

Biologic Therapy for Systemic Sclerosis: A Systematic Review

VEERAPONG PHUMETHUM, SHAHIN JAMAL, and SINDHU R. JOHNSON

ABSTRACT. Objective. Biologic agents are increasingly used in the rheumatic diseases. Their role in patients with systemic sclerosis (SSc) is uncertain. Our aim was to evaluate the effectiveness and safety of biologic agents in SSc. We review the evidence for the use of biologic agents to improve inflammatory arthritis, disability, and skin score, and we review adverse effects with biologic agents in patients with SSc.

Methods. A systematic literature review was performed to identify studies evaluating the use of biologic agents in SSc. Medline, Embase, CINAHL, and Cochrane Database of Systematic Reviews were searched. A standardized abstraction form was used to extract biologic agent, study design, sample size, treatment effect, and adverse effects.

Results. A total of 23 studies from 1413 citations were evaluated. Three studies evaluated infliximab, 3 evaluated etanercept, 3 evaluated antithymocyte globulin, 3 evaluated imatinib, 6 evaluated rituximab, and 1 study each evaluated interferon- γ (IFN- γ), IFN- α , relaxin, delipidated, deglycolipidated *Mycobacterium vaccae*, human anti-transforming growth factor β 1 antibody, and oral type I collagen. Studies of etanercept and infliximab suggest improvements in inflammatory arthritis and Health Assessment Questionnaire Disability Index (HAQ-DI). None of the other biologic agents demonstrated reproducible, statistically significant improvements in joint count, HAQ-DI, or skin score.

Conclusion. Anti-tumor necrosis factor- α agents may improve inflammatory arthritis and disability in SSc. The effect on skin score is uncertain. Adequately powered trials are needed to evaluate efficacy, and longitudinal studies are needed to evaluate longterm safety of these agents in SSc. (J Rheumatol First Release Nov 1 2010; doi:10.3899/jrheum.100361)

Key Indexing Terms:
SCLERODERMA

SYSTEMIC SCLEROSIS
ANTI-TUMOR NECROSIS FACTOR- α

BIOLOGIC

Systemic sclerosis (SSc) is a disease characterized by immune activation and inflammation that leads to fibrosis. It commonly affects the skin, blood vessels, and internal organs. Inflammatory arthritis is an increasingly recognized manifestation of SSc, with a prevalence of 10% to 25%^{1,2}. Using the European League Against Rheumatism Scleroderma Trials and Research Group database, the prevalence of synovitis is 16% (1191/7286)³. Synovial biopsy

studies have shown histological evidence of inflammation with lymphocytic and plasma cell infiltration. On electron microscopy there are prominent microvascular alterations and abundant immature collagen fibrils. Late in the course of the disease there can be evidence of severe synovial fibrosis⁴. In a retrospective cohort study evaluating the use of magnetic resonance imaging (MRI) in patients with SSc who have joint pain, 59% of patients had synovitis, erosions, joint effusion, or tenosynovitis⁵. Rheumatoid factor and anticyclic citrullinated peptide (anti-CCP) antibodies were present in only 40% and 11% of patients with inflammatory MRI findings, respectively⁵. Whether erosive, inflammatory arthritis is a direct manifestation of SSc or reflects an overlap syndrome with rheumatoid arthritis (RA) is controversial³. The low frequency of anti-CCP antibodies in multiple studies of patients with SSc who have an erosive inflammatory arthritis (estimates ranging from 1.5% to 11%) suggest that the condition is a direct manifestation of SSc^{5,6,7}.

Tumor necrosis factor (TNF) is an important proinflammatory cytokine that has been shown to play a pivotal role in the pathogenesis of other autoimmune diseases including RA, psoriatic arthritis (PsA), and ankylosing spondylitis⁸. However, its role in the pathogenesis of SSc is unclear. *In*

From the Prapokklao Hospital, Chantaburi, Thailand; Scleroderma Clinic, Mount Sinai Hospital; Division of Rheumatology, St. Michael's Hospital; and Division of Rheumatology, Toronto Western Hospital, University Health Network, Department of Medicine, Faculty of Medicine, University of Toronto, Toronto, Ontario, Canada.

Dr. S.R. Johnson is supported by a Canadian Institutes of Health Research Clinician Scientist Award and the Norton-Evans Fund for Scleroderma Research.

V. Phumethum, MD, FRCPC, Scleroderma Clinic, Mount Sinai Hospital, Prapokklao Hospital; S. Jamal, MD, MSc, FRCPC, Division of Rheumatology, St. Michael's Hospital; S.R. Johnson, MD, FRCPC, Scleroderma Clinic, Mount Sinai Hospital, Division of Rheumatology, Toronto Western Hospital, Department of Medicine, University of Toronto.

Address correspondence to Dr. S.R. Johnson, Division of Rheumatology, Ground Floor, East Wing, Toronto Western Hospital, 399 Bathurst Street, Toronto, Ontario M5T 2S8, Canada. E-mail: Sindhu.Johnson@uhn.on.ca
Accepted for publication September 7, 2010.

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2010. All rights reserved.

in vivo animal studies demonstrate that antagonists of TNF- α can significantly reduce the accumulation of extracellular matrix and prevent the development of fibrosis in mouse models of silica-induced and bleomycin-induced pulmonary fibrosis^{9,10}. Studies in mice lacking TNF-I or both TNF receptors demonstrated that inhibition of TNF signaling can prevent organ fibrosis^{11,12,13}. Conversely, the majority of *in vitro* studies show antifibrotic effects of TNF, in that it suppresses the production of collagen^{14,15}, reduces the expression of tissue inhibitor of metalloproteinases¹⁶, and stimulates the release of matrix metalloproteinase^{17,18}, thereby preventing the accumulation of extracellular matrix and preventing the development of fibrosis.

The efficacy of TNF antagonists in inflammatory diseases such as RA, spondyloarthropathies, and Crohn's disease has been demonstrated in clinical trials^{19,20,21}. Despite much interest in the use of anti-TNF agents in the treatment of SSc, there is no consensus on their potential therapeutic benefit. There is also some concern about using anti-TNF therapy in SSc because of a potential for worsening disease. A case report indicated that adalimumab was associated with fibrosing alveolitis in a patient with SSc²². Similarly, infliximab has been associated with fibrosing alveolitis in patients with RA²³. Further, anti-TNF agents have also been associated with the development of autoantibodies and drug-induced lupus erythematosus^{24,25}, which may be a potential concern in patients with SSc.

Given the potential benefits of biologic therapies in SSc and their demonstrable efficacy in other rheumatic diseases, our systematic review aims to evaluate the effectiveness and safety of biologic therapies in patients with SSc. In particular, we reviewed the evidence for the use of biologic agents to improve inflammatory arthritis, disability, and skin score, and reviewed the adverse effect profile of these agents in SSc.

MATERIALS AND METHODS

Data sources and search strategy. One investigator and an information specialist through the University Health Network Library Services independently performed the literature search. Sources searched were Ovid Medline (1950 to June week 5, 2010), Embase (1980 to week 26, 2010), CINAHL (1982 to week 26, 2010), and the Cochrane Database of Systematic Reviews (inception to first quarter, 2010). The following key words with mapping of term to subject heading were used in the database searches: [(systemic sclerosis or scleroderma or skin sclerosis or sclerema or scleroderma or en coup de sabre or morphea or dermatosclerosis) and (anti-TNF or mhr 24 or Orenzia or Ctl4 immunoglobulin or Ctl4Ig or Ctl4 Ig or Bms 188667 or Abatacept or Rituxin or Mabthera or Idec C2b8 or rituximab or Rituxan or Anakinra or Kineret or Trudexa or Monoclonal Antibody D2e7 or Humira or adalimumab or Tnr001 or Tnr 001 or Enbrel or etanercept or remicaide or Revellex or avakine or infliximab or immune serum or antisera or immune sera or biologic(s) therapy or biologic(s) intervention or biological product or biologic agent(s) or biologic(s) or tnfalpa or tnf or cachectin or tumour necrosis factor(s) or tumor necrosis factor(s)]. The search was limited to human studies but not limited to the English language. The results of the 2 independent searches were compared to ensure completeness.

Study selection. Titles and abstracts were screened to identify studies that

evaluated the use of biologic agents in SSc. Using the US National Institutes of Health definition, biologic agents were defined as substances that are made from a living organism or its products and are used in the prevention, diagnosis, or treatment of cancer and other diseases. Biologic agents may include antibodies, interleukins, and vaccines. Eligible studies were required to report joint count (tender or swollen joint count), Health Assessment Questionnaire Disability Index (HAQ-DI) score, skin score (using any measure of scleroderma skin assessment), and adverse events as outcomes. Studies were ineligible if they were animal studies, review articles, included individuals < 16 years old, or did not report any of the outcome measures of interest. The reference list of selected articles was hand-searched for relevant publications.

Data abstraction. Two reviewers independently abstracted the following data onto standardized forms: study design (observational study or randomized trial), sample size, treatment(s), tender joint count, swollen joint count, HAQ-DI score, skin score, method of skin assessment, and adverse events. The reviewers were blinded to the names of study authors, institutions, and journals when performing data abstraction. All disagreements were resolved by consensus.

RESULTS

Search results. The search identified 1413 citations; however, screening of titles and abstracts resulted in the exclusion of 1390 citations, leaving 23 articles for full review. Citations were excluded because they were review articles, involved patients with rheumatic diseases other than SSc, were animal studies, or did not describe use of a biologic agent. Of the 23 citations selected, 5 were cohort studies^{26,27,28,29,48} reporting on the use of anti-TNF therapy for treatment in SSc, and 18 studies described the use of other biologic agents in SSc. These included the evaluation of antithymocyte globulin (ATG)^{30,31,32}, imatinib mesylate^{33,34,35}, interferon- γ (IFN- γ)³⁶, IFN- α ³⁷, recombinant human relaxin³⁸, delipidated, deglycolipidated *Mycobacterium vaccae* (PVAC)³⁹, recombinant human anti-transforming growth factor- β 1 (TGF- β 1) antibody⁴⁰, type I collagen⁴¹, and rituximab^{42,43,44,45,46,47}.

Anti-TNF therapy in SSc. Three observational studies evaluated infliximab^{27,28,48} and 3 observational studies evaluated etanercept^{26,29}. No randomized trials evaluating the use of anti-TNF therapy in SSc were identified (Table 1).

Lam, *et al* report a retrospective cohort study of etanercept 25 mg twice weekly or 50 mg once weekly in 18 women with SSc²⁶. The primary outcome was a significant decrease in synovitis on examination and complete resolution of joint pain. The mean age was 44 years (range 25–71 yrs) with a mean duration of therapy of 30 months (range 2–66 mo). Fifteen (83%) of 18 patients treated with etanercept had a significant decrease in signs of inflammation or synovitis and resolution of joint symptoms. The mean HAQ-DI score decreased from 1.08 ± 0.70 to 0.74 ± 0.56 ($p = 0.13$). The mean Rodnan skin score decreased from 6.63 ± 6.35 to 3.94 ± 2.38 ($p = 0.12$). There were no opportunistic infections, anaphylaxis, hospitalizations, or deaths attributed to etanercept therapy. One patient developed a lupus-like reaction and another had a significant decline in lung function.

Table 1. Summary of studies evaluating anti-TNF agents in systemic sclerosis.

Study Characteristics	Lam ²⁶	Denton ²⁸	Bosello ²⁷	Ellman ²⁹	Marie ⁴⁸
Anti-TNF agent	Etanercept	Infliximab	Infliximab, Etanercept	Etanercept	Infliximab
Study design	Observational	Observational	Observational	Observational	Observational
Sample size, n	18	16	4	10, 1 discontinued after 4 wks	1
Country	USA	UK	Italy	USA	France
Study/treatment duration	Mean 30 mo (range 2–66 mo)	26 wks	6 mo	6 mo	2 wks
Other immunosuppressive medication	MTX 9/18 HCQ 5/18 Pred 9/18 Minocycline 2/18	None	MTX with infliximab	Not specified	Pred
Outcome					
Skin score	Improved, not significant ^a	No effect ^d	Improved, not significant ^{**}	4/9 (44%) improvement; 5/9 unchanged	NR
HAQ-DI	Improved, not significant ^b	Improved, not significant [*]	Improved, not significant [†]	1.8 to 1.57 ^{††}	NR
Joint count	Improved ^c	NR	NR	NR	NR
Adverse events	Lupus-like reaction 1/18, lung function decline 1/18	Infusion reactions, 44%	Not specified	n = 1 digital ischemia	<i>A. meyeri</i> pneumonia

^a Mean Rodnan skin score (MRSS) decreased from 6.63 ± 6.35 to 3.94 ± 2.38 (p = 0.12). ^b Mean HAQ-DI score decreased from 1.08 ± 0.70 to 0.74 ± 0.56 (p = 0.13). ^c 15/18 (83%) patients treated with etanercept had a significant decrease in signs of inflammation/synovitis and resolution of joint symptoms. ^d No significant change in median (range) baseline MRSS 26 (11–45) and Week 26 MRSS 22 (6–48). * Nonsignificant improvement in median (range) baseline, HAQ-DI 1.63 (0–3), and Week 26, 1.5 (0–2.63) (p value not given). ** Baseline mean MRSS 17.75, 6-month MRSS 8.25 (p = 0.22). [†] Baseline mean HAQ-DI 2.02, 6-month HAQ-DI 1.25 (p = 0.09). ^{††} p value not given. TNF- α : tumor necrosis factor- α ; MTX: methotrexate; HCQ: hydroxychloroquine; Pred: prednisone; NR: not reported; HAQ-DI: Health Assessment Questionnaire-Disability Index.

Denton, *et al* report an observational study of infliximab 5 mg/kg given at Weeks 0, 2, 6, 14, and 22 in 16 patients with diffuse SSc²⁸. Patients were included who had an increase of > 10% in skin score or an increase of 4 skin score units over a maximum 3-month period. Mean disease duration was 25.7 months (SD 19). The median baseline 17-site modified Rodnan skin score (MRSS) was 26. There was no significant change at 26 weeks (median MRSS 22, range 6–48). The median HAQ-DI score decreased from 1.63 (range 0–3) to 1.5 (range 0–2.63) at 26 weeks, and was reported as not statistically significant (p value not given). Seven (44%) of 16 subjects developed infusion reactions. Anti-infliximab antibodies developed in 5 patients and were associated with suspected infusion reactions.

Bosello, *et al*²⁷ report 4 patients with SSc who were treated with infliximab 3 mg/kg administered at Weeks 0, 2, 6, and 14 with methotrexate (MTX), followed by etanercept 25 mg subcutaneously (SC) twice weekly without concomitant MTX. Infliximab was switched to etanercept to minimize patients' time away from work. Individual baseline MRSS of 35, 12, 16, and 8 improved at 6 months with MRSS of 16, 7, 7, and 3, respectively. Baseline HAQ-DI scores of 1.575, 2.5, 2.25, and 1.75 improved at 6 months with HAQ-DI

scores of 0.375, 1.5, 1.75, and 1.375, respectively. No adverse effects were reported with either infliximab or etanercept.

Ellman, *et al* report use of etanercept 25 mg SC twice weekly for 6 months in 9 patients with diffuse SSc²⁹. Four of the patients had a 44% improvement in the Rodnan skin score and 5 remained unchanged. The HAQ-DI score improved from 1.8 to 1.57. One patient discontinued therapy after 4 weeks because of new digital ischemia, while 3 patients with digital ulcers noted improvement.

Marie, *et al*⁴⁸ report the case of a 44-year-old woman with diffuse SSc and erosive, anti-CCP-positive polyarthritis treated with 2 doses of infliximab (3 mg/kg). One month after the second infusion she developed *Actinomyces meyeri* pneumonia. The effects of infliximab on joint count, HAQ, or skin score were not reported.

Other biologic agents in SSc. Three studies report use of ATG^{30,31,32} and 3 report use of imatinib mesylate^{33,34,35} in SSc (Table 2). Five observational studies^{42,43,44,45,47} and 1 randomized trial⁴⁶ report the use of rituximab in SSc (Table 3).

Grassegger, *et al*³⁶ report a multicenter trial of recombinant IFN- γ 100 μ g SC thrice weekly for 12 months in 44 patients. Of them, 27 patients were randomized to treatment

Table 2. Summary of studies evaluating antithymocyte globulin (AG) and imatinib biologic in systemic sclerosis.

Study Characteristics	Matteson ³⁰	Balaban ³²	Stratton ³¹	van Daele ³³	Sfikakis ³⁴	Chung ³⁵
Agent	AG	AG	AG	Imatinib	Imatinib	Imatinib
Study design	Observational	Observational	Observational	Observational	Observational	Observational
Sample size, n	10	2	13	1	1	2
Country	USA	USA	UK	Netherlands	UK	USA
Outcomes						
Skin score	n = 2 improved* n = 6 stabilized*	Improved [†]	Improved ^{††}	Improved ^a	Improved ^b	Improved ^c
HAQ-DI	n = 2 improved** n = 3 stabilized**	NR	NR	NR	NR	NR
Joint count	NR	NR	NR	NR	NR	NR
Adverse events (AE)	Urticaria, n = 1, serum sickness, n = 1	No serious AE	Serum sickness, n = 5	No serious AE	No serious AE	No serious AE

* Improvement defined as $\geq 25\%$ decrease in skin score; stabilized defined as $\pm 25\%$ baseline skin score. ** Improvement defined as ≥ 0.25 -point decrease, stabilized defined as ± 0.25 -point baseline. [†] Total skin score improved from 25 to 14 in Patient 1, and 22 to 7 in Patient 2. ^{††} Mean Rodnan skin score (MRSS) improved from 28 to 17 after 12 months ($p < 0.01$). ^a MRSS improved from 18 to 13 at 3 months. ^b MRSS improved from 44 at baseline to 33 at 6 weeks and 28 at 6 months. ^c MRSS improved from 36 to 21 at 3 months in Patient 1; 36 to 20 at 6 months in Patient 2. NR: not reported; HAQ-DI: Health Assessment Questionnaire-Disability Index.

Table 3. Summary of studies evaluating rituximab in systemic sclerosis.

Study Characteristics	Caliri ⁴²	McGonagle ⁴³	Daoussis ⁴⁶	Ketari ⁴⁴	Smith ⁴⁵	Lafyatis ⁴⁷
Study design	Observational	Observational	RCT	Observational	Observational	Observational
Sample size, n	1	1	14	1 [†]	8	15
Country	Italy	UK	Greece	Tunisia	Belgium	USA
Outcomes						
Skin score	Improved*	NR	No effect***	NR	Improved ^a	No effect ^c
HAQ-DI	NR	NR	NR	NR	No effect ^b	No effect ^d
Joint count	NR	Improved**	NR	NR	NR	NR
Adverse events (AE)	No serious AE	No serious AE	4 respiratory tract infections	No serious AE	No serious AE	46.7% infusion reactions

* Mean Rodnan skin score (MRSS) improved at 6 months. Baseline, 6-month followup Rodnan skin score, and p value not reported. ** Authors report arthropathy resolved completely. Joint count not reported. *** At 1 year, the rituximab group MRSS improved by 39.25% (mean 13.5 ± 6.84 at baseline vs 8.37 ± 6.45 at 1 yr). Control group improved by 20.80% (MRSS 11.50 ± 2.16 at baseline vs 9.66 ± 3.38 at 1 yr). Between-group differences were not statistically significant ($p = 0.06$). [†] Patient diagnosed with scleroderma-lupus overlap with catastrophic antiphospholipid antibody syndrome. ^a MRSS improved from mean 24.8 ± 3.4 at baseline to 14.3 ± 3.5 at 24 weeks ($p < 0.001$). ^b HAQ-DI mean 1.4 ± 0.6 at baseline to 1.3 ± 0.7 at 24 weeks ($p > 0.05$). ^c No significant change in MRSS between baseline and 6 months, mean change -0.37 , 95% CI $-3.8, 3.0$ ($p = 0.82$). ^d HAQ-DI did not change significantly between baseline mean 0.67 ± 0.32 and 6 months 0.64 ± 0.36 . HAQ-DI: Health Assessment Questionnaire-Disability Index; NR: not reported; RCC: randomized controlled trial.

and 17 patients randomized to the control groups. Mean followup was 7–17 months. Skin scores tended to improve in the treatment group ($p > 0.05$; Table 4).

Black, *et al*³⁷ report a randomized trial of IFN- α 13.5 x 10^6 units in divided doses in 35 patients with diffuse SSc. Because of a withdrawal of 13 patients, an interim analysis was performed. Three deaths and the lack of benefit in the treatment arm resulted in termination of further recruitment. A greater improvement in the placebo group skin score was observed (Table 4).

Seibold, *et al*³⁸ report a randomized trial of recombinant relaxin 25 $\mu\text{g}/\text{kg}/\text{day}$, 100 $\mu\text{g}/\text{kg}/\text{day}$, or placebo for 24 weeks by SC infusion in 72 patients with diffuse SSc.

Patients who received 25 $\mu\text{g}/\text{kg}$ of relaxin per day had lower skin scores than those who received placebo (mean change, -3.6 at 4 weeks, $p = 0.021$; -7.5 at 12 weeks, $p < 0.001$; and -8.7 at 24 weeks, $p = 0.040$). Joint score was defined as the sum of swollen or tender metacarpophalangeal joints (as a unit), wrists, and knees. No effect on joint score or HAQ-DI was observed (Table 4).

Genovese, *et al*³⁹ report a trial of PVAC in 18 patients with SSc randomized to 8 injections of 15 μg PVAC, 50 μg PVAC, or placebo at 3-week intervals. Patients who received PVAC 15 mg demonstrated improvement in MRSS [change score -4.3 (-20.6%)] at 24 weeks. Both the placebo and the PVAC 50 μg groups had worsening of skin disease,

Table 4. Summary of studies evaluating other biologic agents in systemic sclerosis.

Study Characteristics	Grassegger ³⁶	Black ³⁷	Seibold ³⁸	Genovese ³⁹	Denton ⁴⁰	Postlethwaite ⁴¹
Agent	IFN- γ	IFN- α	Relaxin	PVAC	Anti-TGF- β 1	Type I collagen
Study design	RCT	RCT	RCT	RCT	RCT	RCT
Sample size, n	44	35	68	18	45	168
Country	Austria	UK	USA	USA	UK	USA
Outcomes						
Skin score	Improved*	No effect**	Improved***	Improved 15 mg arm ^a	No effect ^b	No effect ^d
HAQ-DI	NR	NR	No effect [†]	No effect	No effect ^c	NR
Joint count	NR	NR	No effect ^{††}	NR	NR	NR
Adverse events (SE)	85% headache, 81% fever, 70% arthralgia/myalgia	n = 3 death, n = 5 flu-like symptoms	Menorrhagia 19–35%	Injection site reaction 67–100%	4 deaths related to disease	No serious AE

* Baseline mean 0.34 (SD 0.31) to 0.34 (SD 0.35) in control group; 0.67 (0.46) to 0.50 (0.30) in treatment group ($p > 0.05$). ** Mean change in IFN- α group was -4.7 (SEM 9.5) vs -7.5 (SEM 7.2) in placebo group ($p = 0.36$). *** Mean change from baseline in relaxin group was -3.6 at 4 wks ($p = 0.021$); -7.5 at 12 wks ($p < 0.001$); and -8.7 at 24 wks ($p = 0.040$). [†] Mean change from baseline in the relaxin 25 μ g/kg group was -0.125 ± 0.10 ($p = 0.154$); 100 μ g/kg group 0.036 ± 0.06 ($p > 0.2$). ^{††} Mean change from baseline in the relaxin 25 μ g/kg group was -0.9 ± 0.06 ($p = 0.133$); 100 μ g/kg group 0.2 ± 0.5 ($p > 0.2$). ^a Mean change from baseline in placebo group 3.5, PVAC 15 μ g group -4.3 , and PVAC 50 μ g group 3.5. ^b No difference in skin score across groups ($p = 0.49$). ^c No statistically significant differences in HAQ-DI scores across 4 treatment groups ranging from 0.80 ± 0.60 in 0.5 mg/kg group to 1.40 ± 0.90 in 5 mg/kg group. ^d No significant difference in mean change in MRSS at 12 months: -2.4 ± 7.3 versus -1.8 ± 7.4 ($p = 0.563$). IFN: interferon; RCT: randomized controlled trial; PVAC: delipidated deglycolipidated *M. vaccae*; TGF: tumor growth factor; HAQ-DI: Health Assessment Questionnaire-Disability Index; NR: not reported.

with MRSS increases of 3.5 (29.8%) and 3.5 (16.7%), respectively. There was no change in HAQ-DI (Table 4).

Denton, *et al*⁴⁰ report a trial of recombinant human anti-TGF- β 1 in 45 patients with diffuse SSc. Patients were randomized to placebo or 10 mg/kg, 5 mg/kg, or 0.5 mg/kg anti-TGF- β 1. There was no difference in the 6-month MRSS change score or HAQ-DI scores across the groups (Table 4).

Postlethwaite, *et al*⁴¹ report a study of 168 patients with SSc randomized to oral bovine type I collagen 500 μ g daily or placebo. There was no difference in the mean change in MRSS at 12 months (-2.4 ± 7.3 vs -1.8 ± 7.4 ; $p = 0.563$; Table 4).

DISCUSSION

There has been much recent interest in the use of biologic therapies in the treatment of SSc. In our systematic review, we have synthesized the 23 published studies that have evaluated the effect of these agents. Five studies have evaluated the use of anti-TNF- α therapies in SSc^{26,27,28,29,48}. The remaining 18 studies evaluate a variety of other biologic agents. Review of these studies gives us insight into the clinical utility of these agents, and potential future directions for research.

Anti-TNF therapy appears to confer benefits on inflammatory arthritis and disability. This finding has particular clinical relevance because inflammatory arthritis confers a significant burden of disease. Patients with SSc who have arthritis experience more severe pain than patients with RA, and more disability than patients with PsA⁴⁹. The first-line therapy for inflammatory arthritis is usually MTX. However, in the setting of pulmonary fibrosis commonly

seen in SSc, many clinicians may not feel comfortable initiating MTX. The use of TNF inhibitors may provide an alternative treatment option. The ability of anti-TNF agents to improve joint symptoms and disability is promising. However, the small sample sizes and observational study designs make these study findings subject to a number of biases. Further, these studies did not report joint counts, a commonly used outcome measure in RA studies. Therefore a comparison of the magnitude of the treatment effect in SSc cannot be compared to RA studies. It is worth noting that the utility of tender and swollen joint counts in SSc may be limited by overlying tight and thickened skin that can prohibit accurate assessment. Because of the potential difficulties associated with joint count assessment in SSc, investigators should consider additional measures of joint involvement (e.g., HAQ-DI, MRI). The studies in our review do indicate a potential beneficial treatment effect. The lack of statistical significance likely reflects the small sample sizes and insufficient power. These studies provide the necessary justification to proceed with larger, adequately powered studies to further evaluate this effect. In the future, the efficacy and safety of combination therapy (anti-TNF and MTX) in SSc-associated inflammatory arthritis should be evaluated because combination therapy has been found to be superior to TNF monotherapy in RA⁵⁰.

Some of the studies report improvement in skin scores with the use of anti-TNF agents. The generalizability of this finding is uncertain for a number of reasons. First, it is uncertain whether the observed skin softening reflected that which can occur as part of the natural history of the diseases, or reflects true treatment effect. Second, the more severe

skin scores in the treatment arm could reflect random variation. When the measurement is repeated, the skin score is lower as a result of “regression to the mean,” rather than true treatment effect. Third, the small observed treatment effects may be within the margin of measurement error. Clements, *et al* have reported an interrater variability of ± 4.6 units with the Rodnan skin score⁵¹. Fourth, the use of different measures of skin score across studies precludes comparisons. Although the majority of studies report the 17-site MRSS^{27,33,34,37,38,40,41,47}, others report the Rodnan total skin score^{29,32}, or the 15-site³⁶ and 18-site MRSS³⁰. Fifth, timing of inclusion into the study may affect the outcome. Patients who are recruited early in their disease or who have active skin involvement are more likely to derive benefit. Some studies explicitly state that patients were recruited early in their disease course or were mandated to have active disease as inclusion criteria, while other studies do not. Given these differences, we are unable to definitively conclude whether anti-TNF agents confer a beneficial effect on skin in SSc. The reported findings do provide a potential signal of benefit, and provide the necessary data required for justification of a larger, adequately powered study.

Anti-TNF agents were generally tolerated in patients with SSc. There was a high frequency of infusion reactions in patients with SSc treated with infliximab, which may be related to the development of anti-infliximab antibodies, and could be minimized with concurrent use of other immunosuppression. There was only 1 report of a lupus-like reaction, and 1 report of a decline in lung function in patients treated with etanercept²⁶. In this study, 2 patients were concomitantly treated with minocycline. Although unclear from the report, if the patient who developed the lupus-like reaction was also treated with minocycline, the lupus-like reaction could also have been attributable to the minocycline. All the published studies are limited in their sample size and duration of followup. Therefore we are unable to draw any conclusions about infrequent adverse effects or longterm safety. Large, longitudinal cohort studies (such as biologic registries) are needed to properly evaluate longterm safety.

Review of other non-TNF biologics in SSc was less promising. Patients treated with IFN- γ showed stabilization of skin scores while those on placebo had progression³⁶. In that study, baseline skin scores between treatment and control groups were not comparable, and IFN- γ treatment was poorly tolerated³⁶. The use of relaxin and PVAC showed a trend toward improved skin scores at low doses but worsening at high doses^{39,40}. The observed improvement in skin score may reflect that natural softening of skin observed in many patients with SSc over time. IFN- α , TGF- β 1, and oral type I collagen did not show promise for the treatment of SSc skin disease or disability. Initial small observational studies^{42,45} suggested improvements in skin score with rituximab, but this effect was not statistically greater than that

observed in the control arm of the randomized trial⁴⁶. Case reports of imatinib demonstrated evidence of skin improvement, and larger studies are currently evaluating this effect^{33,34,35}.

The heterogeneity across studies precluded our ability to aggregate or conduct a metaanalysis of the data. The studies differed in sample selection (inclusion and exclusion criteria), intervention, outcomes assessed (e.g., different skin scores), and study duration. Therefore it would be inappropriate to aggregate data from such variable sources. Future investigators should consider using recommended outcome measures that have demonstrable validity and reliability^{52,53}. This would facilitate quantifiable comparisons across studies.

Despite the limitations of this systematic review, qualitative synthesis of the existing literature and presentation of the individual study findings is useful, especially when considering one of these agents for an individual patient. The results of this systematic review suggest that etanercept and infliximab may be safe (in the short term) and effective for the management of inflammatory arthritis. The utility of anti-TNF agents for skin involvement is less certain. ATG and imatinib may be effective for skin involvement but the longterm safety is unknown. None of the other biologic agent studies were able to demonstrate a significant change in inflammatory arthritis, disability, or skin score. Well designed, adequately powered clinical trials are needed to further evaluate the efficacy, and large, longitudinal studies are needed to evaluate the longterm safety of these agents in SSc.

ACKNOWLEDGMENT

The authors thank Amy Faulkner, Information Specialist at the University Health Network Library Services, for her assistance with this study.

REFERENCES

1. Misra R, Darton K, Jewkes RF, Black CM, Maini RN. Arthritis in scleroderma. *Br J Rheumatol* 1995;34:831-7.
2. La Montagna G, Baruffo A, Tirri R, Buono G, Valentini G. Foot involvement in systemic sclerosis: a longitudinal study of 100 patients. *Semin Arthritis Rheum* 2002;31:248-55.
3. Avouac J, Walker U, Tyndall A, Kahan A, Matucci-Cerinic M, Allanore Y. Characteristics of joint involvement and relationships with systemic inflammation in systemic sclerosis: Results from the EULAR Scleroderma Trial and Research Group (EUSTAR) Database. *J Rheumatol* 2010;37:1488-501.
4. Schumacher HR Jr. Joint involvement in progressive systemic sclerosis (scleroderma): a light and electron microscopic study of synovial membrane and fluid. *Am J Clin Pathol* 1973;60:593-600.
5. Low AH, Lax M, Johnson SR, Lee P. Magnetic resonance imaging of the hand in systemic sclerosis. *J Rheumatol* 2009;36:961-4.
6. Morita Y, Muro Y, Sugiura K, Tomita Y. Anti-cyclic citrullinated peptide antibody in systemic sclerosis. *Clin Exp Rheumatol* 2008;26:542-7.
7. Marrone M, Chiala A, Tampona M, Iannone F, Raho L, Covelli M, et al. [Prevalence of anti-CCP antibodies in systemic sclerosis]. *Reumatismo* 2007;59:20-4.
8. Distler JH, Schett G, Gay S, Distler O. The controversial role of

- tumor necrosis factor alpha in fibrotic diseases. *Arthritis Rheum* 2008;58:2228-35.
9. Piguet PF, Collart MA, Grau GE, Sappino AP, Vassalli P. Requirement of tumour necrosis factor for development of silica-induced pulmonary fibrosis. *Nature* 1990;344:245-7.
 10. Piguet PF, Vesin C. Treatment by human recombinant soluble TNF receptor of pulmonary fibrosis induced by bleomycin or silica in mice. *Eur Respir J* 1994;7:515-8.
 11. Liu JY, Brass DM, Hoyle GW, Brody AR. TNF-alpha receptor knockout mice are protected from the fibroproliferative effects of inhaled asbestos fibers. *Am J Pathol* 1998;153:1839-47.
 12. Ortiz LA, Lasky J, Hamilton RF Jr, Holian A, Hoyle GW, Banks W, et al. Expression of TNF and the necessity of TNF receptors in bleomycin-induced lung injury in mice. *Exp Lung Res* 1998;24:721-43.
 13. Ortiz LA, Lasky J, Lungarella G, Cavarra E, Martorana P, Banks WA, et al. Upregulation of the p75 but not the p55 TNF-alpha receptor mRNA after silica and bleomycin exposure and protection from lung injury in double receptor knockout mice. *Am J Respir Cell Mol Biol* 1999;20:825-33.
 14. Mauviel A, Daireaux M, Redini F, Galera P, Loyau G, Pujol JP. Tumor necrosis factor inhibits collagen and fibronectin synthesis in human dermal fibroblasts. *FEBS Lett* 1988;236:47-52.
 15. Mauviel A, Heino J, Kahari VM, Hartmann DJ, Loyau G, Pujol JP, et al. Comparative effects of interleukin-1 and tumor necrosis factor-alpha on collagen production and corresponding procollagen mRNA levels in human dermal fibroblasts. *J Invest Dermatol* 1991;96:243-9.
 16. Ito A, Sato T, Iga T, Mori Y. Tumor necrosis factor bifunctionally regulates matrix metalloproteinases and tissue inhibitor of metalloproteinases (TIMP) production by human fibroblasts. *FEBS Lett* 1990;269:93-5.
 17. Dayer JM, Beutler B, Cerami A. Cachectin/tumor necrosis factor stimulates collagenase and prostaglandin E2 production by human synovial cells and dermal fibroblasts. *J Exp Med* 1985;162:2163-8.
 18. Meikle MC, Atkinson SJ, Ward RV, Murphy G, Reynolds JJ. Gingival fibroblasts degrade type I collagen films when stimulated with tumor necrosis factor and interleukin 1: evidence that breakdown is mediated by metalloproteinases. *J Periodontol* 1989;24:207-13.
 19. Furst DE, Breedveld FC, Kalden JR, Smolen JS, Burmester GR, Bijlsma JW, et al. Updated consensus statement on biological agents, specifically tumour necrosis factor alpha (TNF-alpha) blocking agents and interleukin-1 receptor antagonist (IL-1ra), for the treatment of rheumatic diseases, 2004. *Ann Rheum Dis* 2004;63 Suppl 2:ii2-12.
 20. Keyser FD, Mielants H, Veys EM. Current use of biologicals for the treatment of spondyloarthropathies. *Exp Opin Pharmacother* 2001;2:85-93.
 21. Olsen NJ, Stein CM. New drugs for rheumatoid arthritis. *N Engl J Med* 2004;350:2167-79.
 22. Allanore Y, Devos-Francois G, Caramella C, Boumier P, Jounieaux V, Kahan A. Fatal exacerbation of fibrosing alveolitis associated with systemic sclerosis in a patient treated with adalimumab. *Ann Rheum Dis* 2006;65:834-5.
 23. Ostor AJ, Crisp AJ, Somerville MF, Scott DG. Fatal exacerbation of rheumatoid arthritis associated fibrosing alveolitis in patients given infliximab. *BMJ* 2004;329:1266.
 24. Shakoor N, Michalska M, Harris CA, Block JA. Drug-induced systemic lupus erythematosus associated with etanercept therapy. *Lancet* 2002;359:579-80.
 25. Gartlehner G, Hansen RA, Jonas BL, Thieda P, Lohr KN. The comparative efficacy and safety of biologics for the treatment of rheumatoid arthritis: a systematic review and metaanalysis. *J Rheumatol* 2006;33:2398-408.
 26. Lam GK, Hummers LK, Woods A, Wigley FM. Efficacy and safety of etanercept in the treatment of scleroderma-associated joint disease. *J Rheumatol* 2007;34:1636-7.
 27. Bosello S, De Santis M, Tulusso B, Zoli A, Ferraccioli G. Tumor necrosis factor-alpha inhibitor therapy in erosive polyarthritis secondary to systemic sclerosis. *Ann Intern Med* 2005;143:918-20.
 28. Denton CP, Engelhart M, Tvede N, Wilson H, Khan K, Shiwen X, et al. An open-label pilot study of infliximab therapy in diffuse cutaneous systemic sclerosis. *Ann Rheum Dis* 2009;68:1433-9.
 29. Ellman MH, MacDonald PA, Hayes FA. Etanercept as treatment for diffuse scleroderma: A pilot study [abstract]. *Arthritis Rheum* 2000;43 Suppl:S392.
 30. Matteson EL, Shbeeb MI, McCarthy TG, Calamia KT, Mertz LE, Goronzy JJ. Pilot study of antithymocyte globulin in systemic sclerosis. *Arthritis Rheum* 1996;39:1132-7.
 31. Stratton RJ, Wilson H, Black CM. Pilot study of anti-thymocyte globulin plus mycophenolate mofetil in recent-onset diffuse scleroderma. *Rheumatology* 2001;40:84-8.
 32. Balaban EP, Zashin SJ, Geppert TD, Lipsky PE, Condie RM. Treatment of systemic sclerosis with antithymocyte globulin. *Arthritis Rheum* 1991;34:244-5.
 33. van Daele PL, Dik WA, Thio HB, van Hal PT, van Laar JA, Hooijkaas H, et al. Is imatinib mesylate a promising drug in systemic sclerosis? *Arthritis Rheum* 2008;58:2549-52.
 34. Sfrikakis PP, Gorgoulis VG, Katsiari CG, Evangelou K, Kostopoulos C, Black CM. Imatinib for the treatment of refractory, diffuse systemic sclerosis. *Rheumatology* 2008;47:735-7.
 35. Chung L, Fiorentino DF, Benbarak MJ, Adler AS, Mariano MM, Paniagua RT, et al. Molecular framework for response to imatinib mesylate in systemic sclerosis. *Arthritis Rheum* 2009;60:584-91.
 36. Grassegger A, Schuler G, Hessenberger G, Walder-Hantich B, Jabkowski J, MacHeiner W, et al. Interferon-gamma in the treatment of systemic sclerosis: a randomized controlled multicentre trial. *Br J Dermatol* 1998;139:639-48.
 37. Black CM, Silman AJ, Herrick AI, Denton CP, Wilson H, Newman J, et al. Interferon-alpha does not improve outcome at one year in patients with diffuse cutaneous scleroderma: results of a randomized, double-blind, placebo-controlled trial. *Arthritis Rheum* 1999;42:299-305.
 38. Seibold JR, Korn JH, Simms R, Clements PJ, Moreland LW, Mayes MD, et al. Recombinant human relaxin in the treatment of scleroderma. A randomized, double-blind, placebo-controlled trial. *Ann Intern Med* 2000;132:871-9.
 39. Genovese MC, Chakravarty EF, Boyle DL, Tutuncu Z, Thorburn CM, Halilhodzic M, et al. A randomized, blinded, parallel group, placebo controlled pilot study evaluating the effect of PVAC treatment in patients with diffuse systemic sclerosis. *J Rheumatol* 2005;32:2345-50.
 40. Denton CP, Merkel PA, Furst DE, Khanna D, Emery P, Hsu VM, et al. Recombinant human anti-transforming growth factor beta-1 antibody therapy in systemic sclerosis: a multicenter, randomized, placebo-controlled phase I/II trial of CAT-192. *Arthritis Rheum* 2007;56:323-33.
 41. Postlethwaite AE, Wong WK, Clements PJ, Chatterjee S, Fessler BJ, Kang AH, et al. A multicenter, randomized, double-blind, placebo-controlled trial of oral type I collagen treatment in patients with diffuse cutaneous systemic sclerosis: I. Oral type I collagen does not improve skin in all patients, but may improve skin in late-phase disease. *Arthritis Rheum* 2008;58:1810-22.
 42. Caliri A, Sangari D, Sferrazza P, Bagnato G, Bagnato G. Effectiveness of rituximab in the treatment of a rare overlap syndrome — rheumatoid arthritis/s. Sjogren/systemic sclerosis — complicated by monoclonal gammopathy [Italian]. *Trends Med* 2009;9:161-2.
 43. McGonagle D, Tan AL, Madden J, Rawstron AC, Rehman A,

- Emery P, et al. Successful treatment of resistant scleroderma-associated interstitial lung disease with rituximab. *Rheumatology* 2008;47:552-3.
44. Ketari JS, Zaghoudi I, Ben Dhaou B, Kochbati S, Mir K, Ben Ali Z, et al. [Catastrophic antiphospholipid syndrome and rituximab: a new report]. *Tunis Med* 2009;87:699-702.
45. Smith V, Van Praet JT, Vandooren B, Van der Cruyssen B, Naeyaert JM, Decuman S, et al. Rituximab in diffuse cutaneous systemic sclerosis: an open-label clinical and histopathological study. *Ann Rheum Dis* 2010;69:193-7.
46. Daoussis D, Liossis SN, Tsamandas AC, Kalogeropoulou C, Kazantzi A, Sirinian C, et al. Experience with rituximab in scleroderma: results from a 1-year, proof-of-principle study. *Rheumatology* 2010;49:271-80.
47. Lafyatis R, Kissin E, York M, Farina G, Viger K, Fritzler MJ, et al. B cell depletion with rituximab in patients with diffuse cutaneous systemic sclerosis. *Arthritis Rheum* 2009;60:578-83.
48. Marie I, Lahaxe L, Levesque H, Heliot P. Pulmonary actinomycosis in a patient with diffuse systemic sclerosis treated with infliximab. *QJM* 2008;101:419-21.
49. Johnson SR, Glaman DD, Schentag CT, Lee P. Quality of life and functional status in systemic sclerosis compared to other rheumatic diseases. *J Rheumatol* 2006;33:1117-22.
50. Gartlehner G, Hansen RA, Jonas BL, Thieda P, Lohr KN. The comparative efficacy and safety of biologics for the treatment of rheumatoid arthritis: a systematic review and metaanalysis. *J Rheumatol* 2006;33:2398-408.
51. Clements PJ, Lachenbruch PA, Seibold JR, Zee B, Steen VD, Brennan P, et al. Skin thickness score in systemic sclerosis: an assessment of interobserver variability in 3 independent studies. *J Rheumatol* 1993;20:1892-6.
52. Johnson SR, Hawker GA, Davis AM. The Health Assessment Questionnaire Disability Index and Scleroderma Health Assessment Questionnaire in scleroderma trials: an evaluation of their measurement properties. *Arthritis Rheum* 2005;53:256-62.
53. Khanna D, Distler O, Avouac J, Behrens F, Clements PJ, Denton C, et al. Measures of response in clinical trials of systemic sclerosis: the Combined Response Index for Systemic Sclerosis (CRISS) and Outcome Measures in Pulmonary Arterial Hypertension related to Systemic Sclerosis (EPOSS). *J Rheumatol* 2009;36:2356-61.