

Rituximab in the Treatment of Interstitial Lung Disease Associated with Antisynthetase Syndrome: A Multicenter Retrospective Case Review

Tracy J. Doyle, Namrata Dhillon, Rachna Madan, Fernanda Cabral, Elaine A. Fletcher, Diane C. Koontz, Rohit Aggarwal, Juan C. Osorio, Ivan O. Rosas, Chester V. Oddis, and Paul F. Dellaripa

ABSTRACT. Objective. To assess clinical outcomes including imaging findings on computed tomography (CT), pulmonary function testing (PFT), and glucocorticoid (GC) use in patients with the antisynthetase syndrome (AS) and interstitial lung disease (ILD) treated with rituximab (RTX).

Methods. We retrospectively identified all patients at 2 institutions with AS-ILD who were treated with RTX. Baseline demographics, PFT, and chest CT were assessed before and after RTX. Two radiologists independently evaluated CT using a standardized scoring system.

Results. Twenty-five subjects at the Brigham and Women's Hospital (n = 13) and University of Pittsburgh Medical Center (n = 12) were included. Antisynthetase antibodies were identified in all patients (16 Jo1, 6 PL-12, 3 PL-7). In 21 cases (84%), the principal indication for RTX use was recurrent or progressive ILD, owing to failure of other agents. Comparing pre- and post-RTX pulmonary variables at 12 months, CT score and forced vital capacity were stable or improved in 88% and 79% of subjects, respectively. Total lung capacity (%) increased from 56 ± 13 to 64 ± 13 and GC dose decreased from 18 ± 9 to 12 ± 12 mg/day. Although DLCO (%) declined slightly at 1 year, it increased from 42 ± 17 to 70 ± 20 at 3 years. The most common imaging patterns on CT were nonspecific interstitial pneumonia (NSIP; n = 13) and usual interstitial pneumonia/fibrotic NSIP (n = 5), of which 5 had concurrent elements of cryptogenic organizing pneumonia.

Conclusion. Stability or improvement in pulmonary function or severity of ILD on CT was seen in most patients. Use of RTX was well tolerated in the majority of patients. RTX may play a therapeutic role in patients with AS-ILD, and further clinical investigation is warranted. (J Rheumatol First Release April 1 2018; doi:10.3899/jrheum.170541)

Key Indexing Terms:

INTERSTITIAL LUNG DISEASE

ANTISYNTHEASE SYNDROME

RITUXIMAB

From the Departments of Medicine and Radiology, Brigham and Women's Hospital, Boston, Massachusetts; Departments of Medicine and Radiology, University of Pittsburgh School of Medicine, Pittsburgh, Pennsylvania, USA. Supported by the US National Institutes of Health (grant number 1 K23 HL119558 to TJD).

T.J. Doyle, MD, MPH, Pulmonary and Critical Care Division, Brigham and Women's Hospital, Harvard Medical School; N. Dhillon, MD, Division of Rheumatology and Clinical Immunology, University of Pittsburgh School of Medicine; R. Madan, MD, Department of Radiology, Brigham and Women's Hospital; F. Cabral, MD, Department of Radiology, Brigham and Women's Hospital; E.A. Fletcher, Pulmonary and Critical Care Division, Brigham and Women's Hospital, Harvard Medical School; D.C. Koontz, BS, Division of Rheumatology and Clinical Immunology, University of Pittsburgh School of Medicine; R. Aggarwal, MD, MS, Division of Rheumatology and Clinical Immunology, University of Pittsburgh School of Medicine; J.C. Osorio, MD, Pulmonary and Critical Care Division, Brigham and Women's Hospital, Harvard Medical School; I.O. Rosas, MD, Pulmonary and Critical Care Division, Brigham and Women's Hospital, Harvard Medical School; C.V. Oddis, MD, Division of Rheumatology and Clinical Immunology, University of Pittsburgh School of Medicine; P.F. Dellaripa, MD, Division of Rheumatology, Immunology and Allergy, Brigham and Women's Hospital.

Address correspondence to Dr. P.F. Dellaripa, Division of Rheumatology, Immunology and Allergy, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, 75 Francis St., Boston, Massachusetts 02115, USA. E-mail: pdellaripa@partners.org

Accepted for publication November 15, 2017.

The antisynthetase syndrome (AS) is rare and characterized by myositis associated with antisynthetase antibodies, including, but not limited to, anti-Jo1, -PL-7 (threonyl), -PL-12 (alanyl), -OJ (isoleucyl), and -EJ (glycyl), as well as additional clinical features including polyarthritides, fever, Raynaud phenomenon, mechanic's hands, and interstitial lung disease (ILD)^{1,2}. ILD is the most serious complication of AS (AS-ILD), occurring in 70–89% of patients, with an increase in morbidity and mortality in affected individuals^{3,4,5,6,7,8}. Some variability in ILD incidence, phenotype, and mortality has been noted based on antibody subtype^{7,8,9}. It is imperative to establish effective treatment options for this serious complication. To date, many therapies have been reported for ILD, with glucocorticoids (GC) remaining as the first line of treatment^{3,5,10}. Because of the lack of prospective trials, specific use of additional immunosuppressive agents is empiric.

Rituximab (RTX) is a chimeric monoclonal antibody against CD20+ B cells, which results in B cell depletion^{11,12}. Evidence for the effectiveness of B cell depletion exists in a number of immune-mediated conditions, including rheuma-

toid arthritis, antineutrophil cytoplasmic antibody-associated vasculitis, and immune thrombocytopenic purpura^{13,14,15,16,17,18}. RTX has also been specifically studied in myositis^{19,20,21,22,23,24} and scleroderma-ILD^{25,26}. Of note, a retrospective analysis of 50 patients with severe ILD of varying etiologies, excluding idiopathic pulmonary fibrosis (IPF), demonstrated an improvement in pulmonary function tests (PFT) at 6 and 12 months post-RTX²⁷. The only randomized trial of RTX therapy in AS was nested within a double-blind, placebo-phase trial of RTX in 200 refractory adult and juvenile dermatomyositis and adult polymyositis cases by Oddis, *et al*, in which 17% of subjects possessed antisynthetase autoantibodies (primarily anti-Jo1)¹⁹. In the Oddis, *et al* prospective trial, 83% of subjects met the predetermined definition of improvement after RTX, but no subgroup analysis was performed and lung function and computed tomography (CT) changes were not assessed as an outcome in the antisynthetase-positive subjects. There are limited data regarding the use of RTX in AS-ILD, mostly derived from retrospective case series/case reports, in which improvements in pulmonary function and high-resolution CT (HRCT) scans were noted^{20,21,28–36}. Adverse events mainly included infectious complications, such as *P. jirovecii* pneumonia, which were occasionally fatal, as well as rash, arrhythmia, and serum sickness^{21,30,31,33,34,37}.

Our objective was to assess clinical outcomes, including pulmonary function, severity of ILD on HRCT, and concurrent GC dosing in a cohort of patients with AS-ILD treated with RTX at 2 institutions.

MATERIALS AND METHODS

Study design and population. We retrospectively identified all patients at the Brigham and Women's Hospital (BWH), Boston, Massachusetts, USA, and the University of Pittsburgh Medical Center (UPMC), Pittsburgh, Pennsylvania, USA, with antisynthetase autoantibodies who presented with ILD and were treated with RTX since 2007 (BWH) and 2005 (UPMC). Inclusion criteria comprised the presence of antisynthetase autoantibodies, a diagnosis of ILD, treatment with RTX, and ≥ 1 PFT and/or CT scan at baseline and again 1–3 years after treatment with RTX. Exclusion criteria included lack of adequate followup or lung transplantation prior to 1 year after administration of RTX. Demographic characteristics, antisynthetase autoantibodies, clinical symptoms, laboratory abnormalities, concomitant GC (i.e., prednisone dose), and other immunosuppressive use, PFT, and HRCT chest imaging findings were extracted from the electronic medical record through April 2016. At BWH, RTX was routinely dosed every 6 months after the initial administration of RTX, whereas at UPMC, RTX was given initially with variable intervals of subsequent administration in the AS-ILD subjects.

Antisynthetase antibody detection. The majority of antisynthetase antibodies were detected using the Myositis Profile available through the Oklahoma Medical Research Foundation Clinical Immunology Laboratory ($n = 21$), which included testing for 12 myositis-specific and myositis-associated antibodies using RNA immunoprecipitation. Testing on the other 4 subjects was performed through a variety of other laboratories. Anti-SSA was measured using standard chemiluminescent immunoassay—certified laboratories at both institutions.

Pulmonary function testing. Serial PFT completed primarily for clinical indications were reviewed. For consistency, only prebronchodilator values

were used for the analyses because bronchodilator therapy was not routinely given. Reference values for spirometry were derived from the Third National Health and Nutrition Examination Survey in the United States³⁸, whereas lung volumes were standardized using predicted equations based on Crapo, *et al*³⁹, and DLCO-predicted equations were based on Cotes, *et al*⁴⁰.

HRCT analysis. Chest CT scans were done at the 2 study sites using standard institutional CT protocols (including axial HRCT images) as clinically indicated. Two radiologists (RM and FC, with 8 and 2 years' experience, respectively, as thoracic radiologists) independently evaluated axial 3–5 mm chest CT scans and 1 mm HRCT scans, recording their subjective assessment of the ILD pattern as usual interstitial pneumonia (UIP), nonspecific interstitial pneumonia (NSIP), or cryptogenic organizing pneumonia (COP). Coexistence of ≥ 1 CT pattern was possible and was also recorded. This was followed by a more detailed quantitative assessment and calculation of a CT severity score (Supplementary Table 1, available from the authors on request). The thin-section CT findings were graded on a scale of 1–6 as shown in Supplementary Table 1. Presence of each of the 6 different types of imaging patterns was assessed independently in 3 (upper, middle, and lower) zones of each lung. The upper zone was defined as the area above the level of the carina, the middle zone as the area between the level of the carina and the level of inferior pulmonary vein, and the lower zone as the area below the level of inferior pulmonary vein. The extent of each abnormality was determined by visually estimating the percentage (to the nearest 5%) of the affected lung parenchyma in each zone. The abnormality score for each zone was calculated by multiplying the percentage area by the point value (the score of 1–6)^{41,42}. The score for each zone was calculated by multiplying the percentage of the area to the nearest 5% by the grading scale score. The 6 zone scores were averaged to determine the total score for each patient. The scores of the 2 observers were averaged.

Statistical analysis. Chest CT scan scores, PFT, and prednisone dose were assessed before and after RTX. Average values were calculated at baseline for all subjects with followup values at any timepoint (1, 2, or 3 yrs), and average values at subsequent timepoints were calculated only for subjects with baseline values for comparison. An improvement in CT severity scores was defined as a $> 10\%$ decrease, while improvement in PFT was defined as a $> 10\%$ increase in forced vital capacity (FVC). Failure is defined by a $> 10\%$ increase in CT score or a $> 10\%$ decline in FVC. Stability was defined as between these values — a $< 10\%$ decrease but $< 10\%$ increase in CT score and a $< 10\%$ increase but $< 10\%$ decrease in FVC⁴³. Paired univariate analyses were conducted with Wilcoxon signed-rank tests where appropriate. Subgroup analyses were based on antisynthetase autoantibodies, ILD pattern, baseline mean HRCT score (194), PFT variables indicating severe disease [FVC $< 50\%$, total lung capacity (TLC) $< 50\%$, and DLCO $< 35\%$; some of which have been associated with increased ILD mortality^{44,45}], use of RTX as initial or rescue therapy, and the presence of concurrent immunosuppression at the time of starting RTX. A p value of < 0.05 was considered statistically significant. All analyses were performed using Statistical Analysis Software version 9.3 (SAS Institute). This project was granted IRB approval by the Partners Human Research Committee at BWH (protocol no. 2014P000110) and UPMC (IRB0409097).

RESULTS

At BWH, 16 consecutive patients treated with RTX for AS-ILD since 2007 were identified. One proceeded to lung transplant 2 months after RTX and was excluded, while 2 additional patients were excluded because of lack of PFT or HRCT followup. There were no deaths recorded in the medical record. At UPMC, 21 consecutive patients with AS-ILD treated with RTX since 2005 were identified. Eight were excluded because of lack of followup PFT and/or HRCT data, 1 of whom died from respiratory failure 5 months after receiving RTX; 1 additional subject proceeded

Table 1. Available baseline and followup CT and PFT for each BWH and UPMC subject.

Subject	Baseline CT	1-yr followup CT	2-yr followup CT	3-yr followup CT	Baseline PFT	1-yr followup PFT	2-yr followup PFT	3-yr followup PFT
BWH								
1	•	•			•		•	•
2	•			•	•	•	•	•
3	•	•			•	•	•	
4	•		•		•	•	•	
5	•		•		•	•	•	•
6	•	•			•	•		
7	•	•			•	•	•	•
8	•		•	•	•		•	•
9	•				•	•		
10	•				•	•	•	
11	•				•	•	•	
12	•	•			•	•		
13	•		•	•				
UPMC								
1					•	•		
2	•	•			•	•		
3	•				•	•		
4	•				•			•
5	•				•	•	•	
6	•	•			•			
7	•	•			•	•		
8	•			•	•	•		•
9					•	•	•	
10	•		•		•			
11					•	•		
12					•	•	•	

CT: computed tomography; PFT: pulmonary function tests; BWH: Brigham and Women's Hospital; UPMC: University of Pittsburgh Medical Center.

to lung transplant 10 months after RTX and was also excluded. Two UPMC subjects included in our study also died: 1 from respiratory failure secondary to progression of underlying ILD 2 years after receiving RTX (Supplementary Table 2, UPMC Subject 4, available from the authors on request), and 1 from anoxic brain injury about 10 years after receiving RTX (Supplementary Table 2, UPMC Subject 6). Available baseline and followup HRCT and PFT for each BWH and UPMC subject are detailed in Table 1.

Of the 25 subjects included in our cohort, the mean age was 49 years, and 20 (80%) were female (Table 2). Eight patients (32%) had a smoking history and 2 (8%) were current smokers. Antisynthetase antibodies were identified in all patients (16 Jo1, 6 PL-12, 3 PL-7). Four subjects were anti-SSA-positive, 15 were negative, and for 6 the information was unavailable. The most common clinical features noted at any time in the clinical course included myositis (n = 19) and Raynaud phenomenon (n = 14), followed by mechanic's hands/rashes/arthritis (n = 12 each), and fever (n = 6). Baseline HRCT (n = 21) revealed the following patterns: 9 NSIP, 4 UIP/fibrotic NSIP, 4 NSIP/COP, 1 COP, 1 fibrotic NSIP/COP, 1 COP/bronchiolitis obliterans, and 1 acute interstitial pneumonitis (AIP)/diffuse alveolar damage

(Figure 1). Abnormalities of the esophagus identified on HRCT included a patulous esophagus (n = 7) and hiatal hernia (n = 1). The Spearman rank correlation coefficient for quantitative CT scoring interobserver agreement between the 2 radiologists was 0.95.

The diagnosis of AS followed the diagnosis of ILD in 18 patients (72%) by a mean of 2.5 years. The mean time to initiation of RTX after ILD identification was 4.4 years. In 21 cases (84%), the principal indication for RTX use was recurrent or progressive ILD owing to failure of other immunosuppressive agents. In 4 subjects (16%), RTX was used as the first GC-sparing agent. Medications used prior to switching to RTX are detailed in Table 1. Twenty-three subjects were taking prednisone at the time of starting RTX (92%) and 12 (48%) were taking additional concurrent immunosuppressive agents at the time of starting RTX: mycophenolate mofetil (MMF; n = 7), tacrolimus and MMF (n = 1), azathioprine (n = 1), cyclophosphamide (n = 1), intravenous immunoglobulin (n = 1), and methotrexate (n = 1). Three patients had documented adverse events after the initial RTX dosing: 1 anaphylaxis and 2 serious gastrointestinal complications requiring surgery; they all later resumed RTX. Infectious complications while taking RTX included

Table 2. Baseline demographics of AS subjects. Values are n (%) or mean \pm SD.

Variables	BWH Cohort, n = 13	UPMC Cohort, n = 12	Total AS Subjects, n = 25
Age at time of RTX, yrs	51 \pm 10	47 \pm 14	49 \pm 12
Sex, female	12 (92)	8 (67)	20 (80)
Race, white	4 (31)	6 (50)	10 (40)
BMI	27.5 \pm 9.6	31.4 \pm 7.6	29.1 \pm 8.9
Ever smoker	4 (31)	4 (33)	8 (32)
Current smoker	0 (0)	2 (17)	2 (8)
Antisynthetase antibodies			
Jo1	6 (46)	10 (83)	16 (64)
PL-12	4 (31)	2 (17)	6 (24)
PL-7	3 (23)	0 (0)	3 (12)
Anti-SSA	1 (8)	3 (25)	4 (16)
Symptoms			
Myositis	12 (92)	7 (58)	19 (76)
Arthritis	9 (69)	3 (25)	12 (48)
Mechanic's hands	9 (69)	3 (25)	12 (48)
Rashes	8 (62)	4 (33)	12 (48)
Raynaud phenomenon	7 (54)	7 (58)	14 (56)
Fever	4 (31)	2 (17)	6 (24)
Medications, prior to RTX/concurrent with RTX initiation			
AZA	9 (69)/1 (8)	7 (58)/0 (0)	16 (64)/1 (4)
MMF	8 (62)/4 (31)	8 (67)/4 (33)	16 (64)/8 (32)
CYC	6 (46)/0 (0)	5 (42)/1 (8)	11 (44)/1 (4)
Tacrolimus	2 (15)/0 (0)	7 (58)/1 (8)	9 (36)/1 (4)
MTX	4 (31)/0 (0)	7 (58)/1 (8)	11 (44)/1 (4)
IVIG	4 (31)/0 (0)	3 (25)/1 (8)	6 (24)/1 (4)
LEF	3 (23)/0 (0)	0 (0)/0 (0)	3 (12)/0 (0)
ETN	1 (8)/0 (0)	0 (0)/0 (0)	1 (4)/0 (0)
Baseline laboratory values			
CK	872 \pm 1537	3744 \pm 7273	2308 \pm 5337
Aldolase	18 \pm 16	54 \pm 72	29 \pm 42
ESR	24 \pm 17	36 \pm 41	28 \pm 28
CRP	6.2 \pm 4.7	30.8 \pm 36.2	10.1 \pm 14.6

Data are missing for the following: race (n = 3), BMI (n = 3), ever smoker (n = 1), anti-SSA (n = 6), myositis (n = 2), mechanic's hands (n = 7), Raynaud phenomenon (n = 6), fever (n = 9), arthritis (n = 4), rash (n = 9), CK (n = 2), aldolase (n = 11), ESR (n = 8), CRP (n = 11). AS: antisynthetase syndrome; BWH: Brigham and Women's Hospital; UPMC: University of Pittsburgh Medical Center; RTX: rituximab; BMI: body mass index; PL-12: alanyl; PL-7: threonyl; AZA: azathioprine; MMF: mycophenolate mofetil; CYC: cyclophosphamide; MTX: methotrexate; IVIG: intravenous immunoglobulin; LEF: leflunomide; ETN: etanercept; CK: creatine kinase; ESR: erythrocyte sedimentation rate; CRP: C-reactive protein.

pneumonia (n = 1), influenza (n = 1), bronchitis (n = 1), *Clostridium difficile* colitis/urinary tract infection (n = 1), diverticulitis (n = 1), varicella zoster (n = 2), cellulitis (n = 1), and sinus infection (n = 1).

We compared the baseline and 1-year post RTX findings in the 21 subjects with either PFT and/or HRCT imaging at 1 year of followup and then analyzed subjects with up to 3-year PFT (n = 7) and/or HRCT (n = 4) followup (Table 3, Figure 2; and Supplementary Table 2, available from the authors on request). The average CT scan score at 1 year (n = 8) was stable or improved in 88% of subjects while the FVC (n = 19) was similarly stable or improved in 79%. The average TLC in all subjects (n = 7) increased from 56 \pm 13 to 64 \pm 13 at 1 year, with a statistically significant average individual TLC increase of 8.3% (p = 0.016). Similar trends

were seen at 24 months. Based on very limited 36-month followup data for only 7 subjects, there was a statistically significant average individual increase in FVC at 3 years of 21% (p = 0.016). Although the average DLCO in all subjects declined slightly from 42 \pm 17 to 36 \pm 16 at the 1-year timepoint, there was an increase at 2 years to an average of 53 \pm 26. The GC dose was stable or decreased in 88% of subjects at 1 year with an average drop of 6 mg and a statistically significant average individual decrease of 6.2 mg (p = 0.041). When comparing subjects who received repeat RTX dosing (n = 17) with those who received a 1-time dose (n = 8), those with repeat RTX dosing showed improvement in all variables, including a significant average increase in FVC and TLC of 9.3% (p = 0.0077) and 8.5% (p = 0.031), respectively (Table 4), despite starting with a lower average

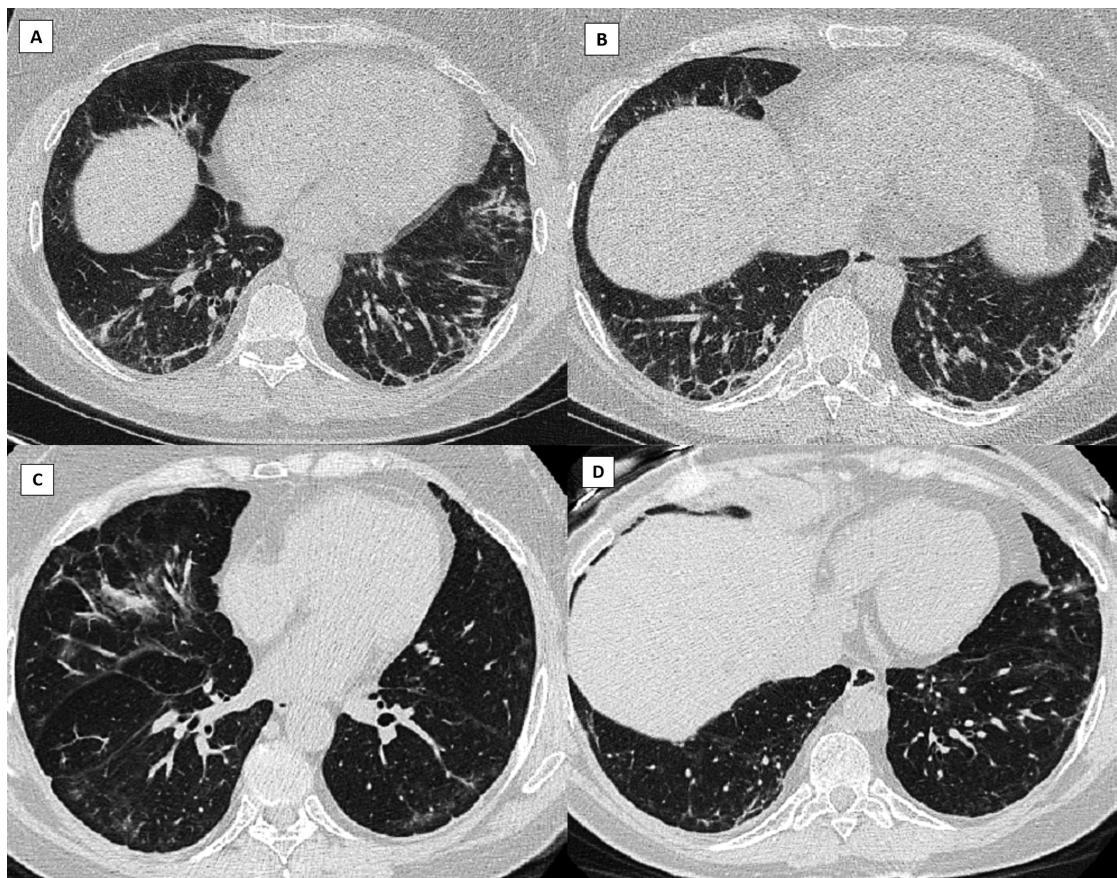


Figure 1. CT scan before and after treatment with RTX for BWH Subject 2. Pre-RTX (CT score 125.1): axial HRCT images in lung windows (A and B) demonstrate subpleural reticulation, coarse linear bands, ground-glass abnormality, and patchy consolidation within the lower lobes, lingula, and right middle lobe in a predominantly peripheral, subpleural distribution. Affected areas of lung demonstrate traction bronchiectasis and architectural distortion. No honeycombing was present, and lung volumes were mildly reduced. Pattern of interstitial pneumonitis has overlapping features of both NSIP and COP. More than 3 years post-RTX (CT score 113.7): axial HRCT images (C and D) in lung windows demonstrate definite improvement in bilateral subpleural reticulation, consolidation, and architectural distortion. Residual ground-glass opacity is present with subpleural sparing, and imaging features are consistent with a milder NSIP pattern. CT: computed tomography; RTX: rituximab; BWH: Brigham and Women's Hospital; HRCT: high-resolution CT; NSIP: nonspecific interstitial pneumonia; COP: cryptogenic organizing pneumonia.

FVC, TLC, and DLCO, although a comparison of baseline values between the 2 groups did not reach statistical significance. Those who received a single dose of RTX had a decline in most variables including a worsening CT score, decrease in FVC, and decrease in DLCO, although the numbers were too small to reach statistical significance. Overall, 100% of subjects (5/5) who had repeat RTX dosing had treatment success (defined as stabilization or improvement) regarding CT score, and 93% (13/14) had treatment success regarding FVC. In the single RTX group, those numbers declined to 67% (2/3) for CT score and 40% (2/5) for FVC (Table 4).

Although limited in their interpretability given the insufficient numbers, we did perform subgroup analyses by autoantibody status (Jo1, PL-12, PL-7), radiographic pattern (NSIP, NSIP, and COP, or fibrotic ILD), severity of lung disease (CT fibrosis score \geq or $<$ 194, FVC \geq or $<$ 50%, TLC

\geq or $<$ 50%, DLCO \geq or $<$ 35%), use of RTX as rescue therapy or initial therapy, and absence or presence of concurrent immunosuppression at the time of initiating RTX (Supplementary Table 3, available from the authors on request). Improvement with RTX was more apparent in individuals who had an NSIP pattern on CT scan, or had less severe lung disease based on the FVC or DLCO. Those in whom RTX was started owing to failure of other immunosuppressive agents and those who were taking concurrent immunosuppression also had a more notable response. However, small subject numbers limited the generalizability of the above findings and prohibited paired statistical analyses in many subgroups.

DISCUSSION

In our study of 25 AS-ILD subjects treated with RTX at 2 academic referral centers, we observed stability or

Table 3. Comparison of pre- and post-RTX CT scan score, PFT measurements, and prednisone dose for all subjects with followup at 1–3 years (n = 25). Values are mean ± SD or n (%) unless otherwise specified.

Variables	Baseline	1-yr Followup	2-yr Followup	3-yr Followup
CT scan				
Average score	194 ± 58, n = 15	195 ± 55, n = 8	187 ± 52, n = 5	161 ± 55, n = 4
Paired average Δ (p value)*	NA	–2.0 (0.84)	–0.9 (0.88)	–38.9 (0.63)
Improved/stable**	NA	2/5 (88)	0/4 (80)	1/2 (75)
PFT				
FVC, % predicted	57 ± 17, n = 22	60 ± 20, n = 19	65 ± 22, n = 12	82 ± 20, n = 7
Paired average Δ (p value)	NA	+3.7 (0.18)	+7.7 (0.15)	+21.1 (0.016)
Improved/stable**	NA	8/7 (79)	6/4 (83)	7/0 (100)
TLC, % predicted	56 ± 13, n = 7	64 ± 13, n = 7	67 ± 24, n = 5	75 ± 19, n = 2
Paired average Δ (p value)*	NA	+8.3 (0.016)	+8.6 (0.44)	+23.5 (0.50)
DLCO, % predicted	42 ± 17, n = 19	36 ± 16, n = 17	53 ± 26, n = 9	70 ± 20, n = 4
Paired average Δ (p value)*	NA	–3.34 (0.46)	+9.6, (0.074)	+14.8 (0.25)
Prednisone				
Average dose	18 ± 9, n = 20	12 ± 12, n = 17	10 ± 8, n = 6	6 ± 6, n = 4
Paired average Δ (p value)*	NA	–6.2 (0.041)	–3.3 (0.63)	–15.1 (0.13)
Improved/stable**	NA	11/4 (88)	2/2 (67)	4/0 (100)

*Paired average Δ represents difference in the mean of paired observations with the p value as calculated using the Wilcoxon signed-rank test. A p value of < 0.05 was considered statistically significant. **Improved is defined by > 10% decrease in CT score or > 10% improvement in FVC. Stable is defined by < 10% decrease but < 10% increase in CT score or < 10% increase but < 10% decline in FVC (PIOPED criteria). Failure is defined by a > 10% increase in CT score or > 10% decline in FVC. Values are compared to baseline timepoint. RTX: rituximab; CT: computed tomography; PFT: pulmonary function tests; FVC: forced vital capacity; TLC: total lung capacity; NA: not applicable; PIOPED: pulmonary embolism diagnosis.

improvement in CT imaging and/or physiologic testing (FVC, TLC, and DLCO) in most patients at 1 and 3 years of followup. Further, there was a significant steroid-sparing effect and RTX was generally well tolerated. Subgroup analyses demonstrated a benefit to repeat RTX dosing over a single RTX cycle, but in general, it seemed that the greatest benefit to RTX was seen at the 3-year followup, although the low number of patients with data at this endpoint limits the generalizability of the conclusions.

Previous studies addressing RTX for treatment of AS-ILD have also shown a favorable response^{20,21,28–34,36}, although investigations are mostly limited to case reports and retrospective studies. In small case series of patients with AS-ILD following treatment with RTX, objective improvement in PFT (FVC, TLC, and DLCO) has been noted^{20,21,28,30,33,34,35,36}, as has improvement or resolution of ground-glass opacities and stability, or improvement of fibrosis on HRCT^{20,28,30,33,35}. In 1 similar retrospective study by Andersson, *et al*, ILD extent was found to be reduced or stabilized in 21/23 patients and FVC increased by an average of 24% in 24 patients followed longitudinally for up to 60 months³³. In another small study by Marie, *et al*³⁵ of 7 AS subjects with refractory ILD treated with RTX at Day 0, Day 14, and Month 6, there was a statistically significant improvement in the median FVC/DLCO and an improvement (n = 5) or stability (n = 2) in HRCT at 1 year of followup, as well as a decrease or resolution of respiratory symptoms in all subjects. Further, these subjects had a significant decrease in the concurrent median daily prednisone dose from 20 mg/day to 9 mg/day³⁵. However, unlike our study, this was a single-center study that did not assess individual change and

had insufficient power to analyze subgroups. There has been 1 prospective trial by Allenbach, *et al*, in which 10 antisynthetase-positive subjects who had failed conventional treatment received RTX at Day 0, Day 15, and Month 6³¹. That study demonstrated an improvement or stabilization of ILD as measured by FVC and DLCO in 9/10 subjects and either a > 50% decrease in steroid dose or discontinuation of ≥ 1 other immunomodulatory/immunosuppressive drug in 6/10 patients. Only 1 subject had a reduction in interstitial infiltrates on CT scan, and improvement in ILD (5/10 subjects) was defined as a 10% increase in FVC or 15% increase in DLCO, while stabilization (4/10 subjects) was defined as a < 10% decrease in FVC or < 15% decrease in DLCO at Month 12.

Although our study and previous case series and retrospective studies have reported favorable responses in AS-ILD treated with RTX, the mechanism of its therapeutic effect is uncertain. It is unclear whether the B cell-deleting mechanism attributed to RTX represents the mechanism of action leading to its effectiveness in AS-ILD. Studies in other immune-mediated diseases suggest that RTX may lead to normalization of autoreactive T cells^{46,47}, and the repopulation of the B cells following RTX may be antigenically naive, transitional B cells, suggesting a possible “resetting” of the immune system that contributes to a therapeutic effect⁴⁸. A retrospective study of 50 systemic autoimmune rheumatic disease-related ILD patients with myositis improved their PFT following B cell depletion²⁷, suggesting a key pathogenic role of T cells in myositis-ILD. Future mechanistic studies should address the pathogenic potential of specific antibodies in AS and whether B cell depletion after

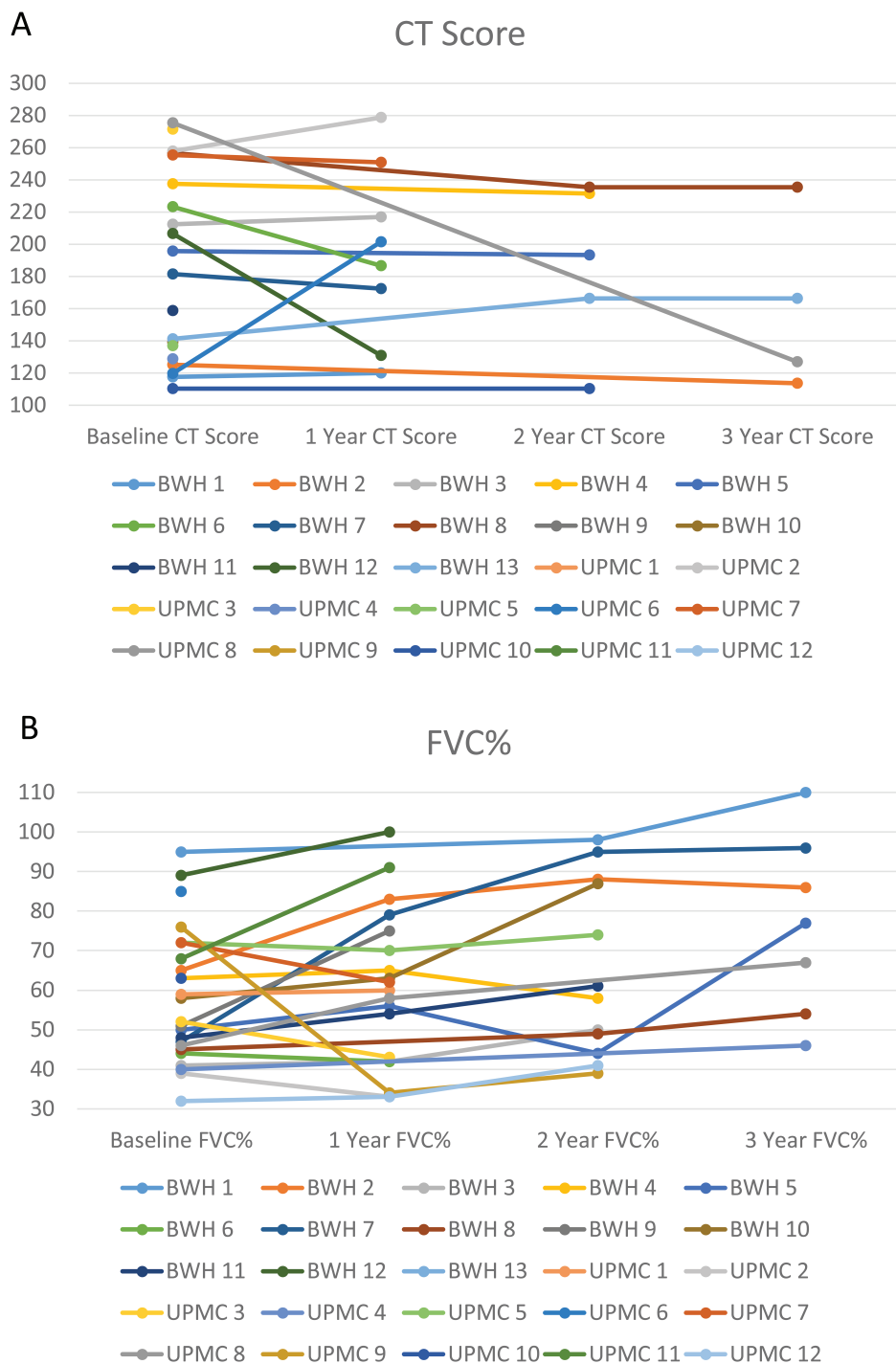


Figure 2. Graph of (A) CT score, and (B) FVC by subject over 3 years of followup. (A) Line graph of CT scores demonstrating stability or improvement (decrease in CT score) in > 75% of subjects over the 3 years of followup. (B) Line graph of FVC demonstrating stability or improvement in > 79% of subjects over the 3 years of followup. CT: computed tomography; FVC%: forced vital capacity, % predicted; BWH: Brigham and Women's Hospital; UPMC: University of Pittsburgh Medical Center.

RTX is responsible for its favorable effect in AS-ILD. In addition, the future elucidation of antisynthetase auto-antibody-specific differences or whether the presence of other

biomarkers is useful in predicting a response to B cell depletion should be investigated to guide treatment algorithms in AS-ILD.

Table 4. Comparison of pre- and post-RTX CT scan score and PFT measurements comparing averages at baseline and 1 year with the average individual change (p value) in subjects with repeat RTX dosing versus subjects with a 1-time RTX dose. Values are mean \pm SD or n (%) unless otherwise specified.

Variables	Repeat RTX Dosing, n = 17		One-time RTX Dosing, n = 8	
	Baseline	1-yr Followup	Baseline	1-yr Followup
CT scan	n = 5		n = 3	
Average score	171 \pm 42	152 \pm 40	211 \pm 79	244 \pm 39
Paired average Δ (p value)*	-22.9 (0.31)		32.8 (0.50)	
Improved/Stable**	2/3 (100)		0/2 (67)	
Failure***	0 (0)		1 (33)	
PFT	n = 14		n = 5	
FVC	n = 14		n = 5	
FVC, % predicted	54 \pm 14	63 \pm 20	64 \pm 15	52 \pm 17
Paired average Δ (p value)	+9.3 (0.0077)		-11.8 (0.13)	
Improved/Stable**	8/5 (93)		0/2 (40)	
Failure***	1 (7)		3 (60)	
TLC	n = 6		n = 1	
TLC, % predicted	53 \pm 13	62 \pm 14	70	77
Paired average Δ (p value)	+8.5 (0.031)		NA	
DLCO	n = 12		n = 5	
DLCO, % predicted	38 \pm 12	39 \pm 18	43 \pm 16	29 \pm 8
Paired average Δ (p value)	+1.2 (0.59)		-14.2 (0.13)	

*Paired average Δ represents difference in the mean of paired observations with the p value as calculated using the Wilcoxon signed-rank test. A p value of < 0.05 was considered statistically significant. **Improved is defined by $> 10\%$ decrease in CT score or $> 10\%$ improvement in FVC. Stable is defined by $< 10\%$ decrease but $< 10\%$ increase in CT score, or $< 10\%$ increase but $< 10\%$ decline in FVC (PIOPED criteria). ***Failure is defined by a $> 10\%$ increase in CT score or a $> 10\%$ decline in FVC. Values are compared to baseline timepoint. RTX: rituximab; CT: computed tomography; PFT: pulmonary function tests; FVC: forced vital capacity; TLC: total lung capacity; NA: not applicable; PIOPED: pulmonary embolism diagnosis.

In our study, all patients in the BWH cohort received repeat RTX dosing on average every 6 months, whereas most patients in the UPMC cohort received only a 1-time dose (n = 8), with only 4 receiving repeat dosing at variable intervals. A better objective response in FVC and TLC was noted in patients with repeat RTX dosing, with a trend toward improved HRCT scores. However, the sample size is too small to draw any significant conclusions and there may be inherent bias because the reason for redosing in the UPMC cohort could not be ascertained. This raises the question of whether those subjects who were redosed were the patients who responded favorably, while those who were not redosed did not respond to initial RTX dosing. In general, there is no standard of care regarding the frequency of RTX dosing in myositis given the paucity of data and the variability of third-party payer approval. One retrospective study demonstrated that in 11 AS patients with severe, progressive ILD refractory to other immunotherapy, RTX appeared to stabilize and/or improve ILD in 7 patients during the first 6 months only²⁸, suggesting that frequent dosing versus 1-time RTX dosing was more effective. This observation, our findings, especially the significant improvement at 3 years, and the pathophysiologic mechanisms discussed above suggest that optimal RTX dosing may require additional dosing rather than an initial cycle of RTX, and that RTX may take a long time to have an effect.

Our study has a few important limitations. First, the small sample size limits the discovery of statistically significant findings, particularly regarding the interpretability of subgroup analyses. Second, the retrospective chart review design is limited in that not all subjects have data at all timepoints, and there are very few subjects with 3-year followup, potentially introducing a source of bias. However, all subjects had ≥ 1 HRCT or PFT done > 1 year from the time of initial RTX dosing. To address this limitation, the difference in the mean of paired observations was presented to allow for paired analyses. Third, there was variability in the dosing of RTX at the 2 independent centers, which complicates composite data analyses. Fourth, there is limited generalizability given that our study was conducted in 2 clinical centers. Fifth, there is an inherent difficulty in assessing radiographic outcomes in ILD associated with AS, because there is no validated specific scoring system for autoimmune ILD. Although numerous HRCT severity scoring systems exist for evaluation of patients with IPF, no specific scoring system has yet been proposed for patients with ILD in the setting of AS. Given the presence of multiple radiologic patterns (UIP, NSIP, and COP) in this subset of patients, we adapted a CT scoring system proposed by Fujimoto, *et al*⁴¹ and Ichikado, *et al*⁴², which has previously been used for quantitative assessment of HRCT abnormalities in a variety of clinical scenarios ranging from AIP, acute respi-

ratory distress syndrome, and IPF⁶. The exclusion of the UPMC subject who proceeded to transplant after 10 months might have biased the data analysis in favor of RTX, given that this subject received a lung transplant after declining over the 10 months after RTX. Additional prospective, randomized, multicenter trials should address these limitations.

RTX may play a therapeutic role in patients with AS-ILD. Our study has added a significant body of data to the existing information on RTX in AS-ILD, and suggests that further clinical investigation is warranted into the effectiveness of RTX as a treatment for AS-ILD, pathophysiologic mechanisms of action, ideal dosing interval, and length of treatment⁴.

REFERENCES

- Katzap E, Barilla-LaBarca ML, Marder G. Antisynthetase syndrome. *Curr Rheumatol Rep* 2011;13:175-81.
- Bohan A, Peter JB. Polymyositis and dermatomyositis (first of two parts). *N Engl J Med* 1975;292:344-7.
- Connors GR, Christopher-Stine L, Oddis CV, Danoff SK. Interstitial lung disease associated with the idiopathic inflammatory myopathies: What progress has been made in the past 35 years? *Chest* 2010;138:1464-74.
- Hallowell RW, Danoff SK. Interstitial lung disease associated with the idiopathic inflammatory myopathies and the antisynthetase syndrome: Recent advances. *Curr Opin Rheumatol* 2014;26:684-9.
- Marie I, Hatron PY, Dominique S, Cherin P, Mouthon L, Menard JF. Short-term and long-term outcomes of interstitial lung disease in polymyositis and dermatomyositis: A series of 107 patients. *Arthritis Rheum* 2011;63:3439-47.
- Debray MP, Borie R, Revel MP, Naccache JM, Khalil A, Toper C, et al. Interstitial lung disease in anti-synthetase syndrome: Initial and follow-up CT findings. *Eur J Radiol* 2015;84:516-23.
- Aggarwal R, Cassidy E, Fertig N, Koontz DC, Lucas M, Ascherman DP, et al. Patients with non-Jo-1 anti-tRNA-synthetase autoantibodies have worse survival than Jo-1 positive patients. *Ann Rheum Dis* 2014;73:227-32.
- Marie I, Josse S, Hatron PY, Dominique S, Hachulla E, Janvresse A, et al. Interstitial lung disease in anti-jo-1 patients with antisynthetase syndrome. *Arthritis Care Res* 2013;65:800-8.
- Tomonaga M, Sakamoto N, Ishimatsu Y, Kakugawa T, Harada T, Nakashima S, et al. Comparison of pulmonary involvement between patients expressing anti-pl-7 and anti-jo-1 antibodies. *Lung* 2015;193:79-83.
- Aggarwal R, Oddis CV. Therapeutic approaches in myositis. *Curr Rheumatol Rep* 2011;13:182-91.
- Perosa F, Prete M, Racanelli V, Dammacco F. Cd20-depleting therapy in autoimmune diseases: From basic research to the clinic. *J Intern Med* 2010;267:260-77.
- Leandro MJ, Cambridge G, Ehrenstein MR, Edwards JC. Reconstitution of peripheral blood b cells after depletion with rituximab in patients with rheumatoid arthritis. *Arthritis Rheum* 2006;54:613-20.
- Edwards JC, Szczepanski L, Szechinski J, Filipowicz-Sosnowska A, Emery P, Close DR, et al. Efficacy of b-cell-targeted therapy with rituximab in patients with rheumatoid arthritis. *N Engl J Med* 2004;350:2572-81.
- Emery P, Fleischmann R, Filipowicz-Sosnowska A, Schechtman J, Szczepanski L, Kavanaugh A, et al. The efficacy and safety of rituximab in patients with active rheumatoid arthritis despite methotrexate treatment: Results of a phase iib randomized, double-blind, placebo-controlled, dose-ranging trial. *Arthritis Rheum* 2006;54:1390-400.
- Cohen SB, Emery P, Greenwald MW, Dougados M, Furie RA, Genovese MC, et al. Rituximab for rheumatoid arthritis refractory to anti-tumor necrosis factor therapy: Results of a multicenter, randomized, double-blind, placebo-controlled, phase III trial evaluating primary efficacy and safety at twenty-four weeks. *Arthritis Rheum* 2006;54:2793-806.
- Stone JH, Merkel PA, Spiera R, Seo P, Langford CA, Hoffman GS, et al. Rituximab versus cyclophosphamide for ANCA-associated vasculitis. *N Engl J Med* 2010;363:221-32.
- Arnold DM, Dentali F, Crowther MA, Meyer RM, Cook RJ, Sigouin C, et al. Systematic review: Efficacy and safety of rituximab for adults with idiopathic thrombocytopenic purpura. *Ann Intern Med* 2007;146:25-33.
- Gottenberg JE, Guillemin L, Lambotte O, Combe B, Allanore Y, Cantagrel A, et al. Tolerance and short term efficacy of rituximab in 43 patients with systemic autoimmune diseases. *Ann Rheum Dis* 2005;64:913-20.
- Oddis CV, Reed AM, Aggarwal R, Rider LG, Ascherman DP, Levesque MC, et al. Rituximab in the treatment of refractory adult and juvenile dermatomyositis and adult polymyositis: A randomized, placebo-phase trial. *Arthritis Rheum* 2013;65:314-24.
- Nalotto L, Iaccarino L, Zen M, Gatto M, Borella E, Domenighetti M, et al. Rituximab in refractory idiopathic inflammatory myopathies and antisynthetase syndrome: Personal experience and review of the literature. *Immunol Res* 2013;56:362-70.
- Unger L, Kampf S, Luthke K, Aringer M. Rituximab therapy in patients with refractory dermatomyositis or polymyositis: Differential effects in a real-life population. *Rheumatology* 2014;53:1630-8.
- Fasano S, Gordon P, Hajji R, Loyo E, Isenberg DA. Rituximab in the treatment of inflammatory myopathies: A review. *Rheumatology* 2017;56:26-36.
- Aggarwal R, Bandos A, Reed AM, Ascherman DP, Barohn RJ, Feldman BM, et al. Predictors of clinical improvement in rituximab-treated refractory adult and juvenile dermatomyositis and adult polymyositis. *Arthritis Rheumatol* 2014;66:740-9.
- Mahler EA, Blom M, Voermans NC, van Engelen BG, van Riel PL, Vonk MC. Rituximab treatment in patients with refractory inflammatory myopathies. *Rheumatology* 2011;50:2206-13.
- Jordan S, Distler JH, Maurer B, Huscher D, van Laar JM, Allanore Y, et al. Effects and safety of rituximab in systemic sclerosis: An analysis from the European Scleroderma Trial and Research (EUSTAR) group. *Ann Rheum Dis* 2015;74:1188-94.
- Daoussis D, Liossis SN, Tsamandas AC, Kalogeropoulou C, Kazantzis A, Korfiatis P, et al. Is there a role for B-cell depletion as therapy for scleroderma? A case report and review of the literature. *Semin Arthritis Rheum* 2010;40:127-36.
- Keir GJ, Maher TM, Ming D, Abdullah R, de Lauretis A, Wickremasinghe M, et al. Rituximab in severe, treatment-refractory interstitial lung disease. *Respirology* 2014;19:353-9.
- Sem M, Molberg O, Lund MB, Gran JT. Rituximab treatment of the anti-synthetase syndrome: A retrospective case series. *Rheumatology* 2009;48:968-71.
- Dasa O, Ruzieh M, Oraibi O. Successful treatment of life-threatening interstitial lung disease secondary to antisynthetase syndrome using rituximab: A case report and review of the literature. *Am J Ther* 2016;23:e639-45.
- Bauhammer J, Blank N, Max R, Lorenz HM, Wagner U, Krause D, et al. Rituximab in the treatment of jo1 antibody-associated antisynthetase syndrome: Anti-ro52 positivity as a marker for severity and treatment response. *J Rheumatol* 2016;43:1566-74.
- Allenbach Y, Guiguet M, Rigolet A, Marie I, Hachulla E, Drouot L, et al. Efficacy of rituximab in refractory inflammatory myopathies associated with anti-synthetase auto-antibodies: An open-label, phase II trial. *PLoS One* 2015;10:e0133702.

32. Vandenbroucke E, Grutters JC, Altenburg J, Boersma WG, ter Borg EJ, van den Bosch JM. Rituximab in life threatening antisynthetase syndrome. *Rheumatol Int* 2009;29:1499-502.
33. Andersson H, Sem M, Lund MB, Aalokken TM, Gunther A, Walle-Hansen R, et al. Long-term experience with rituximab in anti-synthetase syndrome-related interstitial lung disease. *Rheumatology* 2015;54:1420-8.
34. Lepri G, Avouac J, Airo P, Anguita Santos F, Bellando-Randone S, Blagojevic J, et al. Effects of rituximab in connective tissue disorders related interstitial lung disease. *Clin Exp Rheumatol* 2016;34 Suppl 100:181-5.
35. Marie I, Dominique S, Janvresse A, Levesque H, Menard JF. Rituximab therapy for refractory interstitial lung disease related to antisynthetase syndrome. *Respir Med* 2012;106:581-7.
36. Sharp C, McCabe M, Dodds N, Edey A, Mayers L, Adamali H, et al. Rituximab in autoimmune connective tissue disease-associated interstitial lung disease. *Rheumatology* 2016;55:1318-24.
37. Belhassen-Garcia M, Rabano-Gutierrez A, Velasco-Tirado V, Romero-Alegria A, Perez-Garcia ML, Martin-Oterino JA. Atypical progressive multifocal leukoencephalopathy in a patient with antisynthetase syndrome. *Intern Med* 2015;54:519-24.
38. Hankinson JL, Odencrantz JR, Fedan KB. Spirometric reference values from a sample of the general U.S. population. *Am J Respir Crit Care Med* 1999;159:179-87.
39. Crapo RO, Morris AH, Clayton PD, Nixon CR. Lung volumes in healthy nonsmoking adults. *Bull Eur Physiopathol Respir* 1982;18:419-25.
40. Cotes JE, Chinn DJ, Quanjer PH, Roca J, Yernault JC. Standardization of the measurement of transfer factor (diffusing capacity). Report Working Party Standardization of Lung Function Tests, European Community for Steel and Coal. Official statement of the European Respiratory Society. *Eur Respir J Suppl* 1993;16:41-52.
41. Fujimoto K, Taniguchi H, Johkoh T, Kondoh Y, Ichikado K, Sumikawa H, et al. Acute exacerbation of idiopathic pulmonary fibrosis: High-resolution CT scores predict mortality. *Eur Radiol* 2012;22:83-92.
42. Ichikado K, Suga M, Müller NL, Taniguchi H, Kondoh Y, Akira M, et al. Acute interstitial pneumonia: Comparison of high-resolution computed tomography findings between survivors and nonsurvivors. *Am J Respir Crit Care Med* 2002;165:1551-6.
43. Collard HR, King TE Jr, Bartelson BB, Vourlekis JS, Schwarz MI, Brown KK. Changes in clinical and physiologic variables predict survival in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2003;168:538-42.
44. Ryerson CJ, Vittinghoff E, Ley B, Lee JS, Mooney JJ, Jones KD, et al. Predicting survival across chronic interstitial lung disease: The ILD-GAP model. *Chest* 2014;145:723-8.
45. Berry CE, Wise RA. Interpretation of pulmonary function test: Issues and controversies. *Clin Rev Allergy Immunol* 2009; 37:173-80.
46. Stasi R, Del Poeta G, Stipa E, Evangelista ML, Trawinska MM, Cooper N, et al. Response to B-cell depleting therapy with rituximab reverts the abnormalities of T-cell subsets in patients with idiopathic thrombocytopenic purpura. *Blood* 2007;110:2924-30.
47. Cutler C, Miklos D, Kim HT, Treister N, Woo SB, Bienfang D, et al. Rituximab for steroid-refractory chronic graft-versus-host disease. *Blood* 2006;108:756-62.
48. Anolik JH, Barnard J, Owen T, Zheng B, Kemshetti S, Looney RJ, et al. Delayed memory B cell recovery in peripheral blood and lymphoid tissue in systemic lupus erythematosus after B cell depletion therapy. *Arthritis Rheum* 2007;56:3044-56.