ABSTRACT. Objective. To investigate the relationship between genetic variations of rheumatoid arthritis (RA) susceptibility and joint morbidity.

Methods. We used data from Genetic Analysis Workshop 15. The Illumina linkage panel IV included 5858 single-nucleotide polymorphisms (SNP), with 5744 SNP passing quality control filters. The phenotypic variables analyzed were the level of rheumatoid factor (RF) and score on the Joint Alignment and Motion (JAM) scale. We modified the scale, dividing by RF values relevant to disease severity. Linkage analysis for affected sibling pairs was done using the MERLIN program, and family-based association tests were carried out using PLINK and FBAT software.

Results. We found a high peak (LOD = 3.29; NPL Z = 4.07) near the HLA-DRB1 region on chromosome 6. The linkage at 6p24 at rs1410766 [LOD = 2.66; nonparametric linkage (NPL) Z = 3.23] was statistically significant. Two other regions also showed possible linkage peaks: chromosome 7q30 at rs322812 (LOD = 2.47; NPL Z = 3.39) and chromosome 15p34 at rs347117 (LOD = 1.95; NPL Z = 2.80). For the family-based association study, 7 SNP related to clinical RA severity were detected.

Conclusion. Genetic variations may lead to an enhanced risk of joint damage and increased levels of RF. Further studies are needed to elucidate the roles of other genes involved in RA and to explore whether the clinical signs of RA are associated with particular genetic variations. (First Release Dec 23 2009; J Rheumatol 2010;37:291–5; doi:10.3899/jrheum.090526)

Key Indexing Terms:
- LINKAGE STUDY
- ASSOCIATION STUDY
- RHEUMATOID ARTHRITIS
- JOINT ALIGNMENT AND MOTION SCALE

Rheumatoid arthritis (RA) is a chronic and progressive inflammatory disease occurring in the synovial membrane that can lead to joint deformity and cartilage destruction. Its characteristic feature is the production of autoantibodies. Although there are still no established markers specific to RA, rheumatoid factor (RF), an autoantibody against the Fc portion of immunoglobulin G, has modest specificity and has been used in classification criteria for RA by the American College of Rheumatology.

RF production is the leading indicator of RA severity. However, RF production can also be detected in healthy subjects and is sometimes associated with age. Further, other diseases and environmental exposures can play a role in triggering RF production. In a clinical setting, some patients with RA who have elevated RF levels appear to suffer more severe symptoms, such as progressive loss of joint motion and irreversible joint deformity, while other patients with RA, even those with high RF levels, appear to have relatively minor problems with joint function and disease severity.

Although the progress and severity of RA can be measured by using objective biochemical examinations, given the current state of information, the disparity between these predictors and real symptoms is significant. While this disparity may be explained by phenotypic heterogeneity and individual variability, genetic variants could account for the majority of the difference.

We propose that a certain phenotype, manifested as clinical symptoms representing the degree of RA severity, could be influenced by genetic predisposition. These symptoms include joint deformity and loss of joint motion, 2 prominent features of RA responsible for the majority of unfavorable disease outcomes such as limited daily activity, reduced quality of life, and treatment compliance problems. We investigated genetic variations to RA susceptibility in relation to these symptoms of joint morbidity, using a linkage study and family-based association tests. Although it is unlikely that RF production is responsible for the joint destruction of RA, RF involves the formation of an immune complex that interacts with complement and effector cells to
damage the target cells, potentially promoting joint inflammation and immune dysregulation. Thus, the synovium may convert to lymphoid granulation tissue. The measurement of joint morbidity used for this study was the joint alignment and motion (JAM) scale. To compensate for the disparity between objective markers and clinical symptoms, we modified the scale by dividing JAM scores by RF values to obtain a ratio relevant to disease susceptibility. Patients with lower ratios presented less severe symptoms in terms of joint mobility and/or higher RF levels.

MATERIALS AND METHODS

Study population. We used the resources of the 15th Genetic Analysis Workshop database and the North American Rheumatoid Arthritis Consortium (NARAC) family collection. Families were recruited from 12 NARAC centers in the United States according to the following criteria: (1) at least 1 sibling had documented erosions on hand radiographs, and (2) at least 1 sibling had disease onset between age 18 and 60 years. The dataset included 723 families with 1560 siblings with RA. Additionally, the analysis of RA affection status included data for patients with RA and 349 controls in the same dataset.

The Illumina linkage panel IV was used for analysis of genome-wide single-nucleotide polymorphisms (SNP) in all the families (Illumina, San Diego, CA, USA). The linkage panel contained 5858 SNP markers, and 98.1% of these markers passed quality control filters (minor allele frequency > 5%, SNP call rate > 90%, and Hardy-Weinberg equilibrium p value > 0.01), giving a total yield of 5744 SNP. The results of the analysis of NARAC Caucasian families have been published. Detailed information on the study subjects is available at the NARAC Website.

We used RF levels and the JAM scale as phenotypic measures. In affected siblings, the mean values were 261.6 IU/ml (range 8–6920 IU/ml) for RF levels and 30.8 (range 0–118) on the JAM scale. RF is an IgM antibody directed against the Fc region of IgG and a commonly used serologic marker for the diagnosis of RA. The JAM scale, as a measure of joint motility, represents the severity or limiting aspect of either motion or alignment. It consists of a 5-point scale, with scores indicating the percentage of normal range of motion (ROM) and alignment: a score of 0 represents normal ROM and alignment; 1 = 0–5% decrease in ROM or malalignment; 2 = 6%–25% decrease in ROM or mild malalignment; 3 = 26%–75% decrease in ROM or joint subluxation; and 4 = 76%–100% decrease in ROM, joint fusion, or joint dislocation.

Statistical analysis. As a result of the joint susceptibility to RA, we calculated a JAM-RF ratio. This value was obtained by dividing the JAM scale score by RF level (JAM-RF ratio = JAM scale score/RF).

Linkage analysis for affected sibling pairs was done using the MERLIN program developed by Abecasis, et al. From the linkage analysis, we obtained the nonparametric linkage (NPL) Z score and LOD score using the method of Kong and Cox. In addition, we generated graphic plots along the entire length of each chromosome. Genetic map positions were based on the Rutgers Map Build 36. The empirical p value was estimated for the NPL Z and LOD scores from 10,000 replicates. In each simulation, markers with similar allele frequencies were also generated under the assumption of no linkage.

We performed the transmission disequilibrium test with PLINK software and the family-based association test with FBAT software (-e option) using the additive genetic model.

RESULTS

Table 1 describes the structures of the 723 families included in our study. Most families had 2 affected sibling pairs (N = 637, 88%), and 990 affected sibling pairs were included in this analysis. A total of 1640 subjects, including family members and affected siblings, composed our study population.

Figure 1 displays the multipoint nonparametric LOD score of a genome-wide scan for the JAM-RF ratio over chromosomes 1–22. There was significant evidence of linkage to chromosome 6 at approximately 5 cM near rs197987 and rs412735 on chromosome 6, and the region was included in HLA-DRB1. The high peak (LOD = 9.83) for RA affection status was observed about 5 cM from rs1402405 and rs1410766 (LOD = 2.66; NPL Z = 3.23). The high peak (LOD = 3.29; NPL Z = 4.07) was also observed on chromosome 6 at approximately 5 cM near rs197987 and rs451174. There were 2 suggestive regions, including chromosome 7q30 at rs322812 (LOD = 2.47; NPL Z = 3.39) and chromosome 15p34 at rs347117 (LOD = 1.95; NPL Z = 2.80).

For chromosomes 6, 7, and 15, we performed a complementary analysis for measurements related to RA (Figure 2). Four analyses were conducted on RA status, RF level, JAM score, and JAM-RF ratio. The linkage peak was dependent on RA diagnostic measurements, and the values differed largely among the measurements, while very similar peak regions were detected in chromosomes 6, 7, and 15. The highest peak (LOD = 9.83) for RA affection status was observed about 5 cM from rs1402405 and rs412735 on chromosome 6, and the region was included in HLA-DRB1.

Table 2 shows the results of family-based association tests and corresponding LOD scores for the JAM-RF ratio. There were 7 SNP on chromosomes 6, 7, and 15 with a p value < 0.05, but none of these SNP reached statistical significance following a Bonferroni correction. The SNP on chromosome 6 showed the strongest association. The highest LOD scores among these SNP were for rs2040587 (LOD = 1.76) and rs10099130 (LOD = 1.72), but these SNP were not highly significant in association tests (p < 0.05).

DISCUSSION

We performed a quantitative trait linkage analysis and family-based association test for joint susceptibility to RA, as measured by the JAM-RF ratio, for affected sibling pairs. As expected, the JAM-RF ratio appears to be associated with the HLA-DRB1 region on chromosome 6 (LOD = 3.29; NPL Z = 4.07). This is consistent with reports that found strong evidence for linkage between RA and chromosome 6, which contains the candidate gene HLA-DRB1.

Table 1. Affected sibling pair families.

<table>
<thead>
<tr>
<th>Description</th>
<th>Number</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. of families</td>
<td>723</td>
</tr>
<tr>
<td>No. of families with 2 affected</td>
<td>637</td>
</tr>
<tr>
<td>sibling pairs</td>
<td>66</td>
</tr>
<tr>
<td>No. of families with 3 affected</td>
<td>14</td>
</tr>
<tr>
<td>sibling pairs</td>
<td>5</td>
</tr>
<tr>
<td>No. of families with 4 affected</td>
<td>1</td>
</tr>
<tr>
<td>sibling pairs</td>
<td>1</td>
</tr>
<tr>
<td>No. of affected siblings</td>
<td>1560</td>
</tr>
<tr>
<td>No. of affected sibling pairs</td>
<td>990</td>
</tr>
<tr>
<td>No. of typed individuals</td>
<td>1640</td>
</tr>
</tbody>
</table>
HLA-DRB1 haplotype and disease severity as characterized by positive RF and higher degrees of joint deformity. Based on these results, we propose that HLA-DRB1 is not only the major region of genetic susceptibility to RA but is also a potential region for the joint deformity and loss of joint motion that occurs with RA.

The strongest finding was the significant evidence of linkage to region 6p24 (LOD = 2.66; NPL Z = 3.23). Among the many genes in the interval, an interesting candidate is bone morphogenetic protein 6 (BMP6) gene (MIM 112266), which is located less than 1 cM from the linkage region of 6p24. The BMP are a family of secreted signaling molecules of the transforming growth factor-β superfamily. Several studies have reported a role for BMP6 in skeletal development and bone formation. Rickard, et al. suggested that BMP6 production plays a role in mediating the effect of estrogen on development of bone and cartilage. Cheng, et al. demonstrated a specific role for osteoblasts during mesenchymal stem cell differentiation. Further, work by Lories, et al. supported the hypothesis that BMP6 could be a potent regulator of bone and cartilage repair in arthritis. Based on these results, we postulate that BMP6 may be responsible for helping to regulate joint motion and alignment during the progression of RA.

We observed 2 novel linkage regions on chromosomes 7q30 (LOD = 2.47; NPL Z = 3.39) and 15p34 (LOD = 1.95; NPL Z = 2.80) for JAM-RF ratio. No previous studies have reported linkage of these regions to RA. However, considering that the linkage analysis is dependent on phenotypic variants, in addition to other RA diagnostic measurements that display a similar pattern as shown in Figure 2, these 2 regions may contain candidate genes related to joint alignment and motion susceptibility to RA.

None of the findings in our study of potential linkage regions (chromosomes 6, 7, and 15) were significant after we applied a Bonferroni correction. There were 7 SNP with a p value < 0.05, for which only 3 genes could be found: triadin (TRDN), Src homology 2 domain-containing family member 4 (SHC4), and hypothetical protein FLJ23834. TRDN is a protein that colocalizes with the ryanodine receptor on the sarcoplasmic reticulum in both skeletal muscle and cardiac muscle. The triadin gene is a putative regulator of human muscular pathologies. In contrast, the Shc family proteins serve as phosphotyrosine adaptor molecules in various receptor-mediated signaling pathways. The Shc family of docking proteins, ShcD/Shc4, expressed in adult brain and skeletal muscle, may play a role in mediating downstream signaling of a muscle-specific kinase receptor. FLJ23834 is a putative calcium-dependent cell adhesion protein, and its function has not been established.

We could not confirm a causal link between these genes and clinical RA susceptibility as measured by the JAM-RF ratio. As a result, the association could be due to linkage disequilibrium between a marker and a phenotype. We also failed to detect statistically significant SNP following use of the Bonferroni correction. However, based on the plausible relationship between the novel genes and their potential roles in RA, our finding implies that the genes obtained from our family-based association tests may be responsible for the clinical susceptibility to RA as approximated by the JAM-RF ratio. In addition, other unknown genes could be present near these novel linkage regions.

Of interest is that no gene gave consistent results in both linkage analysis and genome-wide association tests. It is likely that the SNP used in the association analysis did not cover all of the genetic variations in the linkage regions because of the restricted number of SNP in the panel. Another reason may be the lack of power of linkage analysis for complex traits, which could be influenced by multiple genetic, physical, and environmental features. The Type I error of each test may also account for the discordant results between linkage and association analysis.
Research exploring genetic susceptibility to RA has increased, revealing significant evidence for linkage over wide chromosome regions to RA and the presence of RA-specific antibodies, such as RF and anti-citrullinated protein antibody.\(^{11,19,20,32,33}\) The occurrence of RA has also been consistently associated with HLA-linked genes, as well as tumor necrosis factor, HLA-B, PADI4, CTLA4, TRAF1, STAT4, and PTPN2.\(^{34-38}\) Currently, genome projects have resulted not only in identification of genetic factors affecting health and disease but also in improved prediction and care for several diseases. Further, this progress has suggested new directions such as personalized medicine. Although major advances have been made in understanding its potential causes and in generating new therapeutic strategies, RA remains the most common autoimmune disease, with an unknown etiology and, at present, no cure. In light of these considerations, the association of specific gene variations with clinical susceptibility to RA could be used to formulate more effective diagnosis and treatment strategies. In addition, more aggressive treatments and intensive behavioral interventions would contribute to better diagnosis and management of RA.

We found that 2 loci show significant evidence of linkage at 6p21–24, and 2 other regions show suggestive linkage peaks at 7q30 and 15p34. Seven SNP related to joint susceptibility to RA were detected from family-based association tests. These results imply that genetic variations may affect the risk of joint damage and levels of RF. Further studies are needed to elucidate the roles of other genes involved in RA and to explore whether the clinical signs of RA are associated with particular genetic variations.

REFERENCES

Table 2. Results of family-based association tests using the JAM-RF ratio and corresponding LOD scores.

<table>
<thead>
<tr>
<th>Chr</th>
<th>Marker</th>
<th>cM</th>
<th>Region</th>
<th>Gene</th>
<th>Allele</th>
<th>Frequency of Allele</th>
<th>FBAT&lt;sup&gt;17&lt;/sup&gt; p</th>
<th>PLINK&lt;sup&gt;16&lt;/sup&gt; p</th>
<th>MERLIN&lt;sup&gt;13&lt;/sup&gt; LOD</th>
</tr>
</thead>
<tbody>
<tr>
<td>6</td>
<td>rs1009130</td>
<td>68.22</td>
<td>6p11.2</td>
<td>—</td>
<td>A/G</td>
<td>0.26</td>
<td>0.00435</td>
<td>0.00589</td>
<td>1.72</td>
</tr>
<tr>
<td>6</td>
<td>rs2063875</td>
<td>125.65</td>
<td>6p22.31</td>
<td>—</td>
<td>A/T</td>
<td>0.42</td>
<td>0.001183</td>
<td>0.00733</td>
<td>-0.07</td>
</tr>
<tr>
<td>6</td>
<td>rs873460</td>
<td>126.96</td>
<td>6q22.32</td>
<td>TRDN</td>
<td>C/T</td>
<td>0.39</td>
<td>0.00229</td>
<td>0.00155</td>
<td>-0.02</td>
</tr>
<tr>
<td>7</td>
<td>rs41261</td>
<td>115.08</td>
<td>7q22.2</td>
<td>FLJ23834</td>
<td>C/T</td>
<td>0.41</td>
<td>0.03774</td>
<td>0.01274</td>
<td>0.01</td>
</tr>
<tr>
<td>7</td>
<td>rs2040587</td>
<td>123.59</td>
<td>7q13.2</td>
<td>—</td>
<td>C/T</td>
<td>0.37</td>
<td>0.01944</td>
<td>0.01336</td>
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</tr>
<tr>
<td>15</td>
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<td>15q21.1</td>
<td>SHC4</td>
<td>C/T</td>
<td>0.43</td>
<td>0.02245</td>
<td>0.00940</td>
<td>0.93</td>
</tr>
</tbody>
</table>

JAM-RF: joint alignment and motion-rheumatoid factor; Chr: chromosome; TRDN: triadin; FLJ23834: hypothetical protein; SHC4: Scr homology 2 domain-containing family member 4.


