The Pathogenesis of Rheumatoid Arthritis: Pivotal Cytokines Involved in Bone Degradation and Inflammation

CLIFTON O. BINGHAM III

ABSTRACT. Proinflammatory cytokines, notably interleukin 1 (IL-1) and tumor necrosis factor-α (TNF-α), play an important role in initiating and perpetuating inflammatory and destructive processes in the rheumatoid joint. These cytokines regulate many nuclear factor κB inducible genes that control expression of other cytokines, cell adhesion molecules, immunoregulatory molecules, and proinflammatory mediators. The expression of cyclooxygenase-2 and inducible nitric oxide synthase (iNOS) and thereby production of prostaglandins (PG) and NO are regulated by cytokines. PGE₂ and NO further promote inflammation and likely participate in destructive mechanisms in the rheumatoid joint. In some experimental systems, the effects of IL-1 and TNF-α appear synergistic, and correspondingly, concomitant inhibition of both cytokines provides greater than additive antiarthritic effects. Although the actions of IL-1 and TNF-α show a large degree of overlap, some differences have been observed in animal models. However, in patients with active rheumatoid arthritis, blockade of either cytokine results in clinical improvement and less radiographic progression. (J Rheumatol 2002;29 Suppl 65:3–9)

Key Indexing Terms:INDUCIBLE NITRIC OXIDE SYNTHASECYCLOOXYGENASE-2INDUCIBLE NITRIC OXIDE SYNTHASEINTERLEUKIN 1RHEUMATOID ARTHRITISTUMOR NECROSIS FACTOR-α

In rheumatoid arthritis (RA), an immunological trigger begins an inflammatory process that ultimately manifests clinically by typical signs and symptoms of disease, such as joint swelling and tenderness¹. Inflammation is also responsible for stimulating destructive mechanisms in the joint, which lead to structural damage and subsequently to functional declines and disability. The rheumatoid joint contains numerous cell types that are involved in these inflammatory and destructive processes^{2,3}. The inflamed synovial membrane contains synovial macrophages and fibroblasts (synoviocytes), whereas plasma cells, dendritic cells, T lymphocytes, and mast cells are found in the subsynovial layer. The composition of the synovial fluid varies, but it is principally composed of neutrophils that have infiltrated from the circulation. As synovial proliferation continues, pannus invades from the joint margins, triggering cartilage thinning that is mediated in part by the release of matrix metalloproteinases from synovial fibroblasts in addition to chondrocyte mediated destruction and failure of repair mechanisms. Bone destruction is also initiated through the activation of osteoclasts.

From the New York University School of Medicine and Department of Rheumatology and Medicine, NYU Hospital for Joint Diseases, New York, NY, USA.

Supported by an unrestricted educational grant from Amgen, Inc. C.O. Bingham III, MD.

Address reprint requests to Dr. C.O. Bingham III, NYU-Hospital for Joint Diseases, Department of Rheumatology and Medicine, 301 E. 17th St., Room 1410, New York, NY 10003. E-mail: clifton.bingham@med.nyu.edu

The synovial macrophage plays an important role in orchestrating inflammation and joint destruction in the rheumatoid joint⁴. These macrophages are activated by Th1 cytokines, including interferon-γ (IFN-γ), interleukin 12 (IL-12), and IL-18, which are released following T cell activation by antigen-presenting cells. Macrophage activation may also result from direct contact with T cells, as well as via stimulation by immune complexes or bacterial products found in the synovial fluid⁵. Once activated, macrophages release multiple cytokines and other inflammatory mediators that further amplify the inflammatory and destructive processes¹.

CYTOKINES

Cytokines are formed following cell activation (Figure 1), with increased transcription of cytokine genes leading to the synthesis and subsequent release of these mediators. The cytokines then interact with specific cell surface receptors, which initiate intracellular signal transduction in target cells and, ultimately, biological responses. Cytokines exert autocrine effects by interacting with receptors on cells that produce them, paracrine effects by binding to receptors on adjacent cells in the local microenvironment, and endocrine effects by acting on distant cells or organs⁶.

Each cytokine may have multiple, pleiotropic actions, and as a result, cytokine functions are frequently redundant. For example, IL-1 and tumor necrosis factor- α (TNF- α) share many proinflammatory actions in RA, whereas IL-4 and IL-13 cause overlapping antiinflammatory effects in RA

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2002. All rights reserved.

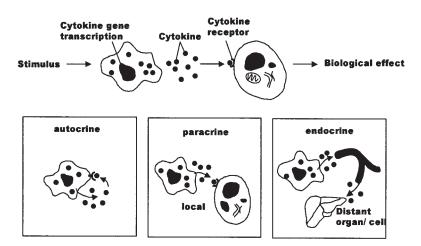


Figure 1. Steps leading to biological effects of cytokines. From Bingham, et al⁶, with permission.

while serving as integral mediators of asthmatic and allergic responses. When multiple cytokines are present, they may interact in a synergistic or antagonistic manner depending on the system under investigation^{7,8}. Cytokines can influence the production of one another, as is the case with IL-1 and TNF- α , each of which can induce the expression of the other⁹. Finally, cytokines can affect the expression and/or function of cytokine receptors, thereby regulating cytokine action.

The rheumatoid joint contains a variety of proinflammatory cytokines besides IL-1 and TNF-α, which include IL-6, IL-15, IL-16, IL-17, IL-18, IFN-γ, granulocyte macrophage-colony stimulating factor, and chemokines such as IL-8, macrophage inflammatory protein-1α and monocyte chemoattractant protein-1. Under normal physiologic conditions, the actions of these proinflammatory cytokines are maintained in balance by antiinflammatory cytokines, such as IL-4, IL-10, IL-11, and IL-13, and by natural cytokine antagonists, including IL-1 receptor antagonist (IL-1ra), soluble type 2 IL-1 receptor, soluble TNF receptor (sTNF-RI), and IL-18 binding protein. In the rheumatoid joint, however, the balance swings in favor of the proinflammatory cytokines¹⁰.

IL-1 and TNF-α have numerous functions throughout the body, many of which are important in RA^{2,11,12}. Both cytokines upregulate expression of cell adhesion molecules on endothelial cells, which is important in the recruitment of inflammatory cells to the inflammatory site^{13,14}. Both IL-1 and TNF activate a variety of cell types found in the rheumatoid joint, including macrophages, fibroblast-like synoviocytes, chondrocytes, and osteoclasts, resulting in the release of other proinflammatory mediators and degradative enzymes¹⁵⁻¹⁸. Both stimulate proliferation of synovial cells leading to pannus formation. Both cytokines influence immunological activity by causing T cell and B cell activation, and both stimulate hepatocytes to release acute phase

reactants, notably IL-6. Both have actions in the central nervous system: TNF- α stimulates endogenous glucocorticoid production via an action in the hypothalamus, and IL-1 induces fever and slow wave sleep.

A significant body of evidence suggests that IL-1 and TNF- α are important in RA pathogenesis. First, both cytokines are arthritogenic when injected into the joints of experimental animals^{19,20}. Second, spontaneous arthritis develops in transgenic animals overexpressing either cytokine²¹⁻²³. Moreover, a spontaneous erosive arthritis is found in IL-1ra knockout mice²⁴. Third, IL-1 or TNF- α blockade reduces inflammation and joint destruction in many animal models of RA^{25,26}. Synergistic effects are evident, with simultaneous inhibition of both cytokines. Finally, the most important piece of evidence comes from controlled clinical trials, where IL-1 antagonism or TNF- α blockade reduces the signs and symptoms of active RA and slows radiographic evidence of joint destruction²⁷⁻³⁴.

The similar biological actions of IL-1 and TNF- α may be explained, in part, by the similar intracellular signaling pathways that their receptors activate. The type 1 IL-1 receptor and TNF-receptors (TNF-R) activate a family of adapter proteins called TNF-R associated factors (TRAF) TRAF2 in the case of TNF- α and TRAF6 in the case of IL-1³⁵⁻³⁷. In turn, these TRAF activate downstream signaling pathways including nuclear factor κB (NF-κB)-mediated transduction mechanisms. NF-kB is normally found in the cytoplasm in an inactive complex with IkB. The TRAF activate an enzyme known as IkB kinase, which phosphorylates IκB, leading to its dissociation from the complex and resulting in the liberation of active NF-κB. The NF-κB then enters the nucleus and binds to promoter regions of multiple genes, resulting in their transcription and subsequently in generation of multiple products.

Many of the products of NF-κB inducible genes are involved in inflammation and immune responses (Table 1).

These genes encode adhesion molecules, cytokines, growth factors, cytokine receptors, immunoregulatory molecules, and acute phase proteins. In addition, NF-κB-inducible genes encode enzymes that are important in inflammatory mediator biosynthesis: cyclooxygenase (COX-2) and inducible nitric oxide synthase (iNOS). These enzymes are responsible for production of prostanoids and nitric oxide, respectively.

INFLAMMATORY LIPID MEDIATORS

The eicosanoids are a family of lipid mediators that are derived from arachidonic acid found in membrane phospholipids. Arachidonic acid is released by the action of phospholipase A₂ (PLA₂) enzymes and is then converted into prostanoids, prostaglandins, and thromboxanes, through the intermediate COX-1 and 2, and into leukotrienes through the intermediate 5-lipoxygenase enzyme acting in concert with 5-lipoxygenase activating protein. The intermediate prostaglandin product PGH, formed by COX-1 and 2 is subsequently converted into prostaglandin or thromboxane end products by terminal synthase enzymes. For example, PGE₂ synthase (PGES) is responsible for producing PGE₂. Many of the enzymes involved in prostaglandin synthesis exist in several forms. One form may be expressed constitutively, including cytosolic group IV-α PLA₂ (cPLA₂), COX-1, and cytosolic PGES (cPGES), whereas other forms of these enzymes are induced preferentially in inflammation and following cytokine stimulation, including COX-2, microsomal PGES (mPGES), and multiple types of secretory PLA₂ (sPLA₂) 38,39 .

COX-2 is highly expressed in the rheumatoid synovium. Strong COX-2 immunostaining was found in the synovial lining layer, synovial vessel endothelium, and in cells of the subsynovium⁴⁰. COX-2 expression was significantly higher in specimens from patients with RA as compared with osteoarthritis (OA). In comparison, COX-1 expression was

 $Table\ 1$. A partial list of NF-κB inducible genes involved in immune responses and inflammation.

Class	Genes
Inflammatory mediators	COX-2, iNOS
Cell adhesion molecules	ICAM-1, VCAM-1, E-selectin
Cytokines	IL-1, IL-2, IL-3, IL-6, IL-8, IL-12,
	TNF-α, IFN-α, IFN-β
Growth factors	G-CSF, M-CSF, GN-CSF
Cytokine receptors	IL-2R
Immunoregulatory molecules	Igk light chain, MHC class I and II, T cell receptor α and β
Acute phase proteins	SAA, complement factors B, C3 and C4

ICAM: intercellular adhesion molecule; VCAM: vascular cellular adhesion molecule; G-CSF: granulocyte colony stimulating factor; M-CSF: macrophage colony stimulating factor; GM-CSF: granulocyte macrophage-colony stimulating factor; SAA: serum amyloid A.

almost exclusively localized to the synovial lining layer, with no significant difference between RA and OA samples. Both IL-1 and TNF- α upregulated expression of COX-2 and mPGES in rheumatoid synoviocytes without affecting COX-1 or cPGES^{41,42}. The stimulation of COX-2 and mPGES was inhibited in a concentration-dependent manner by dexamethasone.

The sPLA, family consists of a large group of related molecules. The low molecular weight group IIA sPLA, was originally isolated from RA synovial fluid; it is induced by both IL-1 and TNF-α. Moreover, this sPLA, has proinflammatory activity of its own. At concentrations found in RA synovial fluid, type IIA sPLA, augmented TNF-α induced PGE₂ production by cultured synovial fibroblasts⁴². The expression and regulation of other members of the sPLA, family are largely unknown, but it is the subject of active investigation. It is apparent that these enzymes do not function solely by cleaving arachidonic acid from membrane phospholipids, but they may have unique receptor-mediated functions, such as inducing expression of COX-2 and other enzymes. It is notable that the expression of group V sPLA, in rheumatoid synovium closely parallels that of COX-2, with intense staining found in the synovial lining layer, blood vessel endothelial cells, and subsynovial lymphoid aggregates (Figure 2).

Upregulation of sPLA2, COX-2, and mPGES leads to enhanced production of PGE₂. This prostaglandin causes several effects that may be relevant to bone and cartilage erosion in RA^{43,44}. In vitro systems and animal models have demonstrated that PGE₂ induces bone resorption by osteoclasts¹⁸. On the basis of knockouts of the 4 PGE receptor subtypes, PGE₂ causes bone resorption by a cAMP dependent mechanism via the EP4 receptor⁴⁵. This effect of PGE, is consistent with observations that nonsteroidal antiinflammatory drugs, such as indomethacin, and COX-2 selective inhibitors retard osteoclast formation and activation 18. Moreover, COX-2 knockouts show impaired bone resorption in response to parathyroid hormone (PTH) or vitamin D. PGE, also stimulates new bone formation by osteoblasts by inducing expression of RANKL (receptor activator of NF-κB ligand), and depending on the experimental system, PGE, affects the synthesis and degradation of type II collagen and proteoglycans. Finally, at relatively high concentrations, PGE2 stimulates release of matrix metalloproteinases that degrade cartilage.

NITRIC OXIDE

Nitric oxide (NO) is another mediator that appears to be important in cartilage and bone destruction⁴⁶. Like the prostaglandins, NO is produced through constitutive and inducible pathways, which are responsible for its "house-keeping" and pathogenic roles, respectively. Like COX-2, the transcriptional control of iNOS is regulated by cytokines, such as IL-1 and TNF- α , as well as other cellular

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2002. All rights reserved.

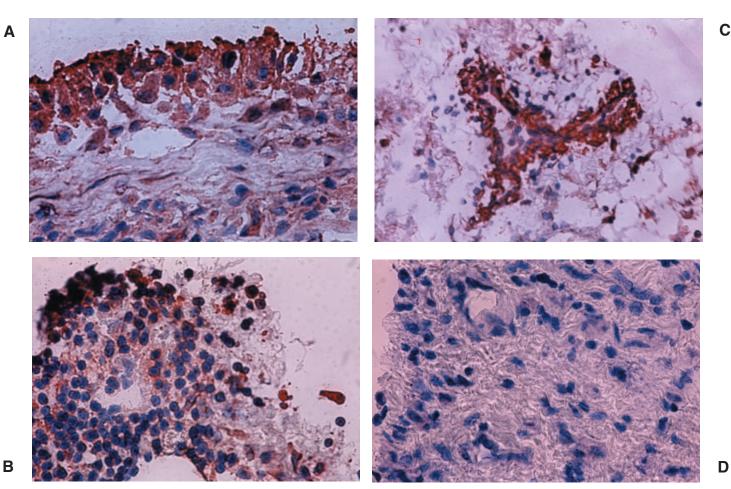


Figure 2. Immunohistologic localization of group V phospholipase A_2 (PLA₂) in rheumatoid synovium. Synovial biopsies were obtained from 3 patients with RA, and evaluated using standard immunohistochemistry. An affinity purified rabbit antipeptide group V PLA₂ antibody⁵⁴ or affinity purified rabbit IgG, both at 10 µg/ml, were applied to acetone fixed sections. Biotinylated secondary antibody was applied; slides were developed with 3-amino-9-ethylcarbazole with H_2O_2 , counterstained with Gill's hematoxylin, and examined under light microscopy. Representative samples are shown with group V PLA₂ antibody (A, B, C) or with control antibody (D): A. Synovial lining layer; B. synovial lining, subsynovial lymphoid aggregate; C. blood vessel, endothelial cells; and D. control antibody.

stimuli (Figure 3)⁴⁷. The iNOS enzyme catalyzes the conversion of L-arginine in the presence of molecular oxygen and the cofactor NADPH into NO and L-citrulline. Among its many biological actions, NO activates matrix metalloproteinases, inhibits collagen and proteoglycan synthesis by chondrocytes, and promotes vasodilation, which leads to fluid and cellular influx into an inflammatory site. NO also combines with reactive oxygen species to produce peroxynitrite, which promotes chondrocyte apoptosis⁴⁸.

In murine models, NO derived from iNOS may be essential for IL-1 induced bone resorption⁴⁹. In iNOS deficient mice, IL-1 induced osteoclastic bone resorption was disrupted, but resorption caused by parathyroid hormone or vitamin D was unaffected. The defect in IL-1 induced bone resorption was attributed to a defect in NF-κB nuclear translocation and binding to promoter sites in the iNOS deficient animals.

SYNERGISM OF IL-1 AND TNF-α

Intraarticular injection of IL-1 or TNF- α into a rabbit knee joint markedly increases the number of infiltrating leukocytes⁷. The nature of the inflammatory infiltrate was predominantly monocytic with TNF- α , whereas it was both neutrophilic and monocytic with IL-1. When administered in combination at submaximal doses, IL-1 and TNF- α were synergistic with respect to both neutrophil and monocyte infiltration. The number of neutrophils and monocytes were 2.8 and 1.8 times higher, respectively, than would be expected if the effects were simply additive.

Similarly, greater than additive effects have been observed in animal models of RA when IL-1 and TNF- α blockers were combined⁵⁰. In established collagen induced arthritis in rats, treatment with either recombinant human IL-1ra or PEGylated sTNF-RI partially reduced several disease variables evaluated histologically, including inflammation, pannus, cartilage damage, and bone resorption

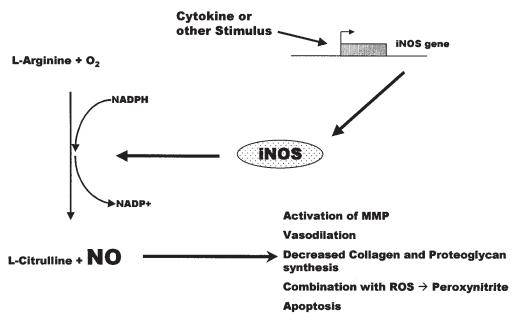


Figure 3. Nitric oxide induction by cytokines and other stimuli.

(Figure 4). When both agents were administered in combination, inflammation was reduced in an additive manner, whereas the other 3 variables were almost completely suppressed.

Some differences between IL-1 and TNF- α have been seen in animal models, and differences may be apparent in individual patients treated with inhibitors of these cytokines. In the rat adjuvant arthritis model, for example, IL-1ra inhibited angiogenesis in the joint, whereas blocking TNF- α did not reduce the number of capillaries⁵¹. In murine

collagen induced arthritis, IL-1 blockade inhibited cartilage and bone destruction, whereas TNF- α inhibition only reduced joint inflammation⁵². Nevertheless, it is important to recognize that animal models of RA do not necessarily reflect clinical disease. Importantly, treatment with either IL-1 or TNF- α blockers reduces inflammation and slows joint destruction in patients with active RA²⁷⁻³⁴. It remains to be determined, however, whether patients with poor treatment responses to either an IL-1 or TNF- α blocker will have better responses when an inhibitor of the other cytokine is

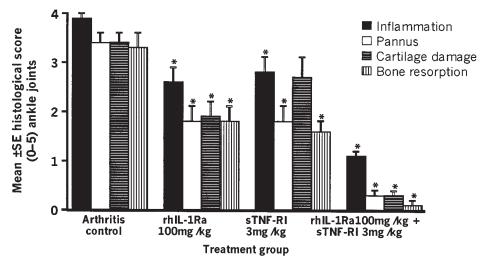


Figure 4. Synergism of combined IL-1 and TNF- α blockade in established collagen induced arthritis in rats. Shown are the effects of recombinant human IL-1ra 100 mg/kg and PEGylated sTNF-RI 3 mg/kg alone and in combination on histological scores of inflammation, pannus, cartilage damage, and bone resorption. *P < 0.05, 2 tailed t test to control. From Bendele, et al⁵⁰ with permission.

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2002. All rights reserved.

administered. The question of synergy of effect using IL-1 and TNF inhibitors is being addressed in ongoing clinical trials of patients with rheumatoid arthritis; however, one report has suggested an increased incidence of severe infections when combinations of biologics are administered⁵³.

REFERENCES

- Choy EHS, Panayi GSP. Cytokine pathways and joint inflammation in rheumatoid arthritis. N Engl J Med 2001;344:907-16.
- Feldman M, Brennan FM, Maini RN. Role of cytokines in rheumatoid arthritis. Annu Rev Immunol 1996;14:397-440.
- Moreland LW. Potential biologic agents for treating rheumatoid arthritis. Rheum Dis Clin North Am 2001;27:445-91.
- Kinne RW, Brauer R, Stuhlmuller B, Palombo-Kinne E, Burmester GR. Macrophages in rheumatoid arthritis. Arthritis Res 2000; 2:189-202.
- Vey E, Dayer JM, Burger D. Direct contact with stimulated T cells induces the expression of IL-1ß and IL-1 receptor antagonist in human monocytes: involvement of serine/threonine phosphatases in differential regulation. Cytokine 1997;9:480-7.
- Bingham CO III, Abramson SA. Mechanisms of inflammation. In: Tannenbaum H, Russell AS, editors. Mechanisms in rheumatology. Toronto: Core Health Services; 2001.
- Henderson B, Pettipher ER. Arthritogenic actions of recombinant IL-1 and tumour necrosis factor-alpha in the rabbit: evidence for synergistic interactions between cytokines in vivo. Clin Exp Immunol 1989;75:306-10.
- van Roon JA, Lafeber FP, Bijlsma JW. Synergistic activity of interleukin-4 and interleukin-10 in suppression of inflammation and joint destruction in rheumatoid arthritis. Arthritis Rheum 2001; 44:3-12.
- Nawroth PP, Bank I, Handley D, Cassimeris J, Chess L, Stern D. Tumor necrosis factor/cachectin interacts with endothelial cell receptors to induce release of interleukin 1. J Exp Med 1986;163:1363-75.
- Arend WP. Cytokine imbalance in the pathogenesis of rheumatoid arthritis: the role of interleukin-1 receptor antagonist. Semin Arthritis Rheum 2001;30 Suppl 2:1-6.
- 11. Gabay C, Arend WP. Treatment of rheumatoid arthritis with IL-1 inhibitors. Springer Semin Immunopathol 1998;20:229-46.
- Charles P, Elliott MJ, Davis D, et al. Regulation of cytokines, cytokine inhibitors, and acute-phase protein proteins following anti-TNF-α therapy in rheumatoid arthritis. J Immunol 1999;163:1521-8.
- Lindsley HB, Smith DD, Cohick CB, Koch AE, Davis LS. Proinflammatory cytokines enhance human synoviocyte expression of intercellular adhesion molecule-1. Clin Immunol Immunopathol 1993;68:311-20.
- Proudman SM, Cleland LG, Mayrhofer G. Effects of tumor necrosis factor-α, interleukin 1β, and activated peripheral blood mononuclear cells on the expression of adhesion molecules and recruitment of leukocytes in rheumatoid synovial xenografts in SCID mice. J Rheumatol 1999;26:1877-89.
- Alvaro-Gracia JM, Zvaifler NJ, Brown CB, Kaushansky K, Firestein GS. Cytokines in chronic inflammatory arthritis. VI. Analysis of the synovial cells involved in granulocyte-macrophage colony-stimulating factor production and gene expression in rheumatoid arthritis and its regulation by IL-1 and tumor necrosis factor-α. J Immunol 1991;146:3365-71.
- Schlaak JF, Schwarting A, Knolle P, Meyer zum Buschenfelde KH, Mayet W. Effects of Th1 and Th2 cytokines on cytokine production and ICAM-1 expression on synovial fibroblasts. Ann Rheum Dis 1995;54:560-5.
- 17. Henrotin YE, De Groote DD, Labasse AH, et al. Effects of

- exogenous IL-1 β , TNF- α , IL-6, IL-8, and LIF on cytokine production by human articular chondrocytes. Osteoarthritis Cartilage 1996;4:163-73.
- Lader CS, Flanagan AM. Prostaglandin E₂, interleukin 1ß, and tumor necrosis factor-α increase human osteoclast formation and bone resorption in vitro. Endocrinology 1998;139:3157-64.
- Staite ND, Richard KA, Aspar DG, Franz KA, Galinet LA, Dunn CJ. Induction of an acute erosive monarticular arthritis in mice by interleukin-1 and methylated bovine serum albumin. Arthritis Rheum 1990;33:253-60.
- Saez-Llorens X, Jafari HS, Olsen KD, Nariuchi H, Hansen EJ, McCracken GH Jr. Induction of suppurative arthritis in rabbits by Haemophilus endotoxin, tumor necrosis factor-α and interleukin-18. J Infect Dis 1991;163:1267-72.
- Ghivizzani SC, Kang R, Georgescu HI, et al. Constitutive intra-articular expression of human IL-1ß following gene transfer to rabbit synovium produces all major pathologies of human rheumatoid arthritis. J Immunol 1997;159:3604-12.
- Niki Y, Yamada H, Seki S, et al. Macrophage- and neutrophil-dominant arthritis in human IL-1ß transgenic mice. J Clin Invest 2001;107:1127-35.
- Keffer J, Probert L, Cazlaris H, et al. Transgenic mice expressing human tumour necrosis factor: a predictive genetic model of arthritis. EMBO J 1991;10:4025-31.
- Horai R, Saijo S, Tanioka H, et al. Development of chronic inflammatory arthropathy resembling rheumatoid arthritis in interleukin-1 receptor antagonist-deficient mice. J Exp Med 2000;191:313-20.
- Arend WP, Malyak M, Guthridge CJ, Gabay C. Interleukin-1 receptor antagonist: role in biology. Annu Rev Immunol 1998;16:27-55.
- Maini RN, Taylor PC. Anti-cytokine therapy for rheumatoid arthritis. Annu Rev Med 2000;51:207-29.
- Bresnihan B, Alvaro-Gracia JM, Cobby M, et al. Treatment of rheumatoid arthritis with recombinant human interleukin-1 receptor antagonist. Arthritis Rheum 1998;41:2196-204.
- Jiang Y, Genant HK, Watt I, et al. A multicenter, double-blind, dose-ranging, randomized, placebo-controlled study of recombinant human interleukin-1 receptor antagonist in patients with rheumatoid arthritis. Radiologic progression and correlation of Genant and Larsen scores. Arthritis Rheum 2000;43:1001-9.
- Cohen S, Hurd E, Cush J, et al. Treatment of rheumatoid arthritis with anakinra, a recombinant human interleukin-1 receptor antagonist (IL-1ra), in combination with methotrexate. Arthritis Rheum 2002;46:614-24.
- Moreland LW, Schiff MH, Baumgartner SW, et al. Etanercept therapy in rheumatoid arthritis. A randomized, controlled trial. Ann Intern Med 1999;130:478-86.
- Weinblatt ME, Kremer JM, Bankhurst AD, et al. A trial of etanercept, a recombinant tumor necrosis factor receptor:Fc fusion protein, in patients with rheumatoid arthritis receiving methotrexate. N Engl J Med 1999;340:253-9.
- Bathon JM, Martin RW, Fleischmann RM, et al. A comparison of etanercept and methotrexate in patients with early rheumatoid arthritis. N Engl J Med 2000;343:1586-93.
- 33. Maini R, St. Clair EW, Breedveld F, et al. Infliximab (chimeric anti-tumour necrosis factor α monoclonal antibody) versus placebo in rheumatoid arthritis patients receiving concomitant methotrexate: a randomized phase III trial. Lancet 1999;354:1932-9.
- Lipsky PE, van der Heijde DM, St. Clair EW, et al. Infliximab and methotrexate in the treatment of rheumatoid arthritis. N Engl J Med 2000;343:1594-602.
- 35. Bradley JR, Pober JS. Tumor necrosis factor receptor-associated factors. Oncogene 2001;20:6482-91.
- 36. Inoue J, Ishida T, Tsukamoto N, et al. Tumor necrosis factor

- receptor-associated factor (TRAF) family: adapter proteins that mediate cytokine signaling. Exp Cell Res 2000;254:14-24.
- Gravallese EM, Galson DL, Goldring SR, Auron PE. The role of TNF-receptor family members and other TRAF-dependent receptors in bone resorption. Arthritis Res 2001;3:6-12.
- Katori M, Majima M. Cyclooxygenase-2: its rich diversity of roles and possible application of its selective inhibitors. Inflamm Res 2000;49:367-92.
- Jamal OS, Conaghan PG, Cunningham AM, et al. Increased expression of human type IIa secretory phospholipase A₂ antigen in arthritic synovium. Ann Rheum Dis 1998;57:550-8.
- Siegle I, Klein T, Backman JT, Saal JG, Nusing RM, Fritz P. Expression of cyclooxygenase 1 and cyclooxygenase 2 in human synovial tissue: differential elevation of cyclooxygenase 2 in inflammatory joint diseases. Arthritis Rheum 1998;41:122-9.
- Stichtenoth DO, Thoren S, Bian H, Peters-Golden M, Jakobsson PJ, Crofford LJ. Microsomal prostaglandin E synthase is regulated by proinflammatory cytokines and glucocorticoids in primary rheumatoid synovial cells. J Immunol 2001;167:469-74.
- Bidgood MJ, Jamal OS, Cunningham AM, Brooks PM, Scott KF. Type IIA secretory phospholipase A₂ up-regulates cyclooxygenase-2 and amplifies cytokine-mediated prostaglandin production in human rheumatoid synoviocytes. J Immunol 2000;165:2790-7.
- Raisz LG. Prostaglandins and bone: physiology and pathophysiology. Osteoarthritis Cartilage 1999;7:419-21.
- Abramson SB. The role of COX-2 produced by cartilage in arthritis. Osteoarthritis Cartilage 1999;7:380-1.
- Miyaura C, Inada M, Suzawa T, et al. Impaired bone resorption to prostaglandin E₂ in prostaglandin receptor EP4-knockout mice. J Biol Chem 2000;275:19819-23.

- 46. Lotz M. The role of nitric oxide in articular cartilage damage. Rheum Dis Clin North Am 1999;25:269-82.
- Grabowski PS, Macpherson H, Ralston SH. Nitric oxide production in cells derived from the human joint. Br J Rheumatol 1996; 35:207-12.
- 48. Lotz M, Hashimoto S, Kuhn K. Mechanisms of chondrocyte apoptosis. Osteoarthritis Cartilage 1999;7:389-91.
- van't Hof RJ, Armour KJ, Smith LM, et al. Requirement of the inducible nitric oxide synthase pathway for IL-1-induced osteoclastic bone resorption. Proc Natl Acad Sci USA 2000;97:7993-8.
- Bendele AM, Chlipala ES, Scherrer J, et al. Combination benefit of treatment with the cytokine inhibitors interleukin-1 receptor antagonist and PEGylated soluble tumor necrosis factor receptor type I in animal models of rheumatoid arthritis. Arthritis Rheum 2000;43:2648-59.
- Coxon A, Bolon B, Estrada J, et al. Differential control of angiogenesis by anakinra (IL-1ra) and PEG sTNF-R1 [abstract]. Arthritis Rheum 2001;44 Suppl:S242.
- Joosten LAB, Helsen MMA, Saxne T, van de Loo FA, Heinegard D, van den Berg WB. IL-1αβ blockade prevents cartilage and bone destruction in murine type II collagen-induced arthritis, whereas TNF-α blockade only ameliorates joint inflammation. J Immunol 1999:163:5049-55.
- Schiff MH, Bulpitt K, Weaver AA, et al. Safety of combination therapy with anakinra and etanercept in patients with RA [abstract]. Arthritis Rheum 2001;44 Suppl:S79.