+ classical monocytes</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>% of monocytes</td>
<td>$84.3 \pm 5.4$</td>
<td>$78.9 \pm 5.3$</td>
<td>$&lt; 0.001$</td>
</tr>
<tr>
<td>Per µl of blood</td>
<td>$469 \pm 194$</td>
<td>$364 \pm 124$</td>
<td>$0.02$</td>
</tr>
<tr>
<td>CD14**CD16+ intermediate monocytes</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>% of monocytes</td>
<td>$5.5 \pm 3.3$</td>
<td>$5.0 \pm 1.9$</td>
<td>$0.60^*$</td>
</tr>
<tr>
<td>Per µl of blood</td>
<td>$28 \pm 15$</td>
<td>$23 \pm 11$</td>
<td>$0.21$</td>
</tr>
<tr>
<td>CD14**CD16+ nonclassical monocytes</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>% of monocytes</td>
<td>$2.9 \pm 2.2$</td>
<td>$5.5 \pm 2.3$</td>
<td>$&lt; 0.001$</td>
</tr>
<tr>
<td>Per µl of blood</td>
<td>$15 \pm 10$</td>
<td>$25 \pm 12$</td>
<td>$&lt; 0.001$</td>
</tr>
<tr>
<td>Total monocytes (per µl of blood)</td>
<td>$542 \pm 211$</td>
<td>$462 \pm 148$</td>
<td>$0.12$</td>
</tr>
</tbody>
</table>

* By Student’s $t$ test for natural logarithm-transformed data to get a normal distribution.

Table 4. Pearson’s correlation coefficients between inflammatory markers, disease activity, monocyte subsets size, and monocytic CD11b and CD11c expression in patients with ankylosing spondylitis. Data in boldface are statistically significant.

<table>
<thead>
<tr>
<th></th>
<th>hs-CRP</th>
<th>Fibr.</th>
<th>IL-6</th>
<th>IL-18</th>
<th>TNF-α</th>
<th>BASDAI</th>
</tr>
</thead>
<tbody>
<tr>
<td>sICAM-1</td>
<td>0.15</td>
<td>0.30</td>
<td>0.08</td>
<td>0.22</td>
<td>–0.04</td>
<td>0.11</td>
</tr>
<tr>
<td>sVCAM-1</td>
<td>–0.11</td>
<td>–0.19</td>
<td>–0.21</td>
<td>–0.02</td>
<td>0.06</td>
<td>–0.16</td>
</tr>
<tr>
<td>Pentraxin-3</td>
<td>–0.04</td>
<td>–0.08</td>
<td>0.17</td>
<td>0.07</td>
<td>–0.19</td>
<td>0.23</td>
</tr>
<tr>
<td>Classical monocytes</td>
<td>0.33</td>
<td>0.37</td>
<td>0.58**</td>
<td>0.05</td>
<td>0.00</td>
<td>0.03</td>
</tr>
<tr>
<td>Classical monocytes, per µl of blood</td>
<td>0.39</td>
<td>0.42</td>
<td>0.56**</td>
<td>–0.04</td>
<td>0.03</td>
<td>0.05</td>
</tr>
<tr>
<td>CD11b expression on classical monocytes</td>
<td>0.19</td>
<td>0.30</td>
<td>0.26</td>
<td>0.04</td>
<td>–0.12</td>
<td>0.47*</td>
</tr>
<tr>
<td>CD11c expression on classical monocytes</td>
<td>–0.21</td>
<td>–0.19</td>
<td>–0.02</td>
<td>–0.03</td>
<td>0.04</td>
<td>0.02</td>
</tr>
<tr>
<td>Intermediate monocytes</td>
<td>0.08</td>
<td>–0.06</td>
<td>0.02</td>
<td>–0.02</td>
<td>0.08</td>
<td>0.11</td>
</tr>
<tr>
<td>Intermediate monocytes, per µl of blood</td>
<td>0.27</td>
<td>0.28</td>
<td>0.35</td>
<td>0.13</td>
<td>0.10</td>
<td>0.28</td>
</tr>
<tr>
<td>CD11b expression on intermediate monocytes</td>
<td>0.20</td>
<td>0.33</td>
<td>0.27</td>
<td>0.04</td>
<td>–0.15</td>
<td>0.49*</td>
</tr>
<tr>
<td>CD11c expression on intermediate monocytes</td>
<td>–0.06</td>
<td>–0.06</td>
<td>–0.02</td>
<td>0.09</td>
<td>0.02</td>
<td>0.03</td>
</tr>
<tr>
<td>Nonclassical monocytes</td>
<td>–0.19</td>
<td>–0.18</td>
<td>–0.53*</td>
<td>–0.07</td>
<td>–0.01</td>
<td>–0.02</td>
</tr>
<tr>
<td>Nonclassical monocytes, per µl of blood</td>
<td>–0.13</td>
<td>–0.04</td>
<td>–0.32</td>
<td>0.01</td>
<td>0.02</td>
<td>0.13</td>
</tr>
<tr>
<td>CD11b expression on nonclassical monocytes</td>
<td>0.10</td>
<td>0.23</td>
<td>0.13</td>
<td>0.04</td>
<td>–0.19</td>
<td>0.52*</td>
</tr>
<tr>
<td>CD11c expression on nonclassical monocytes</td>
<td>–0.09</td>
<td>–0.11</td>
<td>0.08</td>
<td>0.05</td>
<td>0.10</td>
<td>–0.03</td>
</tr>
</tbody>
</table>

Levels of hs-CRP, IL-6, TNF-α, sVCAM-1, PTX-1, and percentage of intermediate monocytes were log-transformed prior to analysis. Because of multiple comparisons, exclusively $p$ values $\leq 0.001$ were inferred significant. * $p \leq 0.001$, ** $p < 0.0001$. Fibr: fibrinogen; BASDAI: Bath Ankylosing Spondylitis Disease Activity Index; hs-CRP: high-sensitivity C-reactive protein; TNF-α: tumor necrosis factor-α; IL: interleukin; sICAM-1: soluble intercellular adhesion molecule-1; sVCAM-1: soluble vascular cell adhesion molecule-1.
The AS patients with a BASDAI score ≥ 4 and < 4 did not differ in terms of traditional risk factors (p > 0.1), levels of sICAM-1 (270 ± 70 vs 262 ± 50 ng/ml, p = 0.7, respectively), sVCAM-1 [541 (491–607) vs 591 (500–658) ng/ml, p = 0.08], hs-CRP [7.02 (2.57–18.20) vs 5.05 (2.95–13.60) mg/l, p = 0.8], PTX-3 [631 (421–739) vs 551 (483–772) pg/ml, p = 0.6], TNF-α [1.77 (1.50–2.20) vs 1.80 (1.58–2.07) pg/ml, p = 0.8], IL-6 [3.09 (2.24–6.98) vs 3.55 (1.80–5.55) pg/ml, p = 0.5], IL-18 [446 ± 137 vs 422 ± 153 pg/ml, p = 0.6], MCP-1 [252 ± 72 vs 264 ± 74 pg/ml, p = 0.6], and size of monocyte subsets (p = 0.4–0.8; Table 4).

The expression of CD11c was similar in patients with AS who had high disease activity, low disease activity, and the controls on classical (1475 ± 638 vs 1522 ± 497 vs 1695 ± 1231 MFI, respectively), intermediate (3381 ± 1025 vs 3468 ± 1040 vs 3983 ± 2612 MFI), and nonclassical monocytes (3620 ± 972 vs 3809 ± 1029 vs 3635 ± 2359 MFI; all intergroup p = 0.3–0.9).

**DISCUSSION**
Our salient finding was a higher number of classical CD14++CD16− blood monocytes and its association with elevated IL-6 levels in DMARD-naive patients with AS. This study is the first, to our knowledge, to report a shift in blood monocyte subpopulations in AS. Additionally, in AS we observed increased sICAM-1, a marker of endothelial activation and an endothelial ligand for monocytic β2-integrin CD11b/CD18. Moreover, on all 3 monocyte subsets the expression of CD11b was decreased in AS.
subjects with low disease activity compared to those with high disease activity, or healthy controls.

The shift in monocyte subsets in AS differed from the previously observed expansion of CD14/CD16− double positive monocytes18,20 or CD14++CD16+ intermediate monocytes19 in RA. This might result from distinct pathogenesis of these 2 forms of inflammatory arthritis. However, we have not studied patients with RA, which limits comparisons of our findings with those reports18,19,20.

That the observed increase in CD14++CD16− monocyte counts in AS correlated to 6-fold elevated IL-6 levels, but not TNF-α or IL-18 concentrations, is compatible with a high secretion of IL-6 by classical monocytes from healthy individuals in response to lipopolysaccharide31,32. Because IL-6 (besides inducing the synthesis of acute-phase reactants) plays a role in the differentiation of Th1733 and Th2 cells34, this relationship may reflect the contribution of a shift in blood monocyte subpopulations to the pathogenesis of AS, reportedly associated with a 10-fold increase in the percentage of Th17 cells among peripheral blood mononuclear cells without in vitro stimulation35. Nevertheless, it is unknown whether classical monocytes have the same properties in patients with AS and healthy subjects.

Previous reports relating monocyte heterogeneity to CV prognosis are inconsistent. A higher frequency of intermediate monocytes but not classical monocytes independently predicted adverse CV outcome in 94 patients receiving dialysis and in 119 nondialysis CKD subjects followed for about 3 and 5 years, respectively13,14. In 951 patients referred for elective coronary angiography, mainly with prevalent coronary artery disease (CAD) or cerebrovascular disease, both classical and intermediate monocytes emerged as univariate predictors of major CV events during a mean followup of 2.6 years, whereas upon multivariate

Figure 2. Elevated CD11b expression on classical (A), intermediate (B), and nonclassical (C) monocytes in subjects with ankylosing spondylitis and a BASDAI score ≥ 4 (n = 21) compared to the remaining patients (n = 26). Data are shown as means ± SE of the mean; p values by Student’s t test. BASDAI: Bath Ankylosing Spondylitis Disease Activity Index score; MFI: mean fluorescence intensity.
adjustment the statistical significance was retained exclu-
sively for intermediate monocytes. Therefore, our findings
might suggest that excessive CV risk in AS might be
independent of a shift in favor of classical monocytes.

On the other hand, classical monocytes, but not other
monocyte subsets, independently predicted ischemic CV
events in 700 randomly selected participants of the cardio-
vascular arm of the Malmö Diet and Cancer study over a
15-year followup. Additionally, in a group of high-risk
patients with stable CAD, the proportion of classical
monocytes was elevated at the expense of nonclassical
monocytes, as in our AS subjects, compared to low-risk and
medium-risk groups delineated on the basis of the number
of traditional risk factors in 80 subjects with CAD. Further,
Urra, et al found a positive association between classical
monocytes (which exhibited the highest platelet-monocyte
interactions) and 3-month mortality in 46 consecutive
patients with stroke.

However, the published observations on the association
between adverse CV outcome and classical CD14++CD16−
monocytes or intermediate CD14++CD16+ monocytes may be
not entirely contradictory because Cros, et al have
provided evidence (by means of whole-genome expression
arrays) that both classical and intermediate human
monocytes resemble murine Ly6C\text{high}Gr\text{+} monocytes,
exhibiting preferential homing to activated endothelium and
atherosclerotic plaques in experimental models of athero-
sclerosis. In addition, preferential transendothelial
migration in response to MCP-1 was demonstrated for
human classical monocytes as compared to the other
monocyte subsets. Moreover, classical monocytes are the
main source of reactive oxygen species known to impair
the activity of nitric oxide, an endogenous antiatherogenic
molecule.

Thus, an increased frequency of classical monocytes in
our subjects with AS who had elevated sICAM-1, an index
of endothelial activation and an endothelial ligand for
monocytic β2-integrin CD11b/CD18, supported by a
relative increase in the expression of CD11b on classical and
intermediate monocytes by over 50% in patients with active
disease compared to those with low disease activity, might
reflect enhanced leukocyte-endothelial interactions and
chronic inflammation within the vascular wall. Because
BASDAI score had been reportedly associated with tradi-
tional risk factors, including metabolic syndrome compo-
nents, the leukocyte-endothelial interactions could
potentiate pro-atherogenic effects of the classical risk
factors in patients with high disease activity. Admittedly,
monocytic CD11b expression did not differ between the
patients with AS who had active disease and the controls;
nevertheless, a decreased CD11b expression on monocytes
of the subjects with AS who had low disease activity could
hypothetically reflect an effective compensatory antiinflam-
matory mechanism that was absent in those with a BASDAI
score ≥ 4. This suggestion remains speculative and requires
further investigation.

On the other hand, the biological significance of
relatively small differences in sICAM-1 levels and the
frequency of classical monocytes remains unclear. Never-
theless, compared to controls, patients with AS exhibited
higher average relative elevations of these variables
(classical monocytes: 469 vs 364 per μl of blood (29%);
sICAM-1: 251 vs 202 ng/ml (25%)) than those previously
reported in participants who later experienced an ischemic
CV event versus the event-free subjects in prospective
studies (classical monocytes: 344 vs 297 per μl of blood
(16%) in the Malmö Diet and Cancer study; sICAM-1: 289 vs 244 ng/ml (18%) in the Atherosclerosis Risk in
Communities study).

It is noteworthy that PTX-3, elevated in patients with AS,
is released locally from cells involved in atherogenesis and
(despite its atheroprotective effects) can be considered a
marker of vascular inflammation. Further, higher PTX-3
levels predicted CV mortality independently of hs-CRP and
traditional risk factors in subjects free of prevalent CV
disease. However, we have provided no direct evidence
supporting the notion of a better ability of PTX-3, adhesion
molecules, or monocyte subsets to predict accelerated
atherogenesis compared to classical risk factors and conven-
tional inflammatory markers in AS. To verify this specu-
lative concept, further studies are warranted, relating these
variables to coexistent atherosclerotic CV disease or
subclinical atherosclerosis in patients with AS.

Several limitations of our study need to be acknowl-
edged. First, the small number of subjects constrains the
conclusions based on our findings. Second, we have
measured cytokines and chemokines in plasma, whereas the
assessment of cytokine generation in response to various
stimuli and the expression of chemokine receptors by
different monocyte subsets would add information to the in
situ plasma levels. Third, we are not able to
exclude the influence of nonsteroidal antiinflammatory
drugs on the activation of both monocytes and endothelial
cells in patients with AS. Fourth, blood monocyte subpop-
lizations might be affected by lipids, and a lower level of
HDL cholesterol was observed in subjects with AS.
However, no lipid variable correlated to the size of any of
the monocyte subsets or their β2-integrin expression in our
study. Finally, although we made every effort to ensure
similar characteristics in both groups, the percentage of men
was higher in AS. Rogacev, et al described higher absolute
counts of classical monocytes in men referred for elective
coronary angiography mainly because of increased numbers
of total monocytes, also present in our data. Nevertheless,
we observed intergroup differences in both relative and
absolute counts of classical monocytes, which argues
against the notion of sex differences in total monocyte
counts as an explanation of our results. In addition, the

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results have not changed after limitation of the analysis to male subjects.

Patients with AS exhibit a shift in blood monocyte subpopulations. Our preliminary cross-sectional findings suggest contribution of immune dysregulation to enhanced monocyte-endothelial interactions in AS, especially in patients with active disease, which possibly can accelerate atherogenesis on a long-term basis. Further longitudinal studies are warranted to determine the relevance of these abnormalities for late CV morbidity.

REFERENCES


