

Transforming Growth Factor β 1 Gene Polymorphism in Patients with Systemic Sclerosis

YOSHIKI SUGIURA, SHOGO BANNO, YOSHIFUJI MATSUMOTO, TAKASHI NIIMI, TAKEO YOSHINOUCI, YOSHIHITO HAYAMI, TAIYO NANIWA, and RYUZO UEDA

ABSTRACT. Objective. To determine whether transforming growth factor β 1 (TGF β 1) gene DNA polymorphism is associated with pathogenesis in the fibrosis of patients with systemic sclerosis (SSc).

Methods. Eighty-seven Japanese patients with SSc including 30 with diffuse type and 57 with limited type together with 110 unrelated controls were investigated. Pulmonary fibrosis was determined in 34 SSc patients using high-resolution chest computed tomography. TGF β 1 genetic polymorphisms were analyzed in 2 loci; T869C (Leu10Pro) in codon 10 at exon 1, and C-509T in the promoter region using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP).

Results. Neither the genotype of T/C polymorphism in T869C nor C/T polymorphism in C-509T revealed any difference in distribution between SSc and controls. In the group of SSc patients with pulmonary fibrosis, a weak but significantly high frequency ($p = 0.05$) of TC+CC (the presence of C allele) in T869C, and CT+TT (the presence of T allele) in C-509T was found. Compared with controls, the pulmonary fibrosis group showed no difference in the highly frequent alleles.

Conclusion. Our results suggest that TGF β 1 polymorphisms do not play a role in the pathogenesis of SSc, even though there remains the possibility of a risk factor for genetic susceptibility to pulmonary fibrosis. (J Rheumatol 2003;30:1520-3)

Key Indexing Terms:

TRANSFORMING GROWTH FACTOR DNAPOLYMORPHISM SYSTEMIC SCLEROSIS

Systemic sclerosis (SSc) is a multisystem connective tissue disease characterized by cutaneous and visceral fibrosis, prominent small vessel damage, and an autoimmune phenomenon. Several cytokines and chemokines are known to induce the secretion of collagen and the formation of fibrosis in SSc. Transforming growth factor β (TGF β) is a multifunctional cytokine which controls cell differentiation and proliferation. In mammals, there are 3 isoforms, TGF β 1, β 2 and β 3 that possess nearly identical biological properties. The overexpression of TGF β 1 is increased in patients with idiopathic pulmonary fibrosis and autoimmune hepatic fibrosis, as well as in patients with SSc. Immunohistological localization of TGF β 1 expression was detected in the alveolar epithelium and interstitium in the lungs of patients with SSc¹. Using skin biopsies from patients with an inflammatory and sclerotic stage of SSc, mRNA for TGF β 1, 2 and 3 were detected in inflammatory skin areas, but not in sclerotic or healthy skin². In addition, the blockade of TGF β

signaling with TGF β antibodies or a TGF β 1 antisense oligonucleotide inhibited the increased mRNA expression in SSc fibroblasts³. TGF β 1 was predicted to be one of the key cytokines in the pathogenesis of SSc.

The genetic polymorphisms of TGF β 1 regulate its expression, and may have a role in predisposing patients to fibrotic disease, especially SSc.

In a UK Caucasian population study by Susol, *et al*⁴, several microsatellite markers including TGF β 1, 2 and 3 and a tissue inhibitor of metalloproteinase-1 (TIMP1) were studied. They demonstrated that there was no association between microsatellite markers for TGF β 1 and SSc. However, another recent report showed a positive association in a UK population⁵. The SSc patients in that study had a higher frequency of C allele at codon 10 than controls. Thus, the association of TGF β 1 polymorphism was ambiguous. We investigated whether TGF β 1 DNA polymorphism is linked to the pathogenesis of fibrosis in SSc patients.

MATERIALS AND METHODS

Patients. We studied 87 Japanese patients with SSc, as well as 110 Japanese controls in the same geographic area. All patients fulfilled the criteria of the American College of Rheumatology for SSc. All patients and controls provided written informed consent. Thirty SSc patients had the diffuse type, and the other 57 had the limited type classified as defined by Leroy, *et al*⁶. Pulmonary fibrosis was seen using high-resolution chest computed tomography (CT) scans in 34 SSc patients. No patients showed clinical evidence of renal failure suggesting renal sclerosis.

From the Department of Internal Medicine and Molecular Science, Nagoya City University Graduate School of Medical Science, Nagoya, Japan.

Y. Sugiura, MD; S. Banno, MD; Y. Matsumoto, MD; T. Niimi, MD; T. Yoshinouchi, MD; Y. Hayami, MD; T. Naniwa, MD; R. Ueda MD, Professor.

Address reprint requests to Dr. S. Banno, Department of Internal Medicine and Molecular Science, Nagoya City University Graduate School of Medical Science, Kawasumi 1, Mizuho-ku, Nagoya-city, Aichi 467-8601, Japan. E-mail: sbannos@med.nagoya-cu.ac.jp

Submitted May 21, 2002; revision accepted December 11, 2002.

Genotype analysis. Genomic DNA from peripheral blood was isolated. We analyzed 2 TGF β 1 polymorphisms: T869C (Leu10Pro) in codon 10 (exon1) in the signal peptide sequence region, and C-509T in the promoter region. The genotypes of these 2 loci were identified using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP), as described⁷ (Figure 1). Genotype distributions in SSc patients and controls were analyzed with Fisher's exact test. A p value < 0.05 was considered statistically significant.

RESULTS

There was no significant difference in the distribution among SSc patients and controls of either T869C or C-509T. When comparing TC+CC (the presence of C allele) with TT homozygous genotype in T869C, we found no significant difference between them (Table 1). Moreover, there was no difference between CT+TT (the presence of T allele) and CC homozygous in C-509T.

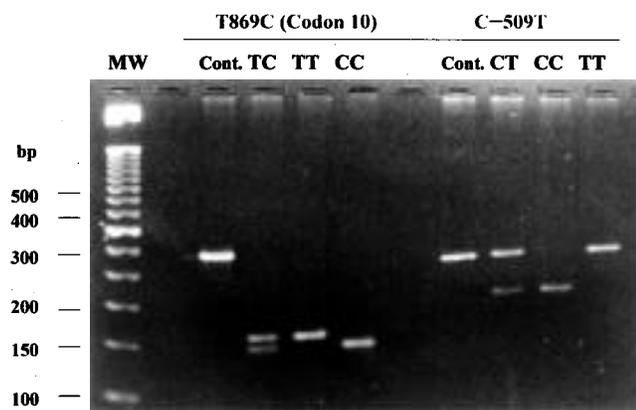


Figure 1. Ethidium bromide-stained 4% agarose gel showing the representative determination of TGF β 1 polymorphism of T869C in codon 10, and C-509T in the promoter region using PCR-RFLP. After digestion of PCR products (294 bp) by MspA1 in T869C (codon 10), T allele and C allele were characterized by a 161 bp and 149 bp fragment, respectively. After digestion of PCR products (265 bp) by Bsu36 in C-509T, the C allele was represented by a 195 bp fragment, and undigested T allele was shown at 265 bp. Three genotypes in T869C (codon 10) were determined, i.e., homozygous for alleles TT and CC, and heterozygous for TC. The polymorphism in C-509T was also determined, i.e., homozygous for the alleles CC and TT, and heterozygous for CT. MW: 50 bp ladder of molecular size markers; Cont.: undigested amplicon.

Table 1. Distribution of TGF β 1 polymorphism at T869C and C-509T in patients with systemic sclerosis (SSc) and controls.

	n	Age Mean \pm SD	Women/Men	TT n (%)	TC n (%)	CC n (%)	TT n (%)	TC+CC n (%)
T869C								
Controls	110	44.5 \pm 18.9	68/42	24 (21.8)	53 (48.2)	33 (30.0)	24 (21.8)	86 (78.2)
SSc	87	56.0 \pm 11.7	74/13	23 (26.4)	38 (43.7)	26 (29.9)	23 (26.4)	64 (73.6)
	n	Age Mean \pm SD	Women/Men	CC	CT	TT	CC	CT+CT
C-509T								
Controls	110	44.5 \pm 18.9	68/42	23 (20.9)	54 (49.1)	33 (30.0)	23 (20.9)	87 (79.1)
SSc	87	56.0 \pm 11.7	74/13	23 (26.4)	38 (43.7)	26 (29.9)	23 (26.4)	64 (73.6)

The distributions of these polymorphisms according to the clinical characteristics of SSc patients are summarized in Table 2. We found no difference in the distribution of either the T869C or C-509T polymorphism between the diffuse and limited type of SSc. However, the TC+CC genotype (the presence of C allele) in T869C, and CT+TT (the presence of T allele) in C-509T was more frequently found in the diffuse type, but this trend was not significant.

Among 87 SSc patients, 34 patients also had pulmonary fibrosis. A significantly higher proportion of polymorphisms in T869C was recognized among SSc patients with pulmonary fibrosis carrying the TC+CC genotype (the presence of C allele) compared to the TT homozygous genotype (the absence of C allele) (p = 0.05). Similarly, the CT+TT genotype (the presence of T allele) in C-509T polymorphism with pulmonary fibrosis was significantly more frequent than the CC genotype (the absence of T allele) (p = 0.05). However, neither the presence of C allele in T869C or of T allele in C-509T revealed any significant difference compared to controls.

DISCUSSION

TGF β 1 has been identified as containing 7 polymorphisms: 3 in the promoter regions at positions -988, -800, and -509; one at position +72 in a non-translated region; 2 in the signal sequence at +869 (Leu10Pro) and +915 (Arg25Pro); and one in the coding region for the precursor part not present in the active form, Ile263Thr. The promoter polymorphism, C-509T, has been recognized in linkage disequilibrium with T869C^{8,9}. We observed that 192 (98.5%) of 195 subjects including SSc and controls showed a linkage between the T869C and T-509C genotypes. Two polymorphisms between -509 and -800 also showed the linkage disequilibrium⁹. In other regions at -988 and codon 263, no difference in the genetic alteration was observed⁹. Another signal sequence at codon 25, G915C, was shown to have a significant association with myocardial infarction, hypertension¹⁰, and fibrotic lung⁸ in Caucasian populations. However, we did not detect this genetic alteration of codon 25. In the Japanese population described previously¹¹, nearly all indi-

Table 2. TGFβ1 polymorphism between diffuse and limited SSc, or between presence or absence of pulmonary fibrosis.

	n	Age Mean ± SD	Women/Men	TT (%)	TC (%)	CC (%)	TT(%)	TC+CC (%)
T869C								
Diffuse SSc	30	55.3 ± 13.1	22/8	6 (20.0)	14 (46.7)	10 (33.3)	6 (20.0)	24 (80.0)
Limited SSc	57	56.3 ± 11.0	52/5	17 (29.9)	24 (42.1)	16 (28.0)	17 (29.8)	40 (70.2)
Pulmonary fibrosis								
Presence	34	55.6 ± 12.3	27/7	5 (14.7)	18 (52.9)	11 (32.4)	5 (14.7)	29 (85.3) }*
Absence	53	56.2 ± 11.5	47/6	18 (34.0)	20 (32.7)	15 (28.3)	18 (34.0)	35 (66.0) }
	n	Age Mean ± SD	Women/Men	CC (%)	CT(%)	TT(%)	CC (%)	CT+CT (%)
C-509T								
Diffuse SSc	30	55.3 ± 13.1	22/8	6 (20.0)	13 (43.3)	11 (36.7)	6 (20.0)	24 (80.0)
Limited SSc	57	56.3 ± 11.0	52/5	17 (29.8)	25 (43.9)	15 (26.3)	17 (29.8)	40 (70.2)
Pulmonary fibrosis								
Presence	34	55.6 ± 12.3	27/7	5 (14.7)	17 (50.0)	12 (35.3)	5 (14.7)	29 (85.3) }*
Absence	53	56.2 ± 11.5	47/6	18 (34.0)	21 (39.6)	14 (26.4)	18 (34.0)	35 (66.0) }

* p = 0.05

viduals were detected as GG genotype, so they could not be used for the polymorphism study.

We failed to detect a difference in the distribution of TGFβ1 polymorphism at T869C and C-509T between SSc patients and controls in a Japanese cohort. These negative associations of TGFβ1 polymorphism confirm 2 previous investigations^{4,12}. One was a small population study by Zhou, *et al*¹² showing a negative association of TGFβ1, in the Oklahoma Choctaw native Americans, exhibiting a high frequency of SSc. This study has been used for several genetic analyses of SSc. The second was a large population study in 191 SSc Caucasian patients by Susol, *et al*⁴. That study found associations indicating a possible role for TGFβ3, β2, and TIMP1 in genetic susceptibility to SSc and for TGFβ3 in determining the degree of cutaneous fibrosis. Other than the above-reported negative associations of TGFβ1, a positive association was found by Crilly, *et al*⁵ in 89 SSc patients in a UK population. In their study, there was no difference between diffuse and limited SSc, similar to our results.

T869C at codon 10 and C-509T promoter polymorphism were thought to be associated with TGFβ1 protein expression. In the Japanese population, the serum concentration of TGFβ1 was significantly higher in the C allele than in the T allele at codon 10 in a study of genetic susceptibility to osteoporosis¹³ or myocardial infarction¹¹. The C allele at codon 10 was more frequent in black than white Americans with hypertension, and this allele was associated with higher levels of TGFβ1 protein and mRNA¹⁴. Otherwise, the C allele at codon 10 was associated with lower TGFβ1 protein expression¹⁵ in patients with cystic fibrosis. Although these studies have limitations regarding the association with TGFβ1 protein expression, if the hypothesis is correct that the C allele at codon 10 and the T allele at C-509T are high

producer alleles, our results are not incompatible with the idea that pulmonary fibrosis is found at significantly higher frequencies in these alleles. Such alleles may be a possible risk factor for genetic susceptibility following overexpression of TGFβ protein in SSc patients with pulmonary fibrosis. In contrast to the findings by Crilly *et al*⁵ in the UK population, the negative associations in our Japanese cohort could be due to the heterogeneity of SSc, or perhaps to some racial difference. Otherwise, the TGFβ3 gene may be the more likely candidate for genetic susceptibility to SSc than the TGFβ1 gene⁴. Further studies should be undertaken on a much larger cohort with accurate characterization of clinical features including skin, lung, renal, or gastro-intestinal fibrosis in SSc patients.

REFERENCES

1. Corrin B, Butcher D, McNulty BJ, Dubois RM, Laurent GJ, Harrison NK. Immunohistochemical localization of transforming growth factor-β1 in the lungs of patients with systemic sclerosis, cryptogenic fibrosing alveolitis and other lung disorders. *Histopathology* 1994;24:145-50.
2. Querfeld C, Eckes B, Huerkamp C, Krieg T, Sollberg S. Expression of TGF-β1, -β2 and -β3 in localized and systemic scleroderma. *J Dermatol Sci* 1999;21:13-22.
3. Ihn H, Yamane K, Kubo M, Tamaki K. Blockade of endogenous transforming growth factor β signaling prevents up-regulated collagen synthesis in scleroderma fibroblasts. *Arthritis Rheum* 2001;44:474-80.
4. Susol E, Rands AL, Herrick A, et al. Association of markers for TGFβ3, TGFβ2, and TIMP1 with systemic sclerosis. *Rheumatology* 2000;39:1332-6.
5. Crilly A, Hamilton J, Clark CJ, Jardine A, Madhok R. Analysis of transforming growth factor β1 gene polymorphisms in patients with systemic sclerosis. *Ann Rheum Dis* 2002;61:678-81.
6. LeRoy C, Medsger Jr TA. Criteria for the classification of early systemic sclerosis. *J Rheumatol* 2001;28:1573-6.
7. Wood NAP, Thomson SC, Smith RM, Bidwell JL. Identification of human TGF-β1 signal (leader) sequence polymorphisms by

- PCR-RFLP. *J Immunol Methods* 2000;234:117-22.
8. Awad MR, El-Gamel Ahmed, Hasleton P, Turner DM, Sinnott PJ, Hutchinson IV. Genotypic variation in the transforming growth factor- β 1 gene. Association with transforming growth factor- β 1 production, fibrotic lung disease, and graft fibrosis after lung transplantation. *Transplantation* 1998;8:1014-20.
 9. Grainger DJ, Heathcote K, Chiano M. Genetic control of the circulating concentration of transforming growth factor type β 1. *Hum Mol Genetics* 1999;8:93-7.
 10. Cambien F, Ricard S, Troesch A, et al. Polymorphisms of the transforming growth factor- β 1 gene in relation to myocardial infarction and blood pressure. *Hypertension* 1996;28:881-7.
 11. Yokota M, Ichihara S, Lin T-L, Nakashima N, Yamada Y. Association of a T29 \rightarrow C polymorphism of the transforming growth factor- β 1 gene with genetic susceptibility to myocardial infarction in Japanese. *Circulation* 2000;101:2783-7.
 12. Zhou X, Tan FK, Stivers DN, Arnett FC. Microsatellites and intragenic polymorphisms of transforming growth factor β and platelet-derived growth factor and their receptor genes in native Americans with systemic sclerosis (scleroderma). *Arthritis Rheum* 2000;43:1068-73.
 13. Yamada Y, Miyauchi A, Takagi Y, Tanaka M, Mizuno M, Harada A. Association of C-509 \rightarrow T polymorphism, alone or in combination with the T869 \rightarrow C polymorphism, of the transforming growth factor- β 1 gene with bone mineral density and genetic susceptibility to osteoporosis in Japanese women. *J Mol Med* 2001;79:149-56.
 14. Suthanthiran M, Li B, Song JO, et al. Transforming growth factor- β 1 hyperexpression in African-American hypertensives: A novel mediator of hypertension and/or target organ damage. *Proc Natl Acad Sci USA* 2000;97:3479-88.
 15. Arkwright PD, Laurie S, Super M, Pravica V, Schwarz MJ, Webb AK, Hutchinson IV. TGF- β 1 genotype and accelerated decline in lung function of patients with cystic fibrosis. *Thorax* 2000; 55:459-62.