Drs. Nakabo and Ohmura reply

To the Editor:

We appreciate the comments of Drs. Mahler and Fritzler¹ in the letter to the editor regarding our article². We will sincerely reply to the concerns raised by the authors.

We acknowledge that interlaboratory differences in anticarbamylated protein (anti-CarP) ELISA are an important issue. Although we slightly modified the original method used by Shi, *et al*³, the prevalence of anti-CarP antibodies in rheumatoid arthritis (RA) using our method was similar to that reported previously⁴. Therefore, we consider our in-house ELISA to be equivalent to those used in other laboratories. Our recent study gives a detailed protocol of our system⁵.

We agree with the comment regarding the relationship between pre-test probability and sensitivity/specificity. We would like to correct the sentence “although the pre-test probability deeply affects the sensitivity and specificity of anti-CarP antibodies in daily clinical practice, our data suggest that anti-CarP antibodies cannot be used for differentiating [anticitrullinated protein antibodies] ACPA-negative RA from non-RA [connective tissue disease] CTD” in the Discussion section to “although sensitivity and specificity differ depending on comparators, our results suggest that anti-CarP antibodies cannot be used to differentiate ACPA-negative RA from non-RA CTD.”

According to the suggestions provided, we prepared Venn diagrams showing the number of patients testing positive for each antibody (Figure 1). Triple-positive patients were mainly observed in the RA group. However, the OR of ACPA, rheumatoid factor (RF), and anti-CarP antibodies for the diagnosis of RA in our cohort were 57.3 (95% CI 35.4–92.9), 14.3 (9.4–21.9), and 2.8 (2.1–3.9), respectively. In the group with ACPA and RF, OR was elevated to 84.3 (48.9–145.4), whereas it was as low as 55.1 (23.8–128.0) in the triple-positive group. The addition of the anti-CarP test to ACPA and RF did not increase OR, at least not in our cohort.

We agree with Drs. Mahler and Fritzler that our cohort does not reflect daily clinical settings, and the combined test of ACPA, RF, and anti-CarP antibodies may contribute to a preclinical diagnosis and very early intervention; however, when we want to exclude the possibility of CTD, our cohort is appropriate for testing the utility of the anti-CarP antibody. The careful exclusion of non-RA CTD is required in any cohort.

Regarding ELISA, using 1 specific protein or peptide as an antigen, we previously reported that albumin is one of the target antigens of anti-CarP antibodies⁵, and the prevalence of anti-CarP albumin antibodies in each CTD was similar to that of anti-CarP antibodies (Figure 2). Previous studies reported that antibodies against carbamylated fibrinogen⁶ and vimen-

tin-derived peptide⁷ coexisted with ACPA. Although we agree that the combination of these specific ELISA may contribute to the diagnosis of RA, the clinical efficacy of these antibodies in seronegative RA may be limited.

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J Rheumatol 2018;45:3; doi:10.3899/jrheum.171415

Figure 1. Venn diagrams of ACPA, anti-CarP antibodies, and RF in RA (A) and non-RA CTD (B). ACPA: anticitrullinated peptide antibodies; anti-CarP: anticarbamylated protein antibodies; RF: rheumatoid factor; RA: rheumatoid arthritis; CTD: connective tissue diseases.

Figure 2. Anti-CarALB antibody prevalence and levels in RA and other CTD. The horizontal line represents the cutoff value. The anti-CarALB level was measured and the cutoff value was selected as described in our previous study⁵. Anti-CarALB: anticarbamylated albumin; RA: rheumatoid arthritis; CTD: connective tissue diseases; ACPA: anticitrullinated peptide antibodies; SLE: systemic lupus erythematosus; PM/DM: polymyositis/dermatomyositis; SSc: systemic sclerosis; MCTD: mixed connective tissue diseases; pSS: primary Sjögren syndrome; BD: Behçet disease; AOSD: adult-onset Still disease; PMR: polymyalgia rheumatica; SpA: spondyloarthritis; HC: healthy controls.