

The Association Between Serum 25-hydroxy Vitamin D Level and Upper Leg Strength in Patients with Knee Osteoarthritis: Results of the Amsterdam Osteoarthritis Cohort

Esmee Koeckhoven, Marike van der Leeden, Leo D. Roorda, Natasja M. van Schoor, Paul Lips, Arjan de Zwart, Joost Dekker, Martin van der Esch, and Willem F. Lems

ABSTRACT. Objective. Vitamin D deficiency, which is common among elderly people, has been linked to muscle weakness. In patients with knee osteoarthritis (OA), the association between muscle strength and serum 25-hydroxy Vitamin D [25(OH)D] level has not been studied comprehensively. The aim of our study was to examine the association between serum 25(OH)D level and muscle strength in patients with knee OA.

Methods. Data of the Amsterdam Osteoarthritis cohort from 319 participants with knee OA were used in a cross-sectional study. Serum 25(OH)D level (nmol/l) was measured by a competitive electrochemiluminescence method. Muscle strength (nm/kg) of the upper leg was measured isokinetically. Univariable and multivariable linear regression analyses were used to calculate the association between serum 25(OH)D level and muscle strength.

Results. Serum 25(OH)D level was significantly associated with muscle strength ($B = 0.036$, 95% CI 0.017–0.054, $p < 0.001$), adjusted for season of blood collection. After adding body mass index (BMI) to the model, this association was no longer significant ($B = 0.011$, 95% CI –0.007 to 0.029, $p = 0.214$). Alcohol consumption, number of comorbidities, and sex were subsequently added and changed the model slightly. Without BMI, this model showed a significant association between serum 25(OH)D level and muscle strength ($B = 0.029$, 95% CI 0.014–0.043, $p < 0.001$).

Conclusion. The observed association between a low serum 25(OH)D level and muscle weakness in patients with knee OA is attenuated by BMI. Further studies are needed to explain the associations among Vitamin D level, muscle strength, and adiposity in patients with knee OA. (J Rheumatol First Release May 15 2016; doi:10.3899/jrheum.150751)

Key Indexing Terms:

VITAMIN D MUSCLE STRENGTH OSTEOARTHRITIS BODY MASS INDEX

From the Amsterdam Rehabilitation Research Center/Reade; Department of Rehabilitation Medicine, and EMGO Institute for Health and Care Research, and Department of Epidemiology and Biostatistics, and Department of Psychiatry, VU University Medical Center; Reade, Department of Rheumatology, Amsterdam, the Netherlands.

E. Koeckhoven, MSc, Amsterdam Rehabilitation Research Center/Reade; M. van der Leeden, PhD, Amsterdam Rehabilitation Research Center/Reade, and Department of Rehabilitation Medicine, and EMGO Institute for Health and Care Research, VU University Medical Center; L.D. Roorda, PhD, Amsterdam Rehabilitation Research Center/Reade; N.M. van Schoor, PhD, EMGO Institute for Health and Care Research, and Department of Epidemiology and Biostatistics, VU University Medical Center; P. Lips, PhD, Department of Internal Medicine/Endocrinology, VU University Medical Center; A. de Zwart, MSc, Amsterdam Rehabilitation Research Center/Reade; J. Dekker, PhD, Department of Rehabilitation Medicine, and EMGO Institute for Health and Care Research, and Department of Psychiatry, VU University Medical Center; M. van der Esch, PhD, Amsterdam Rehabilitation Research Center/Reade; W.F. Lems, PhD, Department of Rheumatology, VU University Medical Center, and Amsterdam Rehabilitation Research Center/Reade.

Address correspondence to M. van der Leeden, Amsterdam Rehabilitation Research Center/Reade, P.O. Box 58271, 1040 HG Amsterdam, the Netherlands. E-mail: m.vd.leeden@reade.nl

Accepted for publication April 8, 2016.

Muscle weakness is a well-established risk factor for activity limitations in patients with knee osteoarthritis (OA)^{1,2}. Determinants of muscle weakness in patients with knee OA include knee pain, disuse, aging, and low-grade inflammation³. In addition, vitamin D deficiency has been linked to muscle weakness because of its direct and indirect effects on muscle cells⁴. The main source of vitamin D is ultraviolet radiation (UV), which synthesizes pre-vitamin D in the skin. Bound to the vitamin D binding protein, pre-vitamin D is transported to the liver where it is hydroxylated into 25(OH)D. Serum 25(OH)D level is generally measured to assess vitamin D status⁵.

The relationship between serum 25(OH)D level and muscle strength in patients with knee OA has been studied only once, to our knowledge⁶. This relatively small study, containing 56 participants younger than 60 years with knee OA and muscle weakness, showed that participants with vitamin D deficiency (< 50 nmol/l) had significantly lower quadriceps strength compared to participants with a

non-deficient serum 25(OH)D level. Vitamin D deficiency has also been related to pain and radiological progression in patients with knee OA^{7,8,9}, although no effect of supplementation could be detected on these outcomes¹⁰.

The observed relationship between vitamin D and muscle strength in knee OA is in line with several observational studies in general populations. A recent systematic review demonstrated that vitamin D deficiency, or hypovitaminosis D, was consistently associated with decrease in muscle function and performance and increase in disability in all ages except for very old individuals¹¹. As to the effect of vitamin D supplementation on muscle strength, there is evidence from a systematic review and metaanalysis that vitamin D supplementation has a small positive effect on muscle strength¹². Subgroup analysis showed a greater effect of vitamin D supplementation in subjects with a baseline serum 25(OH)D level below 30 nmol/l.

Evidence for the role of vitamin D on muscle strength is accumulating, but studies in patients with knee OA are scarce. Because muscle weakness is common in knee OA and of clinical importance, optimizing muscle strength is a focus of rehabilitation in these patients. The association between vitamin D and muscle strength in this population should be clarified to reveal whether low serum 25(OH)D level is a determinant of muscle strength that could be targeted. We hypothesized that a low level of serum 25(OH)D (≤ 50 nmol/l)^{13,14,15} is associated with muscle weakness in patients with knee OA.

The aim of our present study was to examine the cross-sectional association between serum 25(OH)D level and muscle strength in patients with knee OA in a large cohort, taking possible confounders into account.

MATERIALS AND METHODS

Study sample. Data were collected from the Amsterdam Osteoarthritis cohort, made up of participants with a clinical diagnosis of OA of the knee and/or hip according to the American College of Rheumatology criteria¹⁶ and who have been referred to an outpatient rehabilitation center (Reade, Centre for Rehabilitation and Rheumatology; Amsterdam, the Netherlands). Participants with OA of the knee and complete data of serum 25(OH)D level and muscle strength were included in our present study (Figure 1). Exclusion

criteria were total knee replacement or presence of rheumatoid arthritis or any other inflammatory arthritis (i.e., septic arthritis, crystal arthropathy/gout, or spondyloarthropathy). Patients were assessed by rheumatologists, radiologists, and rehabilitation physicians. Rheumatologists established the diagnosis of knee OA and checked the exclusion criteria. Radiologists scored the radiographs of the knee, and rehabilitation physicians assessed the patients on functioning. Demographic, radiographic, biomechanical, clinical, and psychosocial factors related to OA were assessed at the baseline visit to the rehabilitation center. Each participant provided written informed consent according to the declaration of Helsinki. The study was approved by the Medical Ethical Institutional Review Board of Reade. The sample for this study comprises patients recruited between January 2009 and June 2013.

Measurements. In non-fasting blood samples collected by venapuncture, 25-hydroxy vitamin D was measured. The serum was centrifuged and stored at -20°C . The blood samples were analyzed by a Roche Cobas-6000 analyzer [COBAS 25(OH)D total] using a competitive electrochemiluminescence method to measure serum 25(OH)D level. The interassay coefficient of variation was $< 6\%$. The 25(OH)D was released from its binding plasma protein and afterward 25(OH)D was incubated with 25(OH)D binding protein labeled with ruthenium, creating a complex with 25(OH)D and ruthenylated vitamin D binding protein¹⁷. The level of 25(OH)D in nmol/l was used for analyses.

Muscle strength. Muscle strength was tested using an isokinetic dynamometer (EnKnee, Enraf-Nonius), which measured extension and flexion of the knee at $60^{\circ}/\text{s}$ ¹⁸. The participants performed 12 repetitions in total: 3 repetitions per leg for knee extension and 3 repetitions per leg for knee flexion. The average of knee flexor strength and knee extensor strength of both legs was used in the analyses, representing upper leg muscle strength². Average muscle strength of both legs was used because we hypothesized vitamin D deficiency would affect muscle strength in both legs and because the correlation between muscle strength of the right and left leg was high ($r > 0.75$). Muscle strength was adjusted for body weight (nm/kg).

Potential confounders. Based on previous studies investigating the relationship between serum 25(OH)D level and muscle strength in older adults^{19,20,21,22,23,24,25}, the following potential confounders were considered relevant: season of blood collection, age, sex, body mass index (BMI), smoking habits, alcohol consumption, creatinine level, and number of comorbidities. Season of blood collection was dichotomized into summer (April–September) and winter (October–March)^{5,13,23}. Age was calculated from date of birth at the start of inclusion. BMI was calculated as body mass in kg divided by the square of height in meters (kg/m^2). Information on current smoking was assessed by a questionnaire and was reported as “yes” or “no.” Alcohol consumption was also assessed by a questionnaire and was classified into “never,” “ ≤ 8 consumptions per week,” and “ > 8 consumptions per week.” Creatinine level (in $\mu\text{mol}/\text{l}$) was measured by a Roche Cobas-6000 analyzer. The Cumulative Illness Rating Scale was used to collect information about the presence of chronic diseases. This rating scale

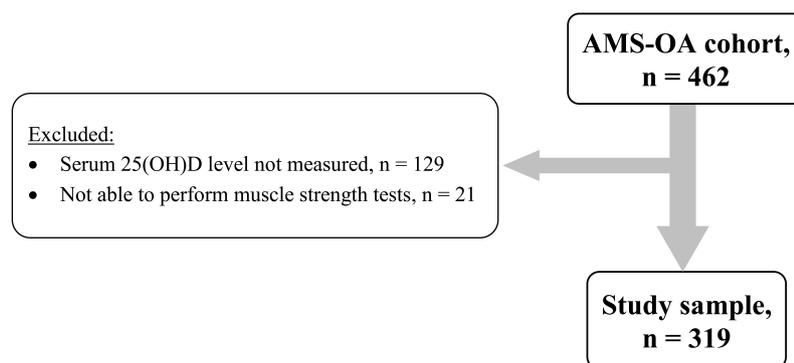


Figure 1. Scheme showing the total study sample and the exclusion of patients with knee osteoarthritis. AMS-OA: Amsterdam Osteoarthritis; 25(OH)D: 25-hydroxy vitamin D.

gathers information related to 13 body systems that can each be scored from 0 (none) to 4 (very severe). The number of body systems on which a patient scored 2 or higher was calculated and used in analyses.

Specific for the knee OA population, the following additional confounders were considered^{26,27}: erythrocyte sedimentation rate (ESR; in mm/h) and C-reactive protein (CRP; in mg/l). ESR values were determined by the standard Westergren method²⁸. The rate at which red blood cells sedimented in 1 h was measured and reported in mm/h. ESR was used as a dichotomized variable (≤ 20 mm/h and > 20 mm/h)²⁶. CRP was analyzed using turbidimetric immunoassay with CRPLX test kits²⁹ and was dichotomized (≤ 3 mg/l and > 3 mg/l)²⁶.

Other variables. Western Ontario and McMaster Universities Osteoarthritis Index (WOMAC) subscales of pain and physical functioning were used descriptively. The WOMAC is a self-administered questionnaire to assess pain, stiffness, and physical function in patients with hip and/or knee OA^{30,31}. WOMAC pain subscale was scored from 0 to 20 and WOMAC physical function subscale was scored from 0 to 68; higher scores represent more pain or activity limitations, respectively. Severity of radiological damage [Kellgren-Lawrence scale (KL)] was also assessed and used descriptively in our study. Radiographs of the tibiofemoral (medial and lateral) joints were made using a weight-bearing posterior-anterior view, semiflexed (7° – 10°) according to Buckland-Wright, *et al*³². The radiographic features (joint space narrowing, osteophyte formation, sclerosis, cysts) within the tibiofemoral joints were scored according to the Osteoarthritis Research Society International atlas in KL grades³³. Scoring of the KL grades was in 5 categories (0–4). Grade 0 represents no osteoarthritic features while grade 4 represents severe joint space narrowing, large osteophyte formation, sclerosis, and cysts.

Statistical analyses. Baseline characteristics for continuous variables [age, BMI, creatinine level, WOMAC physical functioning and pain subscales, muscle strength, and serum 25(OH)D level] were checked for normal distribution and reported as mean, SD, and min/max scores. Categorical variables (sex, season, smoking habits, alcohol consumption, no. comorbidities, ESR, CRP, and KL grade) were reported as percentages. Deficiency was labeled as a serum 25(OH)D level ≤ 50 nmol/l^{13,14,15}, and chi-squared tests and independent t tests were used to compare the patients with deficient levels and those without. To assess the association between muscle strength and serum 25(OH)D level, univariable linear regression analysis was performed. Muscle strength was used as a continuous variable whereas serum 25(OH)D level was used both continuously (10-unit change) and dichotomously (deficient vs non-deficient). The reported regression coefficients (B) represent the change in muscle strength by each 10-unit change of serum 25(OH)D level and the difference in muscle strength of being deficient vs being nondeficient, respectively. Also, the standardized regression coefficient (β) was reported, to assess the effect of various variables on this association in a standardized unit. In multivariable regression analyses, potential confounders were added stepwise into the model. First, season of blood collection was added to the model and was left in the model when testing other potential confounding variables. Variables that changed the regression coefficient (B) of serum 25(OH)D level by more than 10% were considered confounders. Interactions between age and serum 25(OH)D level and BMI and serum 25(OH)D level were considered plausible and were therefore tested. For this purpose, age was dichotomized into ≤ 60 vs > 60 years and BMI into ≤ 30 vs > 30 kg/m².

The same analyses were repeated using 75 nmol/l instead of 50 nmol/l as cutoff value for vitamin D deficiency^{13,14,15}, as well as analyses with extension and flexion strength separately. Data were analyzed using SPSS version 22. A p value ≤ 0.05 was considered significant.

RESULTS

The study sample comprised 319 participants (66.5% women) with a mean age of 60.5 (± 8.3) years (range 33–78 yrs). Table 1 shows the characteristics of the total sample, the

deficient group [serum 25(OH)D level ≤ 50 nmol/l], and the non-deficient group [serum 25(OH)D level > 50 nmol/l]. Overall, the mean serum 25(OH)D level was 60.8 (± 24.6) nmol/l (range 8–135), with 36.1% of the participants being deficient according to the cutoff point of ≤ 50 nmol/l. Using a higher cutoff point (75 nmol/l)¹³, deficiency was observed in 72.7% of the participants. Compared to the participants with a serum 25(OH)D level > 50 nmol/l, participants with a serum 25(OH)D level < 50 nmol/l showed lower muscle strength ($p = 0.008$), higher BMI ($p < 0.001$), higher ESR ($p = 0.011$), higher CRP ($p < 0.001$), and worse physical functioning ($p = 0.028$). In addition, 25(OH)D deficiency was more common in the winter season than in the summer season. Excluded participants [because of unknown serum 25(OH)D level ($n = 129$) and absence of data on muscle strength ($n = 21$); pain ($n = 3$), lack of strength ($n = 10$), and unknown ($n = 8$)] had a higher mean age ($p = 0.002$), worse physical functioning ($p = 0.002$), and higher pain scores ($p = 0.043$).

Table 2 shows the results of the univariable and multivariable linear regression analyses. Serum 25(OH)D level was significantly associated with muscle strength (B = 0.032, 95% CI 0.014–0.050, $p < 0.001$) in univariable analysis. Muscle strength remained significantly associated with serum 25(OH)D level after adjusting for season of blood collection (B = 0.036, 95% CI 0.017–0.054, $p < 0.001$; Model 1). When adding BMI to this model, the association between serum 25(OH)D level and muscle strength was no longer significant (B = 0.011, 95% CI –0.007 to 0.029, $p = 0.214$; Model 2). Model 3, adjusted for season of blood collection, BMI, alcohol consumption, number of comorbidities, and sex, was slightly different from Model 2 (B = 0.013, 95% CI –0.002 to 0.027, $p = 0.083$). In Model 4, the same confounding factors as in Model 3 were incorporated, except for BMI. This model showed a significant association between serum 25(OH)D level and muscle strength (B = 0.029, 95% CI 0.014–0.043, $p < 0.001$). Age, smoking habits, serum creatinine, ESR, and CRP did not change the regression coefficient $> 10\%$ in this model and were therefore omitted from the model. Higher BMI was associated with lower serum 25(OH)D levels (standardized β –0.340, $p < 0.001$) and lower muscle strength (standardized β –0.448, $p < 0.001$). No interaction effect between serum 25(OH)D level and age or serum 25(OH)D level and BMI was observed.

Analyses with 75 nmol/l, instead of 50 nmol/l, as a cutoff value for vitamin D deficiency, as well as analyses with extension and flexion strength separately showed similar results. Further, the frequency of missing data was not different between vitamin D deficient and vitamin D nondeficient participants (Table 1). Univariable and multivariable linear regression analyses using data from participants who had no missing data ($n = 293$) gave similar results (data not shown).

Table 1. Description of the characteristics of the total study sample and the differences of characteristics between deficient and nondeficient serum 25(OH)D levels in patients with knee osteoarthritis. Data are mean \pm SD (min–max) unless otherwise specified.

Characteristics	Total Sample, n = 319	Deficient, \leq 50 nmol/l, n = 115	Nondeficient, $>$ 50 nmol/l, n = 204	p*
%	100	36.1	63.9	
Age, yrs	60.5 \pm 8.3 (33–78)	59.9 \pm 9.1 (33–76)	60.8 \pm 7.8 (40–78)	0.376
Women, %	66.5	67.0	66.2	0.887
BMI, kg/m ²	30.2 \pm 6.0 (20.3–53.9)	32.8 \pm 6.9 (22.2–53.9)	28.7 \pm 4.9 (20.3–43.9)	< 0.001
Summer, %	50.8	39.1	57.4	0.002
Smoking, % yes ^a	7.2	5.6	8.0	0.420
Alcohol consumption, % ^b				0.030
% never	22.1	30.6	17.6	
% 0–8 servings/week	52.4	48.1	54.8	
% \geq 8 servings/week	25.4	21.3	27.6	
Creatinine, μ mol/l ^c	71.4 \pm 14.8 (41–155)	70.3 \pm 14.2 (41–116)	71.9 \pm 15.2 (45–155)	0.385
Comorbidities, n CIRS $>$ 2 ^d	1.0 \pm 1.3 (0–7)	1.1 \pm 1.4 (0–7)	0.9 \pm 1.2 (0–7)	0.312
% ESR $>$ 20 mm/h ^e	15.7	22.6	11.8	0.011
% CRP $>$ 3 mg/l ^f	30.4	42.6	23.5	< 0.001
Bilateral knee OA, %	72.6	75.6	67.3	0.138
KL grade (%) ^g				0.208
0	3.3	5.2	2.2	
1	29.8	34.4	27.4	
2	28.7	28.1	29.1	
3	23.6	16.7	27.4	
4	14.5	15.6	14.0	
WOMAC physical functioning, 0–68 ^h	26.2 \pm 12.7 (0–65)	28.3 \pm 13.5 (0–53)	25.0 \pm 12.0 (0–65)	0.028
WOMAC pain, 0–20 ⁱ	8.1 \pm 3.7 (0–18)	8.4 \pm 4.1 (0–17)	8.0 \pm 3.5 (0–18)	0.335
Total muscle strength, nm/kg	0.87 \pm 0.39 (0.1–2.1)	0.79 \pm 0.41 (0.1–2.1)	0.91 \pm 0.37 (0.1–1.9)	0.008
Extension	1.04 \pm 0.49 (0.1–2.6)	0.95 \pm 0.53 (0.2–2.6)	1.09 \pm 0.47 (0.1–2.4)	0.015
Flexion	0.69 \pm 0.30 (0.1–1.7)	0.62 \pm 0.31 (0.1–1.6)	0.73 \pm 0.30 (0.1–1.7)	0.002
25(OH)D, nmol/l	60.8 \pm 24.6 (8–135)	35.5 \pm 11.1 (8–50)	75.0 \pm 17.6 (51–135)	< 0.001

*p value: deficient vs non deficient. ^{a/b} 12 missing values; ^c 23 missing values; ^d 26 missing values; ^e 1 missing value; ^f 3 missing values; ^g 44 missing values; ^h 9 missing values; ⁱ 8 missing values. BMI: body mass index; CIRS: Cumulative Illness Rating Scale; ESR: erythrocyte sedimentation rate; CRP: C-reactive protein; KL: Kellgren-Lawrence arthritis rating scale; WOMAC: Western Ontario and McMaster Universities Osteoarthritis Index; 25(OH)D: 25-hydroxy vitamin D.

Table 2. Results of the regression analyses of serum 25(OH)D level on upper leg muscle strength, adjusted for potential confounders.

	N	25(OH)D Level: 10-unit Change				25(OH)D: Deficient vs Nondeficient ^a			
		B	β	p	95% CI	B	β	p	95% CI
Crude model	319	0.032	0.204	< 0.001	0.014–0.050	0.120	0.148	0.011	0.027–0.212
Model 1*	319	0.036	0.226	< 0.001	0.017–0.054	0.129	0.159	0.007	0.035–0.223
Model 2**	319	0.011	0.072	0.214	–0.007 to 0.029	0.007	0.009	0.880	–0.083 to 0.097
Model 3#	293	0.013	0.079	0.083	–0.002 to 0.027	0.012	0.015	0.735	–0.059 to 0.084
Model 4 [^]	293	0.029	0.181	< 0.001	0.014–0.043	0.094	0.116	0.012	0.021–0.167

^a \leq 50 nmol/l vs $>$ 50 nmol/l. * Adjusted for season. ** Adjusted for season and BMI. # Adjusted for season, BMI, alcohol consumption, no. comorbidities on CIRS, and sex. [^] Adjusted for season, alcohol consumption, no. comorbidities on CIRS, and sex. B: unstandardized regression coefficient; β : standardized regression coefficient. BMI: body mass index; CIRS: Cumulative Illness Rating Scale; 25(OH)D: 25-hydroxy vitamin D.

DISCUSSION

Our present study was the first, to our knowledge, to investigate the relationship between serum 25(OH)D level and upper leg strength in a large sample of patients with knee OA. A statistically significant association between a low serum 25(OH)D level and muscle weakness was found. However, this association was attenuated by BMI.

The observed association between serum 25(OH)D level and muscle strength is in line with the relatively small study

by Barker, *et al*⁶. Compared to that study, a larger sample of patients with knee OA was included in our present study. Further, in contrast to the study of Barker, *et al*, the association between serum 25(OH)D level and muscle strength was adjusted for several potential confounding factors. Our results showed that the difference between muscle strength in the deficient and nondeficient group was 0.12 nm/kg, indicating a 15% higher muscle strength in the nondeficient group compared to the deficient group. In exercise trials in knee

OA, an average increase in muscle strength of 17% was found, which led to improvements in daily functioning³⁴. We therefore believe that the difference in muscle strength between these groups is of clinical relevance.

Interestingly, BMI strongly affected the association between serum 25(OH)D level and muscle strength and the association was no longer significant when BMI was incorporated into the model. This may be explained by the established association between BMI and muscle strength in knee OA³⁵ and the finding that a reduction in BMI by weight loss resulted in higher muscle strength³⁶. This suggests that BMI has a strong influence on muscle strength, particular in patients with knee OA who have a high BMI (our study sample), which may be the result of fatty infiltration of muscle tissue. Moreover, it is known that 25(OH)D is stored in adipose tissue, because of its lipophilic character^{37,38}. More adipose tissue could consequently result in lower levels of 25(OH)D in serum. In that way, BMI could act as a preceding factor in this association, instead of a confounding factor, causing overcorrection of the model. In future research, stratifying by BMI in the analyses should be encouraged. Other causes for the effect of BMI could be that obese patients may be exposed to sunshine less frequently, owing to limited mobility or clothing habits, or that 25(OH)D production and metabolism are altered in obesity^{37,38}. Further studies are needed to investigate the influence of various factors³⁹, including BMI, on 25(OH)D.

Some limitations of our study have to be considered. First, a cross-sectional study design was used and a causal relationship cannot be established. For causal relationships, an experimental study design is needed. Second, no outdoor activity variable was available in our data. In patients with knee OA, muscle weakness may reduce outdoor activities, which may affect the 25(OH)D level (shortage in UV-exposure) resulting in a lower serum 25(OH)D level. Third, muscle strength was adjusted for body weight (nm/kg). Because muscle strength was adjusted for body weight and BMI was added as a confounding factor into the model, body weight was corrected twice in the model. Because of this double correction, the regression analyses were repeated with muscle strength in Newton meter, showing similar results (data not shown). Further, the majority of the participants excluded because of absence of data on muscle strength (n = 21) had vitamin D deficiency. If these participants were included, this could have increased the between-group difference in muscle strength. Finally, the serum 25(OH)D level was measured by a Roche Cobas-6000 analyzer. Janssen, *et al*⁴⁰ compared several methods — including the Roche Cobas analyzer — with the gold standard (ID–LC–MS/MS) to measure 25(OH)D level in serum. Overall, the Roche Cobas analyzer showed no significant differences in serum 25(OH)D level measured compared to the gold standard. On the other hand, at increasing levels of serum 25(OH)D, it showed an increasing measurement error.

Therefore, this method of measuring serum 25(OH)D level could have affected the vitamin D status measured in our study. However, the average serum 25(OH)D level of our study sample was similar to serum 25(OH)D levels presented in other studies in the same age group in Western Europe^{37,39,41,42}.

Because muscle strength is a risk factor for activity limitation in patients with knee OA^{1,2}, it is important to increase the effectiveness of muscle strengthening exercises in patients with knee OA. According to our results, serum 25(OH)D level could be a factor affecting muscle strength and could therefore be of additive value in the management of patients with knee OA.

The observed association between a low serum 25(OH)D level and muscle weakness in patients with knee OA is attenuated by BMI. Further studies are needed to explain the associations between muscle strength, 25(OH)D, and adiposity in patients with knee OA.

ACKNOWLEDGMENT

We gratefully acknowledge S. Romviel, M. Steenbergen, M. Crins, I. Schaffers, and S. Webster for collecting data.

REFERENCES

1. Dekker J, van Dijk GM, Veenhof C. Risk factors for functional decline in osteoarthritis of the hip or knee. *Curr Opin Rheumatol* 2009;21:520-4.
2. van der Esch M, Holla JF, van der Leeden M, Knol DL, Lems WF, Roorda LD, et al. Decrease of muscle strength is associated with increase of activity limitations in early knee osteoarthritis: 3-year results from the cohort hip and cohort knee study. *Arch Phys Med Rehabil* 2014;95:1962-8.
3. Bennell KL, Hunt MA, Wrigley TV, Lim BW, Hinman RS. Role of muscle in the genesis and management of knee osteoarthritis. *Rheum Dis Clin North Am* 2008;34:731-54.
4. Girgis CM, Clifton-Bligh RJ, Hamrick MW, Holick MF, Gunton JE. The roles of vitamin D in skeletal muscle: form, function, and metabolism. *Endocr Rev* 2013;34:33-83.
5. Lips P. Relative value of 25(OH)D and 1,25(OH)2D measurements. *J Bone Miner Res* 2007;22:1668-71.
6. Barker T, Henriksen VT, Rogers VE, Aguirre D, Trawick RH, Lynn Rasmussen G, et al. Vitamin D deficiency associates with gamma-tocopherol and quadriceps weakness but not inflammatory cytokines in subjects with knee osteoarthritis. *Redox Biol* 2014;2:466-74.
7. McAlindon TE, Felson DT, Zhang Y, Hannan MT, Aliabadi P, Weissman B, et al. Relation of dietary intake and serum levels of vitamin D to progression of osteoarthritis of the knee among participants in the Framingham Study. *Ann Intern Med* 1996;125:353-9.
8. Laslett LL, Quinn S, Burgess JR, Parameswaran V, Winzenberg TM, Jones G, et al. Moderate vitamin D deficiency is associated with changes in knee and hip pain in older adults: a 5-year longitudinal study. *Ann Rheum Dis* 2014;73:697-703.
9. Zhang FF, Driban JB, Lo GH, Price LL, Booth S, Eaton CB, et al. Vitamin D deficiency is associated with progression of knee osteoarthritis. *J Nutr* 2014;144:2002-8.
10. McAlindon T, LaValley M, Schneider E, Nuite M, Lee JY, Price LL, et al. Effect of vitamin D supplementation on progression of knee pain and cartilage volume loss in patients with symptomatic osteoarthritis: a randomized controlled trial. *JAMA* 2013;309:155-62.

11. Halfon M, Phan O, Teta D. Vitamin D: a review on its effects on muscle strength, the risk of fall, and frailty. *Biomed Res Int* 2015;2015:953241.
12. Beaudart C, Buckinx F, Rabenda V, Gillain S, Cavalier E, Slomian J, et al. The effects of vitamin D on skeletal muscle strength, muscle mass, and muscle power: a systematic review and meta-analysis of randomized controlled trials. *J Clin Endocrinol Metab* 2014;99:4336-45.
13. Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, et al. Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2011;96:1911-30.
14. IOM (Institute of Medicine). Dietary reference intakes for calcium and Vitamin D. Washington, DC: National Academies Press; 2011.
15. Mithal A, Wahl DA, Bonjour JP, Burckhardt P, Dawson-Hughes B, Eisman JA, et al. Global vitamin D status and determinants of hypovitaminosis D. *Osteoporos Int* 2009;20:1807-20.
16. Altman R, Asch E, Bloch D, Bole G, Borenstein D, Brandt K, et al. Development of criteria for the classification and reporting of osteoarthritis. Classification of osteoarthritis of the knee. Diagnostic and Therapeutic Criteria Committee of the American Rheumatism Association. *Arthritis Rheum* 1986;29:1039-49.
17. Roche Diagnostics. Elecsys® Vitamin D total assay. [Internet. Accessed April 12, 2016.] Available from: www.cobas.com/content/dam/cobas_com/pdf/product/Elecsys-Vitamin-D-total-assay/Elecsys%20Vitamin%20D%20total%20Factsheet.pdf
18. van der Esch M, Steultjens M, Knol DL, Dinant H, Dekker J. Joint laxity and the relationship between muscle strength and functional ability in patients with osteoarthritis of the knee. *Arthritis Rheum* 2006;55:953-9.
19. Bischoff HA, Stahelin HB, Tyndall A, Theiler R. Relationship between muscle strength and vitamin D metabolites: are there therapeutic possibilities in the elderly? *Z Rheumatol* 2000;59 Suppl 1:39-41.
20. Gerdhem P, Ringsberg KA, Obrant KJ, Akesson K. Association between 25-hydroxy vitamin D levels, physical activity, muscle strength and fractures in the prospective population-based OPRA Study of Elderly Women. *Osteoporos Int* 2005;16:1425-31.
21. Visser M, Deeg DJ, Lips P, Longitudinal Aging Study Amsterdam. Low vitamin D and high parathyroid hormone levels as determinants of loss of muscle strength and muscle mass (sarcopenia): the Longitudinal Aging Study Amsterdam. *J Clin Endocrinol Metab* 2003;88:5766-72.
22. Tieland M, Brouwer-Brolsma EM, Nienaber-Rousseau C, van Loon LJ, De Groot LC. Low vitamin D status is associated with reduced muscle mass and impaired physical performance in frail elderly people. *Eur J Clin Nutr* 2013;67:1050-5.
23. Wicherts IS, van Schoor NM, Boeke AJ, Visser M, Deeg DJ, Smit J, et al. Vitamin D status predicts physical performance and its decline in older persons. *J Clin Endocrinol Metab* 2007;92:2058-65.
24. Sohl E, de Jongh RT, Heijboer AC, Swart KM, Brouwer-Brolsma EM, Enneman AW, et al. Vitamin D status is associated with physical performance: the results of three independent cohorts. *Osteoporos Int* 2013;24:187-96.
25. Mathei C, Van Pottelbergh G, Vaes B, Adriaensen W, Gruson D, Degryse JM. No relation between vitamin D status and physical performance in the oldest old: results from the Belfrail study. *Age Ageing* 2013;42:186-90.
26. Sanchez-Ramirez DC, van der Leeden M, van der Esch M, Gerritsen M, Roorda LD, Verschueren S, et al. Association of serum C-reactive protein and erythrocyte sedimentation rate with muscle strength in patients with knee osteoarthritis. *Rheumatology* 2013;52:727-32.
27. Calton EK, Keane KN, Newsholme P, Soares MJ. The impact of vitamin D levels on inflammatory status: a systematic review of immune cell studies. *PLoS One* 2015;10:e0141770.
28. Patton WN, Meyer PJ, Stuart J. Evaluation of sealed vacuum extraction method (Seditainer) for measurement of erythrocyte sedimentation rate. *J Clin Pathol* 1989;42:313-7.
29. Price CP, Trull AK, Berry D, Gorman EG. Development and validation of a particle-enhanced turbidimetric immunoassay for C-reactive protein. *J Immunol Methods* 1987;99:205-11.
30. Bellamy N, Buchanan WW, Goldsmith CH, Campbell J, Stitt LW. Validation study of WOMAC: a health status instrument for measuring clinically important patient relevant outcomes to antirheumatic drug therapy in patients with osteoarthritis of the hip or knee. *J Rheumatol* 1988;15:1833-40.
31. Roorda LD, Jones CA, Waltz M, Lankhorst GJ, Bouter LM, van der Eijken JW, et al. Satisfactory cross cultural equivalence of the Dutch WOMAC in patients with hip osteoarthritis waiting for arthroplasty. *Ann Rheum Dis* 2004;63:36-42.
32. Buckland-Wright JC, Wolfe F, Ward RJ, Flowers N, Hayne C. Substantial superiority of semiflexed (MTP) views in knee osteoarthritis: a comparative radiographic study, without fluoroscopy, of standing extended, semiflexed (MTP), and schuss views. *J Rheumatol* 1999;26:2664-74.
33. Kellgren JH, Lawrence JS. Radiological assessment of rheumatoid arthritis. *Ann Rheum Dis* 1957;16:485-93.
34. Lange AK, Vanwanseele B, Fiatarone Singh MA. Strength training for treatment of osteoarthritis of the knee: a systematic review. *Arthritis Rheum* 2008;59:1488-94.
35. Maly MR, Calder KM, Macintyre NJ, Beattie KA. Relationship of intermuscular fat volume in the thigh with knee extensor strength and physical performance in women at risk of or with knee osteoarthritis. *Arthritis Care Res* 2013;65:44-52.
36. Henriksen M, Christensen R, Danneskiold-Samsøe B, Bliddal H. Changes in lower extremity muscle mass and muscle strength after weight loss in obese patients with knee osteoarthritis: a prospective cohort study. *Arthritis Rheum* 2012;64:438-42.
37. Wortsman J, Matsuoka LY, Chen TC, Lu Z, Holick MF. Decreased bioavailability of vitamin D in obesity. *Am J Clin Nutr* 2000;72:690-3.
38. Cipriani C, Pepe J, Piemonte S, Colangelo L, Cilli M, Minisola S. Vitamin d and its relationship with obesity and muscle. *Int J Endocrinol* 2014;2014:841248.
39. van Dam RM, Snijder MB, Dekker JM, Stehouwer CD, Bouter LM, Heine RJ, et al. Potentially modifiable determinants of vitamin D status in an older population in the Netherlands: the Hoorn Study. *Am J Clin Nutr* 2007;85:755-61.
40. Janssen MJ, Wielders JP, Bekker CC, Boesten LS, Buijs MM, Heijboer AC, et al. Multicenter comparison study of current methods to measure 25-hydroxyvitamin D in serum. *Steroids* 2012;77:1366-72.
41. van Schoor NM, Knol DL, Deeg DJ, Peters FP, Heijboer AC, Lips P. Longitudinal changes and seasonal variations in serum 25-hydroxyvitamin D levels in different age groups: results of the Longitudinal Aging Study Amsterdam. *Osteoporos Int* 2014;25:1483-91.
42. Hilger J, Friedel A, Herr R, Rausch T, Roos F, Wahl DA, et al. A systematic review of vitamin D status in populations worldwide. *Br J Nutr* 2014;111:23-45.