Effects of FTY720 in MRL-lpr/lpr Mice: Therapeutic Potential in Systemic Lupus Erythematous

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ABSTRACT

Objective. To examine the effects of a novel immunosuppressant, FTY720, on hematolymphoid cells and the clinical course of MRL-lpr/lpr (MRL/lpr) mice genetically predisposed to systemic lupus erythematous (SLE).

Methods. Apoptosis of hematolymphoid cells was determined in vitro by FACScan after staining with propidium iodide or merocyanine 540. From 4 months of age, 15 female MRL/lpr mice received oral administration of 2 mg/kg each of FTY720, methylprednisolone (mPSL), or vehicle, 3 times per week. Therapeutic efficacy was evaluated by levels of anti-dsDNA antibodies in serum and the survival rate. In parallel, T cell proliferation and secretion of interleukin 2 (IL-2) induced by anti-CD3, phenotypes of the spleen, lymph node and bone marrow cells, as well as immunohistochemistry of the kidney, were examined in vitro.

Results. FTY720 at 2 µM induced apoptosis in more than 70% of double negative (CD4–/CD8–) T cells from the spleen of MRL/lpr mice in vitro. Oral FTY720 was tolerated well with no apparent side effects. FTY720 treated and control mice gained weight at an identical pace through to 9 months of age. FTY720 significantly suppressed the production of anti-dsDNA antibodies (FTY720 vs control: 1739 ± 898 U/ml vs 410 ± 356 U/ml at 8 months of age; p < 0.05) and reduced the deposition of IgG in glomeruli compared to control animals. At 9 months of age, the survival rate in the FTY720 treated mice was 86.9% compared to 33.0% in controls (p < 0.01). FTY720 decreased the number of double negative T cells from the spleen and lymph nodes in vivo, and increased T cell proliferation and IL-2 secretion induced by anti-CD3 stimulation in vitro.

Conclusion. FTY720 suppressed the development of autoimmunity and prolonged the lifespan of female MRL/lpr mice. Suppression of autoimmunity, at least in part, may have resulted from an apoptotic potential of FTY720. Hence, it could be useful for primary or adjunctive therapy of human SLE. (J Rheumatol 2002;29:707–16)

Key Indexing Terms:
FTY720
LUPUS
APOTOPSIS

Although the pathogenesis of most autoimmune diseases is yet to be determined, increasing lines of evidence have indicated a role of apoptosis in the development of autoimmune diseases. In several animal models of spontaneous autoimmune diseases, genetic defects causing an abnormal resistance of peripheral T cells to apoptosis have been identified. This applies both to models of systemic lupus erythematous (SLE) (strains homozygous for the lpr or gld mutation) and to an animal model of type I diabetes mellitus (nonobese diabetic (NOD) mouse). Thus, the lpr mutation renders the Fas gene inoperative; the gld mutation affects the Fas ligand; and one of the diabetes susceptible loci of the NOD strain maps to the bcl-2 gene. The induction of apoptosis in these mouse models of autoimmunity, therefore, would ameliorate autoimmune manifestations in them.

This view is supported by corticosteroid and cyclophosphamide, widely used for the treatment of SLE, which exert immunosuppressive activities, in part, by means of inducing apoptosis in lymphocytes. However, their utility is restricted by their undesirable side effects including infection, premature cardiovascular mortality, infertility, and neoplasia. Immunosuppressive drugs with a high desired/adverse reaction ratio and with a higher specificity for effector lymphocytes causing autoimmunity are awaited.

A potential candidate satisfying such criteria might be FTY720 (2-amino-2-(2-[4-octyl-phenyl]ethyl)-1,3-propanediol hydrochloride), a novel immunosuppressant resembling sphingosine. In this regard, it differs from...
conventional immunosuppressants such as FK506, cyclosporin A, and rapamycin. FTY720 administered per os prolongs the survival of skin, cardiac, and liver allografts in rats, as well as that of renal allografts in dogs. It also prevents the development of graft-versus-host disease in rats, with little apparent toxicity. Although the precise molecular mechanism of the immunosuppressive activity of FTY720 has not been fully elucidated, studies have suggested that FTY720 may induce apoptosis in lymphocytes through activation of phospholipase C or some protein kinases, and accelerate homing of lymphocytes to lymphoid tissue by altering the expression of CD62L.

Mice of the MRL-lpr/lpr (MRL/lpr) strain spontaneously develop an autoimmune disease resembling human SLE. The disease is characterized by immune complex mediated glomerulonephritis, enlargement of the spleen and lymph nodes, and production of various autoantibodies such as anti-dsDNA antibodies and rheumatoid factors. The MRL/lpr strain exhibits diminished Fas mediated lymphocyte apoptosis, resulting in accumulation of CD3+CD4−CD8− (double negative) T cells. These abnormally proliferating T cells do not respond normally to anti-CD3 antibody or mitogens such as concanavalin A, resulting in reduced interleukin 2 (IL-2) production. Anti-CD3 antibody or mitogens such as concanavalin A, abnormally proliferating T cells do not respond normally to cyto apoptosis, resulting in accumulation of lymphocytes through activation of phospholipase C or some molecular mechanism of the immunosuppressive activity of FTY720.

Flow cytometry. Methods for immunofluorescent staining were as described. Single cell suspensions of the spleen, lymph node, and bone marrow were stained with fluorochrome conjugated Mab and analyzed in a FACSscan flow cytometer (Becton Dickinson Immunocytometry Systems, San Jose, CA, USA) using Lysis II software (Becton Dickinson). Combinations of FITC-anti-CD3 and PE-anti-CD4 + CD8 Mab were used for dual surface staining to detect CD3+ double negative T cells. Anti-IgM Mab was used to identify mature B cells in the spleen and bone marrow from MRL/lpr mice. To avoid nonspecific staining, the cells were preincubated with a mixture of unconjugated rat anti-mouse CD32 and anti-CD16 antibodies (PharMingen).

Assessment of apoptosis. Experiments were performed in triplicate in 24 well flat bottom plates at a cell concentration of 2 × 10^6 cells/ml. Apoptosis was measured in hemolympidoid cells cultured in vitro for 0, 2, 4, 8, 12, and 24 h. The number of viable cells was determined by staining with erythrosin B at each time point. Apoptosis was confirmed by flow cytometric analysis of cell nuclei stained with propidium iodide (PI) and oligonucleosomal fragmentation of genomic DNA visualized on agarose gels. Briefly, after stimulation, the cells were washed twice in phosphate buffered saline (PBS) containing 0.1% (v/v) NaN3, and 1 ml of 0.2 ml of 100 mg/ml PI (Sigma). The cells were incubated at room temperature for 20 min in the dark and then analyzed by FACSscan. Apoptotic cells appeared in the < 2 N DNA peak. They were distinguished from necrotic cells by analyzing the light scatter profile. Necrotic cells showed a large decline in forward-angle light scatter and side scatter, while apoptotic cells showed a smaller decline in them. Necrotic cell fragments and cell debris were excluded by an appropriate gate, and data were recorded on a linear scale. At least 10,000 cells were evaluated per sample in most experiments. Apoptotic cells in certain T cell subsets were determined by increased binding with a lipophilic fluorescent dye [merocyanine 540 (MC540); Sigma], as well as by decreased forward scatter by flow cytometry. Staining with MC540 was performed as described with a minor modification. Splenic T cells from MRL/lpr mice were cultured for 12 h. After the cells were surface stained with FITC conjugated anti-CD4 + CD8 Mab, they were resuspended in 100 ml of PBS supplemented with 2% FCS. Then MC540 (5 mg/ml) was added to the suspension, and the cells were incubated at room temperature for 10 min. Stained cells were analyzed immediately by FACSscan.

Detection of DNA fragmentation in agarose gels. Splenic T cells (1 × 10^6) from MRL/lpr mice cultured with 2 µM dexamethasone for 4 h were used as a positive control for DNA fragmentation. Splenic T cells (1 × 10^6) from MRL/lpr mice were stimulated as indicated for 4 h and harvested. They were centrifuged and washed once with cold PBS. The cell pellet was resuspended in 0.6 ml of lysate buffer containing 10 mM Tris-HCl, 20 mM EDTA, and 0.25% Triton X-100 (pH 8.0) on ice. After 5 min, the lysate was centrifuged at 15,000 × g for 10 min at 4˚C in an Eppendorf microfuge tube. The supernatant (containing RNA and fragmented DNA, but not intact chromatin) was then placed in a separate microfuge tube with EDTA (12.5 mM) and DNase-free RNase (30 mg/ml) at 37˚C. After 15 min,
Experimental design for in vivo treatment. Mice were randomly divided into 3 groups, each of which was treated with FTY720, methylprednisolone (mPSL), or none (controls). Fifteen MRL/lpr mice at 4 months of age were given per os either 2.0 mg/kg of FTY720 or mPSL 3 times per week, using a curved blunt-end needle inserted into the throat. Control mice were given 0.5 ml of saline. This dose of 2.0 mg/kg FTY720 was adopted from the published report. All experiments were conducted in accord with the guidelines of the Animal Ethics Committee of Chichi Medical School.

Detection of anti-dsDNA antibodies. An anti-dsDNA ELISA kit (MBL, Nagoya, Japan) was used to quantify IgG antibodies to double stranded DNA in sera from individual mice. The assay was performed according to the manufacturer’s instructions and the quantity of anti-dsDNA antibodies was estimated by comparison against the standard curve.

Proliferation of splenic T cells. The method described by Warner, et al was followed for estimating the proliferation of spleen cells in vitro. Splenic T cells obtained through a nylon wool column were cultured at 5 x 10^5 cells/well in flat bottom microtiter plates that had been precoated with 5 ng/ml purified anti-CD3 antibodies. Three days later the cells were pulsed with [3H]-thymidine (1 mCi/well) (NEN, Boston, MA, USA) 5 mg/ml purified anti-CD3 antibodies. Three days later the cells were rinsed with 70% ethanol, air dried, and dissolved in TE buffer (10 mM Tris-HCl, pH 7.5, and 1 mM EDTA). The samples were electrophoresed on a 1.7% agarose gel with TAE buffer (40 mM Tris-acetate, 1 mM EDTA, pH 8.0). DNA was then visualized by ethidium bromide staining. A 1 kb DNA ladder (BRL) obtained after digestion using Hind I was employed as standard size fragments.

RESULTS

Effects of FTY720 on apoptosis in normal BALB/c mice in vitro. Apoptosis was measured in the thymocytes, spleen cells, and bone marrow cells from normal 6-week-old BALB/c mice that had been cultured for 0, 2, 4, 8, 12, and 24 h. Little apoptosis, if any, was seen at time 0 in any of the cell populations, as determined by PI staining (Figure 1). Since the lowest concentration of FTY720 to induce maximal apoptosis of thymocytes cultured for 8–24 h was 2 μM (data not shown), subsequent experiments were performed at this concentration. FTY720 induced a significant increase in apoptosis of the thymocytes (Figure 1A), but not of the spleen cells (Figure 1B). In agreement with published data, DEX increased apoptosis in both the thymocytes and spleen cells to a much higher extent than FTY720 at the same concentrations. In sharp contrast, FTY720 induced apoptosis of the bone marrow cells to a greater degree than DEX (Figure 1C). Thus, the apoptoses of hematolymphoid cells with FTY720 and DEX could be induced by a different mechanism.

Effects of FTY720 on in vitro apoptosis of spleen cells from MRL mice. In MRL/+ mice of the same age, the results were essentially similar to those seen in BALB/c mice (Figure 2A), while in age matched MRL/lpr mice, FTY720 induced apoptosis in about 70% of the spleen cells and to the same extent as DEX (Figure 2B). These results suggested that FTY720 may increase the apoptotic cell death in the CD3+CD4–CD8– double negative T cell population predominant in MRL/lpr mice by 6 months after birth. To confirm this, splenic T cells from 6-month-old MRL/lpr mice were stained with MC540 and a mixture of FITC conjugated anti-CD4 and anti-CD8 Mab. DEX increased apoptosis in both the double negative T cells and CD4+ or CD8+ single positive T cells after 12 h in culture, while FTY720 selectively induced apoptosis in the double negative T cells (Figure 3).

Induction of DNA fragmentation by FTY720 in splenic T cells from MRL/lpr mice. To evaluate whether FTY720 induced DNA fragmentation characteristic of apoptosis in short term cultures, the genomic DNA from splenic T cells was separated by gel electrophoresis. For this purpose, splenic T cells were incubated for 4 h, then harvested and the genomic DNA was obtained. A DNA fragmentation pattern serving as control was of splenic T cells from MRL/lpr mice undergoing apoptosis after incubation in vitro with 2 μM dexamethasone (Figure 4, Lane 1). Lanes 2 and 3 show the DNA fragmentation pattern in splenic T cells from MRL/+ mice incubated in medium and 2 μM FTY720, respectively, with no DNA fragmentation. The splenic T cells from MRL/lpr mice cultured in medium had little or no DNA fragmentation (Lane 4). In contrast, DNA in the splenic T cells from MRL/lpr mice incubated with 2 μM FTY720 showed a fragmentation pattern characteristic of apoptosis (Lane 5). A Hind I digest of DNA is shown for comparison (Lane 6).

Effects of FTY720 on the clinical course of lupus in mice. Survival. Sixteen-week-old female MRL/lpr mice were divided randomly into 3 groups of 15 animals. Each group received orally either 2 mg/kg of FTY720, 2 mg/kg of mPSL, or vehicle alone, 3 times a week, and was observed daily. The FTY720 treated mice gained weight at a pace identical to that of control mice through to 36 weeks of age (Table 1). As shown in Figure 5, FTY720 prolonged the survival of mice at 8, 9, and 10 months of age significantly longer than controls (at 9 mo, 86.7% of FTY720 treatment vs 33.0% of control; p < 0.01), to an extent comparable to that with mPSL.
Kidney disease. Frozen kidney sections from 10-month-old mice treated with FTY720 or mPSL were stained with FITC conjugated anti-mouse IgG. As shown in Figure 6, untreated control mice displayed heavy deposits of IgG in the glomeruli, while the majority of FTY720 treated mice showed a marked decrease in IgG deposition. Essentially similar results were obtained when staining for C1q and C3 in the glomeruli (data not shown).

Production of anti-DNA antibodies. Serum IgG anti-dsDNA antibodies were measured in FTY720 treated, mPSL treated, and control MRL/lpr mice at 6 and 8 months of age. There were no significant differences in the levels of serum anti-dsDNA antibodies among the 3 groups at the start. At the age of 8 months, the mean titer of anti-dsDNA antibodies in the FTY720 treated mice decreased significantly in comparison with controls. The extent of the decrease in the levels
of anti-dsDNA antibodies, however, was higher with mPSL than FTY720 (Figure 7).

**Lymphoid proliferation and cell populations in the spleen.** In a parallel experiment, 16-week-old female MRL/lpr mice were treated with FTY720, mPSL, or vehicle alone for 16 weeks and then examined. Six control animals and one mPSL treated mouse died before the completion of the experiments, probably due to renal failure, and therefore were excluded from further analyses. There were significant reductions in weight of the cervical lymph nodes and spleen, as well as in the cell number in the spleen from FTY720 and mPSL treated mice, in comparison with controls (Table 1). Spleen cells freshly isolated from MRL/lpr mice were subjected to FACScan analysis. As shown in Figure 8, oral FTY720 markedly reduced the frequency of double negative T cells to an extent comparable with that of mPSL. Table 1 summarizes changes in frequency and number of T, B, and myeloid cells in the spleen and bone marrow. Both frequency and number of double negative T cells in the spleen were significantly decreased by FTY720. Although the frequency of B cells was unchanged among the groups, the total number of B cells in the spleen was significantly reduced in both FTY720 and mPSL treated mice in comparison with controls. Since FTY720 increased apoptosis in the bone marrow cells in vitro, the effects of FTY720 on the bone marrow in vivo were examined. The number of Gr-1+ myeloid cells in the bone marrow was not significantly reduced in the FTY720 treated mice compared to controls.

**Proliferation of spleen cells and secretion of IL-2 in vitro.** FTY720 significantly increased the responsiveness of splenic T cells to the stimulation with anti-CD3 (5 mg/ml) in comparison with control mice (Figure 9A). This increase in responsiveness by FTY720 reached almost the same level as that of splenic T cells from MRL/+ mice stimulated with anti-CD3 antibody. The ability to produce IL-2, in response to stimulation with anti-CD3, increased significantly in the FTY720 treated mice, in parallel with an increase in T cell proliferative responsiveness (Figure 9B).

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**Figure 3.** Identification of apoptotic cells in vitro in single positive or double negative splenic T cells from 6-month-old MRL/lpr mice. Splenic T cells from female 6-month-old MRL/lpr mice were cultured 12 h in the absence (untreated) or presence of 2 µM FTY720 or 2 µM DEX. The cells were recovered, stained with a combination of FITC conjugated anti-CD4 and anti-CD8 Mab, as well as MC540, and analyzed by 2 color flow cytometry. Percentage of apoptotic cells was determined as MC540 high in double negative or single positive T cells. Representative results of 5 experiments are shown here.

**Figure 4.** Agarose gel analysis of DNA extracted from splenic T cells from MRL/lpr mice cultured for 4 h. Splenic T cells were cultured 4 h and DNA was extracted and analyzed by gel electrophoresis. Lane 1: Splenic T cells from MRL/lpr mice cultured with 2 µM dexamethasone. Lane 2: Splenic T cells from MRL/+ mice cultured with medium alone. Lane 3: Splenic T cells from MRL/+ mice cultured with 2 µM FTY720. Lane 4: Splenic T cells from MRL/lpr mice cultured with medium alone. Lane 5: MRL/lpr mice cultured with 2 µM FTY720. Lane 6: Control DNA digested with Hinf I.
DISCUSSION

We have shown that FTY720 is efficacious for the treatment of experimental SLE in MRL/lpr mice. Treatment with FTY720 was tolerated well by MRL/lpr mice with no adverse effects such as bone marrow toxicity (Table 1) or hyperglycemia seen in mPSL treated animals (data not shown). Treatment with FTY720 can reduce adenopathy and splenomegaly, possibly through the induction of lymphocyte apoptosis, and prolong the life span in MRL/lpr mice. Our results have confirmed and extended the capacity of FTY720 for such effects. FTY720 induced apoptosis in double negative T cells in vitro, reduced the progression of lupus nephritis in female MRL/lpr mice by inhibiting the development of anti-dsDNA antibodies and decreasing the number of B cells in the spleen, and normalized the proliferation of T cells as well as the production of IL-2 in response to anti-CD3 stimulation by increasing the relative number of functioning T cells.

Several immunomodulating drugs have been evaluated for their therapeutic effects on lupus in MRL/lpr mice. Cyclophosphamide (CYC) was established in 1984 as an effective drug for the treatment of lymphadenopathy, arthritis, and nephritis in MRL/lpr mice. Linomide (LS-2616) and mycophenolate mofetil retard the progression of disease to a degree comparable to that of CYC in MRL/lpr mice. Conflicting results have been published on the therapeutic efficacy of cyclosporin A (CSA). Berden, et al did not find any appreciable effects of CSA, while Mountz, et al described ameliorated immunopathology and prolonged survival with it. Other immunomodulators like FK506 and leflunomide, when tested in MRL/lpr mice, revealed some favorable effects. Our results indicate a therapeutic efficacy of FTY720 on the disease outcome in MRL/lpr mice.

Recent studies have implicated FTY720 directly in apoptotic cell death of lymphocytes, which may explain its

Table 1. Effects of FTY720 and methylprednisolone (mPSL) on body weight, lymphoid organ weight, spleen mononuclear cell (MNC) number, and bone marrow cell (BMC) number in female 32-week-old MRL/lpr mice after 16 weeks of treatment.

<table>
<thead>
<tr>
<th></th>
<th>Control, n = 4 (10)</th>
<th>FTY720, n = 10 (10)</th>
<th>mPSL, n = 9 (10)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body Weight, g</td>
<td>35.4 ± 3.1</td>
<td>36.5 ± 4.2</td>
<td>38.6 ± 3.2</td>
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<tr>
<td>Lymph node weight, g</td>
<td>0.41 ± 0.17</td>
<td>0.13 ± 0.12*</td>
<td>0.08 ± 0.02*</td>
</tr>
<tr>
<td>Spleen</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Weight, g</td>
<td>1.05 ± 0.34</td>
<td>0.60 ± 0.22*</td>
<td>0.50 ± 0.21*</td>
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<tr>
<td>MNC in spleen (×10⁶)</td>
<td>811 ± 263</td>
<td>462 ± 169*</td>
<td>381 ± 160*</td>
</tr>
<tr>
<td>B cells, %</td>
<td>5.9 ± 2.6</td>
<td>5.5 ± 1.9</td>
<td>4.1 ± 2.2</td>
</tr>
<tr>
<td>B cells in spleen (×10⁶)</td>
<td>47.8 ± 16.5</td>
<td>25.4 ± 8.3*</td>
<td>15.6 ± 7.6*</td>
</tr>
<tr>
<td>SP T cells, %</td>
<td>26.4 ± 11.6</td>
<td>75.1 ± 10.9*</td>
<td>71.3 ± 16.0*</td>
</tr>
<tr>
<td>SP T cells in spleen (×10⁶)</td>
<td>214 ± 94</td>
<td>346 ± 50</td>
<td>272 ± 61</td>
</tr>
<tr>
<td>DN T cells, %</td>
<td>57.9 ± 6.9</td>
<td>13.7 ± 8.2*</td>
<td>16.3 ± 7.9*</td>
</tr>
<tr>
<td>DN T cells in spleen (×10⁶)</td>
<td>470 ± 56</td>
<td>63 ± 38*</td>
<td>62 ± 30*</td>
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<td>Bone marrow</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>BMC (×10⁶)</td>
<td>14.3 ± 3.3</td>
<td>12.8 ± 4.8</td>
<td>14.0 ± 2.3</td>
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<tr>
<td>B cells, %</td>
<td>8.4 ± 2.6</td>
<td>8.1 ± 1.9</td>
<td>7.5 ± 1.1</td>
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<tr>
<td>B cells in BM (×10⁶)</td>
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<td>1.0 ± 0.3</td>
<td>1.1 ± 0.2</td>
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<td>Gr–1+ cells, %</td>
<td>68.7 ± 2.9</td>
<td>70.3 ± 2.2</td>
<td>71.8 ± 11.2</td>
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<td>Gr–1+ cells in BM (×10⁶)</td>
<td>9.8 ± 0.5</td>
<td>9.0 ± 0.3</td>
<td>10.1 ± 1.7</td>
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</table>

Statistical method: Mann-Whitney U test; * p < 0.05. Data are presented as means ± SD. † Number of mice at sacrifice. Figures in parentheses show the number of mice at the start of the experiment. SP: single positive; DN: double negative.
immunosuppressive effects. We found that FTY720 induced apoptosis in the thymocytes, but not in the splenic lymphocytes from normal BALB/c mice in vitro (Figure 1). Our results have substantial variance with a recent report of the high sensitivity of splenic lymphocytes to FTY720 in vitro. The reason for this discrepancy is unclear; it may be ascribed to the different concentrations of FTY720 used in their studies and ours. Our experiments were performed with 2 µM, which was found to be the lowest concentration for inducing maximal apoptosis in the thymocytes, whereas Shimizu, et al applied 10 µM, 5 times higher than we used. We confirmed that FTY720 increased apoptosis in the spleen cells in vitro if higher doses (30 µM) were added to culture media, albeit to an extent much lower than that with DEX (data not shown). However, a dose-response curve needs to be drawn to delineate precisely the cell selectivity of FTY720. Unlike in normal mice, FTY720 at 2 µM increased apoptosis in splenic double negative T cells from MRL/lpr mice in a relatively specific manner, whereas DEX at the same concentration induced apoptosis in both double negative and single positive T cells (Figures 2, 3, and 4). Although the reason for this apparent difference between FTY720 and DEX is yet to be determined, there are several possible explanations. First, the machinery of apoptosis triggered by FTY720 may be quite different from that of DEX. A recent study indicated that apoptosis induced by FTY720 involves the activation of c-Jun NH2-terminal kinase and p38 pathways. Second, it is also possible that FTY720 could be intrinsically weaker as a stimulus than DEX; the sensitivity of double negative T cells to apoptosis appears to be higher than that of single positive T cells from MRL/lpr mice. Third, FTY720 may selectively affect the lymphocytes in division and/or activation. Several studies have shown that double negative T cells in MRL/lpr mice are rapidly cycling in a state of activation. Moreover, FTY720 has been reported to increase activation induced peripheral T cell death in which Fas/Fas-Ligand (Fas-L) system is involved. Of particular interest are the recent observations that intracellular signaling of the Fas receptor is coupled to the sphingomyelin-ceramide pathway, which induces apoptosis. It is tempting to speculate that apoptosis induced by FTY720 may be mediated through a sphingomyelin-ceramide cycle by means of bypassing the Fas/Fas-L interaction, because FTY720 resembles sphingosine in chemical structure. Preliminary experiments suggested that the mechanism of apoptosis induced by FTY720 in vitro is similar to that of the membrane permeable ceramide analog, C2-ceramide, which also suppresses autoimmunity and mortality in MRL/lpr mice (Okazaki H, et al, manuscript in preparation). Consistent with our obser-
in vitro, FTY720 markedly decreased the number of double negative T cells in the periphery (Figure 8), resulting in a reduction in adenopathy and splenomegaly. Moreover, FTY720 significantly increased T cell proliferation and IL-2 production to the stimulation with anti-CD3, while mPSL did not (Figure 9). There were no differences between mice treated with FTY720 and mPSL in the degrees of reduction in double negative T cells, which are considered anergic to activation stimulus (Figure 8). Thus, it is possible that glucocorticoids exert immunosuppressive effects on the whole array of T cells nonspecifically, while FTY720 preferentially induces apoptosis in abnormally propagated T cells in MRL/lpr mice. Although FTY720 induced apoptosis in the double negative T cells in MRL/lpr mice in vitro, there is no direct evidence of FTY720 induced apoptosis in lymphoid organs in vivo from this study. In this regard, Nagahara, et al reported that FTY720 induced apoptosis in vivo in circulating T cells in rats and mice. The contribution of the FTY720 induced apoptosis on the therapeutic effect in vivo remains to be determined.

It is necessary to clarify the additional immunosuppressive effects of FTY720 other than the apoptosis-inducing activity. We did not examine the effects of FTY720 on the thymocytes from MRL/lpr mice in vivo. Recently, Yagi, et al reported that in vivo administration of FTY720 did not decrease the number of thymocytes from normal mice by
inhibiting the export of mature thymocytes to the periphery. Since double negative T cells in MRL/lpr mice could be derived from double positive cortical thymocytes, a decrease in double negative T cells in the periphery might be due to the inhibitory effect on T cell migration from the thymus. Moreover, FTY720 induced apoptosis of the bone marrow cells in vitro, while it did not in vivo. One possibility is that the concentration of FTY720 in the bone marrow may not have reached the level used in vitro. A second possibility is that the same mechanism might be operating in the bone marrow by inhibiting the cellular export from the bone marrow to the periphery.

The main cause of death in MRL/lpr mice is renal failure, and the significantly prolonged survival we found with FTY720 was presumably due to the reduced progression of glomerulonephritis. The observation that FTY720 decreased the deposition of IgG in the glomeruli in mice that had already developed renal disease indicates a therapeutic potential of FTY720 on established renal diseases. The mechanism by which FTY720 ameliorates glomerulonephritis is not fully understood. Recently, by establishing class I or II MHC deficient MRL/lpr mouse strains, it has been revealed that lymphoproliferation of double negative T cells and lupus nephritis in MRL/lpr mouse strains were induced by distinct mechanisms. Clearly, a decrease in the number of splenic B cells for a reduced production of pathogenic IgG anti-dsDNA antibodies is among the major factors of such effects. Moreover, a recent study showed that FTY720 decreases the infiltration of T cells into skin allografts in rats. A reduced T cell influx into kidneys with glomerulonephritis could limit the local production of disease-promoting cytokines.

We have shown therapeutic effects of FTY720, a novel apoptosis-inducing drug, in MRL/lpr mice. Probably due to its selective influence on activated T cells, FTY720 had the merit of few, if any, serious side effects in murine experimental SLE. Based on these results, we propose that FTY720 warrants clinical trials as a potential modifier of autoimmune diseases in humans.

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REFERENCES
22. Krause AW, Carley WW, Webb WW. Fluorescent erythrosine B is...


