Progressive Multifocal Leukoencephalopathy in a Patient with Polymyositis: Case Report and Literature Review

To the Editor:

We describe a fatal case of progressive multifocal leukoencephalopathy (PML) in a patient with polymyositis (PM) and antisynthetase syndrome treated with conventional immunosuppressive drugs.

PML is a rare and fatal demyelinating disease caused by the ubiquitous John Cunningham virus (JCV)^{1,2}. JCV infection is usually an asymptomatic event in childhood and about 60%-80% of the normal adult population are seropositive for antibodies against JCV^{3,4}. After primary infection, the virus remains in a latent form in the bone marrow, lymphoid organs, and kidney and some studies have demonstrated that it also can establish latency in healthy brain tissue although this is a matter of debate^{5,6}. The virus may later reactivate, undergoing genomic rearrangement in the noncoding regulatory region, and migrate to the brain, where it infects glial cells and causes a lytic infection known as PML^{7,8}. Reactivation usually occurs during conditions of immunosuppression but there are also reports of PML development in immunocompetent individuals⁹. The mechanisms behind the pathway leading to viral reactivation remain insufficiently defined. The risk of PML in autoimmune diseases has recently been highlighted because of an increasing number of PML cases occurring during treatment with new biological immunosuppressive agents including natalizumab (for multiple sclerosis, MS), rituximab (RTX; for rheumatic diseases), and efalizumab (for psoriasis) and also with the newer immunosuppressant agent mycophenolate mofetil (MMF)¹⁰. In the more rare autoimmune diseases polymyositis/dermatomyositis (PM/DM) there are a few reports of PML developing in patients treated with conventional disease-modifying antirheumatic drugs^{9,11,12,13,14,15,16}, but also in patients during off-label treatment with RTX¹⁷. The increased risk for PML associated with the new biologics has raised the question of whether these treatments involve a biological explanation for the increased risk. The background risk for PML in autoimmune diseases is, however, not known, since epidemiological studies are lacking on the incidence of PML in autoimmune diseases and data to date are dependent on case reports or case series.

A 65-year-old woman negative for human immunodeficiency virus (HIV) and having bronchial asthma, type 2 diabetes mellitus, and substituted hypothyroidism presented in our clinic because of fever, fatigue, symmetrical polyarthritis, myalgia in her proximal muscles, paresthesia, Raynaud's phenomenon, and dyspnea. Her initial laboratory tests revealed slightly elevated erythrocyte sedimentation rate (ESR) 21 mm/h (normal < 20 mm/h), C-reactive protein (CRP) 38 mg/l (normal < 7 mg/l), and markedly elevated levels of alanine aminotransferase, 108 U/I (normal < 45.6 U/l), aspartate aminotransferase, 198 U/l (normal < 36.6 U/l), creatine kinase, 3168 U/I (normal 36-210 U/I), and lactate dehydrogenase, 780 U/I (normal < 210 U/l). White blood cell (WBC) count was within normal limits $(8.7 \times 10^9 \text{/l})$. Immunological tests including rheumatoid factor, antinuclear antibodies, and c-antineutrophil cytoplasmic antibodies were negative, but she was positive for anti-Jo-1 antibodies. A muscle biopsy (from the quadriceps muscle) revealed fibers with signs of regeneration and degeneration and variations in muscle fiber size, but there were no inflammatory infiltrates. Electroneurography demonstrated carpal tunnel syndrome and electromyography confirmed alterations typical for myositis, including spontaneous fibrillations and positive sharp waves in the musculus erector spinae. High-resolution computerized tomography of the lungs confirmed bilateral alveolitis. Lung function tests showed decreased vital capacity (71%) and total lung capacity (66%), indicating restrictive lung disease. She was diagnosed with PM and antisynthetase syndrome with interstitial lung disease and was given prednisone 40 mg/day and oral cyclophosphamide 100 mg/day. In addition she received intraarticular glucocorticoid injections of small joints of the hands. She improved clinically after 12 months, then the cyclophosphamide was replaced by methotrexate (MTX) 20 mg/week and the daily prednisone dose was reduced to 10 mg. Eighteen months after diagnosis of PM and 6 months' treatment with MTX, she developed a flare of myositis and the prednisone dose was increased from 2.5 to 40 mg/day, whereby her symptoms improved. Despite this, she subsequently experienced a flare of arthritis and MTX was switched to oral azathioprine 150 mg/day; however, this was stopped after 6 weeks due to nausea and increased liver enzymes. Oral prednisone 17.5 mg/day was continued.

Four months later, roughly 2 years after she was diagnosed with PM and antisynthetase syndrome, she developed fatigue, vertigo, and balance problems. Neurological examination demonstrated horizontal nystagmus, dysarthria, ataxia of the left extremities, and a mild left-side hemiparesis. Laboratory tests showed CRP and ESR within normal ranges and slightly elevated WBC count (11.8 \times 10⁹/l, with normal lymphocyte count 1.3 \times 10⁹/l). Brain magnetic resonance imaging (MRI) revealed 2 hyperintense white-matter lesions in the left cerebellar hemisphere and 1 hyperintense white-matter lesion in the brain stem, and supratentorial diffuse hyperintense lesions of the white matter. The lesions had no expansive effect or contrast enhancement (Figure 1). The patient was incorrectly suspected to have central nervous system vasculitis. Intravenous (IV) pulse methylprednisone 1000 mg daily on 2 consecutive days was started, but vertigo and balance disturbances worsened. A new MRI scan of the brain showed progression of the cerebellar lesions and appearance of a new pontine lesion without mass effect or contrast enhancement. Despite treatment with IV prednisone 60 mg/day and subcutaneous low molecular weight heparin, the patient's condition worsened and developed progressive tetraparesis and dysphagia. A lumbar puncture showed a normal cell count, slightly elevated albumin, and a positive JCV polymerase chain reaction (PCR) analysis with detection of > 200 copies/ml (SMI, Stockholm, Sweden; exact number of JCV copies not analyzed), which confirmed the diagnosis of PML. Prednisone was tapered from 40 to 12.5 mg/day. Antiviral therapy was discussed but the patient declined any further medication. Her condition deteriorated and she died 4.5 months after the first neurological symptoms had presented. Brain autopsy revealed PML lesions in the brain stem and cerebellum and positive in situ staining for JCV DNA. No supratentorial PML lesions were observed; the diffuse lesions seen on MRI were of degenerative origin. A general autopsy revealed only mild atherosclerosis of the coronary arteries and moderate aortic atherosclerosis.

We performed an updated literature search of PML cases in PM/DM patients with no other autoimmune disease. We identified 8 cases that were treatment-naive for the newer biologics, summarized in Table 1. All patients were HIV-negative and 8 had no other underlying disease; 1 patient also had chronic renal insufficiency. The patients developed PML after a range of 7 months to 5 years of disease duration of PM/DM. Seven patients were diagnosed with DM and 2 with PM. Six were women and the age span at disease onset was 29-72 years. One patient, with a favorable outcome, had no treatment when presenting with symptoms suggestive of PML, and had earlier been treated only with hydroxychloroquine. Three patients were treated only with oral prednisone at time of PML diagnosis (including the present case). One patient had ongoing treatment with MMF at the time of development of PML, 2 patients had ongoing treatment with azathioprine, another 2 had ongoing treatment with cyclosporine, and 1 patient was treated with cyclophosphamide and IV immunoglobulins upon developing PML. Five patients had supratentorial white-matter lesions on MRI, which is the most typical localization of PML; 3 patients had only infratentorial white-matter lesions, including our case, and gadolinium enhancement was seen in 2 patients. One female patient had a negative PCR analysis for JCV DNA in 2 repeated cerebrospinal fluid (CSF) analyses despite multiple supratentorial white-matter PML lesions, and the PML diagnosis was instead confirmed by brain biopsy 13 . That case demonstrates the importance of repeating the PCR analysis for JCV in CSF with ultrasensitive techniques or proceeding to a brain biopsy if suspicion of PML

Six patients died and 3 survived and improved clinically; 2 of the survivors had been treated only with mild immunosuppressants (prednisone, hydroxychloroquine), but the third had been treated with MMF and cyclosporine in addition to prednisone. After PML diagnosis, 4 patients received treatment with cytosine-arabinoside, which might have been suc-

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2012. All rights reserved.

Letter 1299

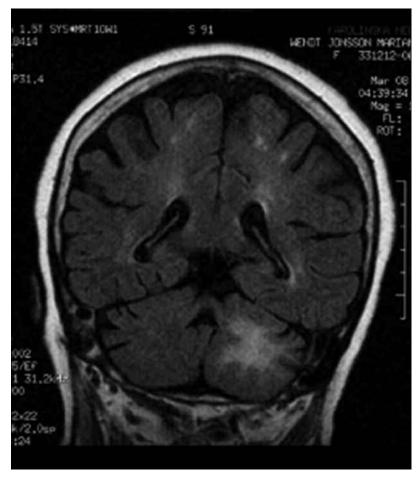


Figure 1. Coronary T2 weighted image brain magnetic resonance imaging (MRI) shows a single hyperintense lesion of the left cerebellar lobe representing progressive multifocal leukoencephalopathy (PML) and supratentorial diffuse hyperintense white-matter lesions. MRI was performed 3 weeks after onset of PML symptoms.

cessful in the 2 that survived. The time to death after onset of PML symptoms ranged between 1 and 5 months.

We also found 1 recent report on a patient with PM who experienced fatal PML during treatment with the biological agent RTX (Table 1, Patient 10)¹⁷. This patient developed clinical signs of PML after 5 infusions of RTX and 6.5 years' duration of PM. The patient was treated with RTX because her treatment had been refractory to conventional immunosuppressive therapy.

We describe a patient with PM and antisynthetase syndrome who developed clinical symptoms of PML after 22 months of conventional immunosuppressive treatment including prednisone, oral cyclophosphamide for 13 months, MTX for 6 months, and azathioprine for 6 weeks. The diagnosis of PML was based on detection of JCV DNA in her CSF. The current diagnostic test for PML is detection of JCV DNA in CSF, using PCR methods, or in brain tissue obtained by biopsy or autopsy. There are, however, reports of negative JCV PCR tests in CSF collected early in the course of PML disease ¹⁸, as well as reports of false-negative results for JCV in CSF despite high levels of virus load, due to genomic variability and use of incorrect sets of primers and probes ¹⁹. Interestingly, the majority of PML cases occurring in patients with MS treated with the new biological agent natalizumab have had much lower JCV load in their CSF (≤ 500 copies/ml) ²⁰ at time of diagnosis, compared to reports of PML not associated with new biological treatments ^{21,22}.

Through the literature search we found another 8 cases of PML after

conventional immunosuppressive treatment. The previously reported cases with PML in patients with PM and DM were similar to our case in their PML features, but 3 of the previously reported 8 cases survived, whereas our patient did not. The surviving patients seemed to have received fewer immunosuppressive agents before developing PML, compared to our case, and 2 of the survivors received antiviral treatment, which may have affected the outcome.

PML has been recognized as a potential risk in patients treated with biological agents. No biological agent has to date been approved for treatment of PM or DM. One phase II randomized double-blind placebo-controlled study using RTX for DM and PM was recently performed, with no reports of cases of PML²³. However, 1 patient with PM developing PML was reported during off-label use of RTX treatment 17. We have also received information on 3 other reported cases of PML in patients with DM or PM treated with RTX (Roche Pharmaceuticals, personal communication). These data may indicate a high number of PML cases in a rare inflammatory disease where the number of patients treated with RTX to date is limited. The prevalence of inflammatory myopathies is about 1/15-50 compared to systemic lupus erythematosus and 1/800 compared to rheumatoid arthritis²⁴. Therefore the number of reported cases with PML in patients with PM/DM might be higher compared to other autoimmune diseases, but to clarify this issue, the background risk of PML in PM and DM must be known.

There is at present no treatment for PML. The most important action is

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2012. All rights reserved.

Table 1. Data on patients with polymyositis/dermatomyositis developing progressive multifocal leukoencephalopathy (PML).

Case	Sex, Age, yrs	Underlying Disease and Duration	IST Before PML	IST at PML	PML Symptoms	MRI Findings	CSF JCV DNA PCR	PML Diagnosis	PML- specific Treatment	PML Outcome
113	F 52	DM, 5 yrs	Prednisone	Prednisone	↓ Psychomotor performance, memory loss, aphasia, N VII palsy	Supratentorial and infratentorial T2 WM lesions, Gd+ cranial nerve roots VII and VIII	Negative × 2	Brain biopsy: pos JCV DNA PCR	CYT-ARA	Improved
213	M 44	DM, 2.5 yrs	Prednisone MMF, IVIG, CYC	Prednisone MMF, IVIG, CYC	↓ Visual acuity hemianopsia	Supratentorial T2 WM lesions	Positive 10 ⁴ –10 ⁵ JCV copies/ml	Pos CSF JCV DNA PCR	CYT-ARA, mirtazapine	Improved
314	F 72	DM, unknown	AZA	AZA	Cognitive deficits, personality changes	Supratentorial Gd+ T2 WM lesions	?	Brain biopsy and autopsy: pos JCV DNA ISH	CYT-ARA	Death in 1 mo
4 ¹¹	F 45	DM, 7 mo, chronic renal insufficiency	Prednisone, IVIG, CYC	Prednisone, IVIG, CYC	Altered consciousness, progressive tetraparesis	Supratentorial T2 WM lesions	?	Autopsy: intranuclear viral bodies and inflammation	None	Death in 1 mo
512	F 43	DM, 5 yrs	Prednisone, AZA	Prednisone	Gait ataxia, dysarthria, corticospinal tract involvement	T2 WM lesions cerebellum and brainstem	Negative	Brain biopsy: pos JCV ab	Cidofovir, cytarabine	Death in 3 mo
69	F 29	DM, unknown	HCQ	None	Left hemiparesis, diplopia, nystagmus, N III, N VI palsy	Supratentorial WM lesions, 1 thalamic lesion	?	"Pos brain biopsy"	IV steroids, mirtazapine	Improved
7 ¹⁶	M 54	PM, unknown	Prednisone, chlorambucil	?	Altered behavior, impaired memory, ataxia	?	Not done?	Brain biopsy: pos JCV ab	None	Death in 5 mo
815	M 67	DM, 4 yrs	Dexamethasone, CPP, IVIG	sirolismus, IVIG		Supratentorial T2 WM lesions	Positive	Not performed	Cidofovir	Death within weeks
9	F 67, present case	PM, 2 yrs	Prednisone, CPP, IVIG, MTX, AZA	Prednisone	Limb ataxia, nystagmus, L hemiparesis, dysarthria	T2 WM lesions cerebellum and brainstem	Positive	Pos CSF JCV DNA PCR; brain biopsy: pos JCV DNA ISH	None	Death in 4.5 mo
10 ¹⁷	F 43	PM 6.5 yrs	Prednisone, MTX, AZA, MMF, CYC, IVIG, rituximab	Prednisone, rituximab	Bilat hemianopsia, ↓ visual acuity	Supratentorial subcortical and cortical T2 and FLAIR WM lesions	Positive	Pos CSF JCV DNA PCR	None	Death in 8 mo

IST: immunosuppressive treatment; DM: dermatomyositis; PM: polymyositis; MMF: mycophenolate mofetil; AZA: azathioprine; CPP: cyclophosphamide; CYC: cyclosporine; HCQ: hydroxychloroquine; MTX: methotrexate; IVIG: intravenous immunoglobulins; Gd+: gadolinium enhancement; CYT-ARA: cytosine-arabinoside; ISH: *in situ* hybridization; ab: antibody; WM: white matter; CSF: cerebrospinal fluid; JCV: John Cunningham virus; MRI: magnetic resonance imaging; PCR: polymerase chain reaction.

withdrawal of immunosuppressive drugs and, if possible, rapid elimination of the specific biological agent.

Administration of the serotonin receptor antagonist mirtazapine has been tried with varying results (serotonin receptors are used by JCV for entry into the cell), but because clinical studies are lacking, this procedure is discouraged. The antimalarial drug mefloquine was shown to inhibit JCV replications *in vitro*²⁵, but a subsequent clinical study was discontinued due to lack of significant efficacy (www.clinicaltrials.gov). There are case reports and series that have reported favorable outcomes of PML after treatment with cytosine-arabinoside, including a few patients with rheumatic diseases^{13,14}; however, a large placebo-controlled trial on the effect of cytosine-arabinoside for HIV-associated PML failed to demonstrate any effect of the drug²⁶. A recent study on the first 35 postmarketing cases of PML in patients with MS treated with natalizumab identified several fac-

tors associated with improved survival, including shorter time from symptom onset to diagnosis of PML, younger age at diagnosis, less disability before PML, and more localized disease on MRI²⁷.

One suggested strategy to assess risk for PML is to screen for seropositivity of JCV antibodies to determine whether the patient has been infected with JCV. This is recommended as a potential tool for stratifying PML risk in persons with MS, prior to initiation of natalizumab treatment²⁸. Patients who are found to be JCV-seronegative should be followed for seroconversion repeatedly during immunosuppressive treatment. By contrast, several studies have demonstrated that screening for JCV DNA in blood, CSF, or urine does not enhance the ability to predict development of PML^{29,30}.

The mechanisms behind JCV reactivation are not understood; immunosuppression is a major causative factor, but host and viral factors are cer-

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2012. All rights reserved.

Letter 1301

tainly important as well. As the use of biological agents in autoimmune disease increases, it is important to improve our knowledge about the mechanisms behind JCV reactivation leading to PML, regardless of treatment, to develop effective strategies to reduce risk of PML. While in theory any immunosuppressive regimen can increase the risk of PML, it had not been seen with MS, Crohn's disease, or psoriasis until the introduction of natalizumab and efalizumab, despite the relatively aggressive immunosuppressive regimens used in these conditions. This may suggest that there is some contribution to the development of PML from the underlying condition being treated.

Patients with autoimmune diseases also carry a risk for reactivation of JCV and PML with milder immunosuppressive treatment, and more rarely, without such treatment. The frequency of PML in autoimmune diseases remains unknown because confirming epidemiological data are sparse. In this context the background prevalence of PML in patients with PM or DM will be important and there is great value in reporting cases such as ours.

MARYAM DASTMALCHI, MD, PhD, Rheumatology Unit, Department of Medicine, Karolinska Institute, Karolinska University Hospital; JUDIT LAKI, MD, PhD, Rheumatology Unit, Department of Medicine, Karolinska Institute, Karolinska University Hospital; Department of Rheumatology, National Health Centre, Budapest, Hungary; INGRID E. LUNDBERG, MD, PhD, Rheumatology Unit, Department of Medicine, Karolinska Institute, Karolinska University Hospital; ELLEN IACOBAEUS, MD, PhD, Neuroimmunology Unit, Department of Clinical Neuroscience, Karolinska Institute, Karolinska University Hospital, Solna 17176, Stockholm, Sweden. Address correspondence to Dr. E. Iacobaeus; E-mail: ellen.iacobaeus@karolinska.se

Supported by the Regional Agreement on Medical Training and Clinical Research (ALF) between Stockholm County Council and Karolinska Institutet.

REFERENCES

- Astrom KE, Mancall EL, Richardson EP Jr. Progressive multifocal leuko-encephalopathy; a hitherto unrecognized complication of chronic lymphatic leukaemia and Hodgkin's disease. Brain 1958;81:93-111.
- Padgett BL, Walker DL, ZuRhein GM, Eckroade RJ, Dessel BH. Cultivation of papova-like virus from human brain with progressive multifocal leucoencephalopathy. Lancet 1971; 1:1257-60.
- Knowles WA, Pipkin P, Andrews N, Vyse A, Minor P, Brown DW, et al. Population-based study of antibody to the human polyomaviruses BKV and JCV and the simian polyomavirus SV40. J Med Virol 2003;71:115-23.
- Stolt A, Sasnauskas K, Koskela P, Lehtinen M, Dillner J. Seroepidemiology of the human polyomaviruses. J Gen Virol 2003;84:1499-504.
- Elsner C, Dorries K. Evidence of human polyomavirus BK and JC infection in normal brain tissue. Virology 1992;191:72-80.
- Grinnell BW, Padgett BL, Walker DL. Distribution of nonintegrated DNA from JC papovavirus in organs of patients with progressive multifocal leukoencephalopathy. J Infect Dis 1983;147:669-75.
- Sabath BF, Major EO. Traffic of JC virus from sites of initial infection to the brain: The path to progressive multifocal leukoencephalopathy. J Infect Dis 2002;186 Suppl 2:S180-6.
- Major EO, Amemiya K, Tornatore CS, Houff SA, Berger JR. Pathogenesis and molecular biology of progressive multifocal leukoencephalopathy, the JC virus-induced demyelinating disease of the human brain. Clin Microbiol Rev 1992;5:49-73.
- Gheuens S, Pierone G, Peeters P, Koralnik IJ. Progressive multifocal leukoencephalopathy in individuals with minimal or occult immunosuppression. J Neurol Neurosurg Psychiatry

- 2010;81:247-54.
- Berger JR. Progressive multifocal leukoencephalopathy and newer biological agents. Drug Saf 2010;33:969-983.
- Gentile S, Sacerdote I, Roccatello D, Giordana MT. Progressive multifocal leukoencephalopathy during cyclosporine treatment. A case report. Ital J Neurol Sci 1996;17:363-6.
- Tubridy N, Wells C, Lewis D, Schon F. Unsuccessful treatment with cidofovir and cytarabine in progressive multifocal leukoencephalopathy associated with dermatomyositis. J R Soc Med 2000;93:374-5.
- Vulliemoz S, Lurati-Ruiz F, Borruat FX, Delavelle J, Koralnik IJ, Kuntzer T, et al. Favourable outcome of progressive multifocal leucoencephalopathy in two patients with dermatomyositis.
 J Neurol Neurosurg Psychiatry 2006;77:1079-82.
- Aksamit AJ. Treatment of non-AIDS progressive multifocal leukoencephalopathy with cytosine arabinoside. J Neurovirol 2001;7:386-90.
- Epker JL, van Biezen P, van Daele PL, van Gelder T, Vossen A, van Saase JL. Progressive multifocal leukoencephalopathy, a review and an extended report of five patients with different immune compromised states. Eur J Intern Med 2009;20:261-7.
- Richardson EP Jr, Johnson PC. Atypical progressive multifocal leukoencephalopathy with plasma-cell infiltrates. Acta Neuropathol Suppl 1975;Suppl 6:247-50.
- Marie I, Guegan-Massardier E, Levesque H. Progressive multifocal leukoencephalopathy in refractory polymyositis treated with rituximab. Eur J Intern Med 2011;22:e13-4.
- Linda H, von Heijne A, Major EO, Ryschkewitsch C, Berg J, Olsson T, et al. Progressive multifocal leukoencephalopathy after natalizumab monotherapy. N Engl J Med 2009;361:1081-7.
- Landry ML, Eid T, Bannykh S, Major E. False negative PCR despite high levels of JC virus DNA in spinal fluid: Implications for diagnostic testing. J Clin Virol 2008;43:247-9.
- Clifford DB, De Luca A, Simpson DM, Arendt G, Giovannoni G, Nath A. Natalizumab-associated progressive multifocal leukoencephalopathy in patients with multiple sclerosis: Lessons from 28 cases. Lancet Neurol 2010;9:438-46.
- Eggers C, Stellbrink HJ, Buhk T, Dorries K. Quantification of JC virus DNA in the cerebrospinal fluid of patients with human immunodeficiency virus-associated progressive multifocal leukoencephalopathy A longitudinal study. J Infect Dis 1999; 180:1690-4.
- Koralnik IJ, Boden D, Mai VX, Lord CI, Letvin NL. JC virus DNA load in patients with and without progressive multifocal leukoencephalopathy. Neurology 1999;52:253-60.
- Oddis CV, Reed AM, Aggarwal R. Rituximab in the treatment of refractory adult and juvenile dermatomyositis (DM) and adult polymyositis (PM) — The RIM study. Arthritis Res 2010;62 Suppl 12:3844.
- Fauci AS, ed. Harrison's rheumatology. Harrison's principles of internal medicine. 16th ed. New York: McGraw-Hill Medical Publishing Division; 2004.
- Brickelmaier M, Lugovskoy A, Kartikeyan R, Reviriego-Mendoza MM, Allaire N, Simon K, et al. Identification and characterization of mefloquine efficacy against JC virus in vitro. Antimicrob Agents Chemother 2009;53:1840-9.
- Hall CD, Dafni U, Simpson D, Clifford D, Wetherill PE, Cohen B, et al. Failure of cytarabine in progressive multifocal leukoencephalopathy associated with human immunodeficiency virus infection. AIDS Clinical Trials Group 243 Team. N Engl J Med 1998;338:1345-51.
- Vermersch P, Kappos L, Gold R, Foley JF, Olsson T, Cadavid D, et al. Clinical outcomes of natalizumab-associated progressive multifocal leukoencephalopathy. Neurology 2011;76:1697-704.
- 28. Gorelik L, Lerner M, Bixler S, Crossman M, Schlain B, Simon K,

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2012. All rights reserved.

- et al. Anti-JC virus antibodies: Implications for PML risk stratification. Ann Neurol 2010;68:295-303.
- Rudick RA, O'Connor PW, Polman CH, Goodman AD, Ray SS, Griffith NM, et al. Assessment of JC virus DNA in blood and urine from natalizumab-treated patients. Ann Neurol 2010;68:304-10.
- Iacobaeus E, Ryschkewitsch C, Gravell M, Khademi M, Wallstrom E, Olsson T, et al. Analysis of cerebrospinal fluid and cerebrospinal fluid cells from patients with multiple sclerosis for detection of JC virus DNA. Mult Scler 2009;15:28-35.

J Rheumatol 2012;39:6; doi:10.3899/jrheum.111126

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2012. All rights reserved.

Letter 1303