# Tubular Urate Transporter Gene Polymorphisms Differentiate Patients with Gout Who Have Normal and Decreased Urinary Uric Acid Excretion

Rosa J. Torres, Eugenio de Miguel, Rebeca Bailén, José R. Banegas, and Juan G. Puig

ABSTRACT. Objective. Primary gout has been associated with single-nucleotide polymorphisms (SNP) in several tubular urate transporter genes. No study has assessed the association of reabsorption and secretion urate transporter gene SNP with gout in a single cohort of documented primary patients with gout carefully subclassified as normoexcretors or underexcretors.

*Methods.* Three reabsorption SNP (SLC22A12/URAT1, SLC2A9/GLUT9, and SLC22A11/OAT4) and 2 secretion transporter SNP (SLC17A1/NPT1 and ABCG2/BRCP) were studied in 104 patients with primary gout and in 300 control subjects. The patients were subclassified into normoexcretors and underexcretors according to their serum and 24-h urinary uric acid levels under strict conditions of dietary control.

**Results.** Compared with control subjects, patients with gout showed different allele distributions of the 5 SNP analyzed. However, the diagnosis of underexcretor was only positively associated with the presence of the T allele of *URAT1* rs11231825, the G allele of *GLUT9* rs16890979, and the A allele of *ABCG2* rs2231142. The association of the A allele of *ABCG2* rs2231142 in normoexcretors was 10 times higher than in underexcretors. The C allele of *NPT1* rs1165196 was only significantly associated with gout in patients with normal uric acid excretion.

*Conclusion.* Gout with uric acid underexcretion is associated with transporter gene SNP related mainly to tubular reabsorption, whereas uric acid normoexcretion is associated only with tubular secretion SNP. This finding supports the concept of distinctive mechanisms to account for hyperuricemia in patients with gout with reduced or normal uric acid excretion. (J Rheumatol First Release Aug 15 2014; doi:10.3899/jrheum.140126)

Key Indexing Terms: GOUT

GENE POLYMORPHISM

ARTHRITIS

Increased serum urate levels in gout may be the result of enhanced purine synthesis, decreased uric acid excretion, or both<sup>1</sup>. In most patients with primary gout, hyperuricemia has been related to a reduced renal uric acid excretion (underexcretors)<sup>2,3</sup>. In contrast, increased uric acid synthesis is suspected in normoexcretors with gout in whom the renal handling of uric acid is presumed to be normal.

The renal handling of uric acid is a complex process involving glomerular filtration and both tubular reabsorp-

Supported by grants from the Fondo de Investigación Sanitaria del Instituto de Salud Carlos III (Healthcare Research Fund of the Carlos III Health Institute; FIS, 08/0009 and 11/0598).

*R.J.* Torres, MD, PhD, Department of Biochemistry; E. de Miguel, MD, PhD, Department of Rheumatology; R. Bailén, MD; J.G. Puig, MD, PhD, Department of Internal Medicine, Metabolic-Vascular Unit, La Paz University Hospital; J.R. Banegas, MD, PhD, Department of Epidemiology and Public Health, Madrid Autonoma University.

Address correspondence to Dr. R. Torres, Servicio de Bioquímica, Edificio de Laboratorios, Planta BAJA, Hospital Universitario "La Paz," Paseo de la Castellana 261, 28046 Madrid, Spain. E-mail: rtorres.hulp@salud.madrid.org Accepted for publication May 21, 2014. tion and secretion, all of which determine a normal net uric acid excretion of almost 10% of the filtered load of urate<sup>2</sup>. In the last decade, several tubular urate transporters have been described that influence either urate reabsorption or secretion<sup>4</sup>.

Several studies, including a number of genome-wide association studies, have identified a substantial association between single-nucleotide polymorphisms (SNP) in as many as 28 genetic loci, including 6 urate-transporter-coding genes (URAT1/SLC22A12, GLUT9/SLC2A9, ABCG2/BCRP, SLC22A11/OAT4, SLC17A1/NPT1, and SLC17A3/NPT4), and serum urate concentrations<sup>5,6,7,8,9,10,11,12</sup>. The essential role of URAT1 and GLUT9 transporters in proximal tubular urate reabsorption has been underscored by their marked functional deficiency in patients with renal hypouricemia (OMIM 220150 and 612076, respectively) attributable to renal urate wasting<sup>13,14,15</sup>. Data have been published relative to gout association with URAT1, GLUT9, ABCG2, OAT4, and SLC17A1/NPT1 SNP; however, a number of studies have shown certain discrepancies. For instance, among several SNP at 8 genetic loci with a significant association with serum urate levels, Yang, et al<sup>9</sup> reported that only 2 loci (GLUT9 and ABCG2) were significantly

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2014. All rights reserved.

Torres, et al: A genetic study in gout

From the Department of Biochemistry, Department of Rheumatology, Department of Internal Medicine, Metabolic-Vascular Unit, La Paz University Hospital; Department of Epidemiology and Public Health, Madrid Autonoma University, IdiPaz, Madrid, Spain.

associated with gout. Several studies<sup>11,16</sup> suggested that NPT1 polymorphisms are associated with gout; however, Stark, et al<sup>5</sup> did not find such an association. It is remarkable that in a number of these studies, the diagnosis of gout was self-reported<sup>5,6,17</sup>, and no study, to our knowledge, has yet assessed the association of several SNP in a single cohort of well-characterized patients with gout classified into normoexcretors and underexcretors. In our study, we analyzed 5 previously gout-related SNP, corresponding to 5 different reabsorption and secretion tubular urate transporter-related genes (Figure 1) in normal subjects and in patients with primary gout, diagnosed according to the American College of Rheumatology criteria, and classified into normoexcretors and underexcretors following a protocol that included renal uric acid excretion rates and serum urate concentrations determined under strict dietary control.

#### MATERIALS AND METHODS

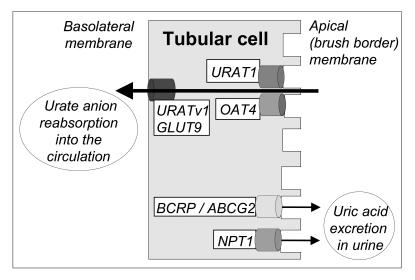
*Patients*. Patients with primary gout were recruited from the outpatient clinic of the Metabolic-Vascular Unit, Division of Internal Medicine at La Paz University Hospital, Madrid. Primary gout was diagnosed according to the American College of Rheumatology criteria<sup>18</sup>. Only patients with 2 or more acute arthritis episodes documented by a physician were selected for our study (n = 141). Secondary gout attributable to enzymopathies was discarded by determination of phosphoribosylpyrophosphate synthetase and hypoxanthine-guanine phosphoribosyltransferase activities in erythrocyte lysates by high-performance liquid chromatography, as previously described<sup>19,20</sup>. The main exclusion criteria were secondary gout (i.e.,

psoriasis, n = 4), marked organ insufficiency that precluded participation on an ambulatory basis (n = 10), and unwillingness to participate (n = 9). All studies were conducted according to the Declaration of Helsinki and were approved by the institutional research and ethics review committees of La Paz University Hospital. Medications that might affect uric acid metabolism were discontinued for a 3-month period prior to the start of our study. Colchicine (0.5 to 1.0 mg/day) was prescribed as a prophylaxis against acute gouty arthritis. All patients signed informed consent forms.

A specialized nurse provided all subjects with detailed instructions on following a weight-maintenance, isocaloric, purine-restricted diet (< 75 mg/24 h of purines, with a protein content of 10%-15%) for 5 days before the 24-h urine sample collection. At the end of the 24-h urine collection and following an overnight fast and rest, venous blood was obtained between 7 AM and 10 AM for uric acid and creatinine determination and DNA extraction. Patients with serum urate levels < 6.5 mg/dl following the 5-day purine-restricted diet were excluded from our study (n = 6). Subjects who did not collect urine appropriately were scheduled for a second visit the following day and instructed to continue the same purine-free diet. None of the subjects had lost  $\geq 1$  kg by the end of the purine-free diet period. Patients with a 24-h creatinine clearance  $< 45 \text{ ml/min}/1.73 \text{ m}^2$  were excluded from our study (n = 8). According to a constructed nomogram<sup>21</sup>, patients with gout were classified into 2 groups: normoexcretors and underexcretors. This nomogram takes into account the individual body surface area and serum urate concentration, the main variable determining urinary uric acid excretion.

Comorbidities were assessed following the definition proposed by the European Society of Hypertension<sup>22</sup>. The control group included DNA and plasma from 300 healthy subjects from the Spanish National DNA Bank. The Spanish National DNA Bank receives, processes, and stores DNA, plasma, and cell samples from donors along with relevant information on health and lifestyle habits related to the samples.

*Methods*. Uric acid and creatinine levels were determined by means of the uricase enzymatic and Jaffé methods, respectively. The RNA-free genomic



*Figure 1*. Schematic diagram showing the 5 tubular urate transporter genes studied in the current model of bidirectional urate anion movement in proximal tubule epithelial cells. Urate reabsorption at the apical membrane is critically regulated by SLC22A12/URAT1, which exchanges urate for intracellular organic anions (e.g., lactate, nicotinate) and monocarboxylates (e.g., pyrazinamide metabolites). The organic acid transporter SLC22A11/OAT4 also contributes to urate reabsorption from the tubular lumen. The hexose transport facilitator SLC2A9/GLUT9 (URATv1) performs voltage-dependent urate anion reabsorption into the circulation at the basolateral membrane. The purine nucleoside transporter BCRP/ABCG2 and the voltage-driven transporter SLC17A1/NPT1 mediate urate secretion at the apical membrane.

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2014. All rights reserved.

The Journal of Rheumatology 2014; 41:9; doi:10.3899/jrheum.140126

DNA samples from 101 patients with gout were isolated from whole blood using a DNA Purification Kit (Puragene, Gentra systems). DNA samples from 300 control subjects were isolated with the same DNA Purification Kit.

Five SNP (rs11231825, rs16890979, rs17300741, rs1165196, and rs2231142) corresponding to 5 different tubular urate transporter-related genes (*URAT1, GLUT9, OAT4, NPT1*, and *ABCG2*, respectively), were analyzed in all subjects. These 5 SNP have previously been associated with serum uric acid levels<sup>6,7</sup> and primary gout<sup>5,6,8,9,10,11,12,23</sup>. Four SNP are in the coding region and 1 SNP is intronic (rs17300741; Table 1). SNP rs16890979, rs1165196, and rs2231142 are nonsynonymous and cause the V253I, T269I, and Q141L amino acid change, respectively, in their corresponding proteins (GLUT9, NPT1, and ABCG2).

Genotyping was performed on each 50-ng genomic DNA sample using the LightCycler 480 System (Roche), with the LightCycler 480 Genotyping Master. For each SNP, a set of primers and HybProbe probes was designed and obtained from TIB MOLBIOMOL (Table 1). The specificity of the amplified PCR product was assessed by performing a melting curve analysis to discriminate between the homozygote for the major frequency allele, the homozygote for the minor frequency allele, and the heterozygote products.

Statistical analysis. The analysis plan for our study specified a minor allele frequency of  $25\%^{24}$ , a significance level of 5% ( $\alpha$ ), and a 3:1 ratio (control/case) to provide about 95% power to detect a difference of  $\pm 20\%$ . Thus, a sample size of 400 subjects (300 controls and 100 cases) was estimated (Query Advisor program, version 5.0). Mean values with SD and percentages were used to describe the patient characteristics. The differences between groups were evaluated with t tests for continuous variables and a logistic-regression model for dichotomous variables.

The Hardy-Weinberg equilibrium<sup>25</sup> for genotypic frequencies for the 5 study SNP in the control subjects was calculated using the chi-squared test. The association of the 5 different SNP genotypes and serum urate levels was performed by 1-way ANOVA with a posthoc Bonferroni test. We used a Bonferroni correction of p < 0.00625, equivalent to p < 0.05 significance, to correct for multiple testing, assuming independent tests for the 5 SNP.

OR and 95% CI were calculated to assess the relationship between SNP and the diagnosis of primary gout and gout subgroups (normoexcretors and underexcretors). A multivariate logistic regression analysis was used to compare allele distribution of the 5 SNP in the different subject groups.

A genetic risk score was generated for every individual by counting the number of alleles of those genes that were associated with serum urate levels in our population (rs11231825 T of *URAT1*, rs16890979 G of *GLUT9*, and rs2231142 A of *ABCG2*; range 0 to 6 points) in control subjects and patients with primary gout<sup>6,9,26</sup>. The correlation between the risk score and serum urate concentrations was performed using a logistic regression model by means of the chi-squared test. A risk score comparison

between controls and patients with gout was conducted using the Mann-Whitney U test.

All p values are 2-sided, and p values of < 0.05 were considered to indicate statistical significance (Statview SAS version 9.2).

#### RESULTS

Participant characteristics. A total of 104 patients with primary gout completed our study. According to the uric acid nomogram<sup>21</sup>, 16 patients were classified as normoexcretors (15.4%) and 88 as underexcretors (84.6%; Table 2). The mean daily uric acid excretion was 344 mg/day/1.73 m<sup>2</sup> (95% CI, 310-378 mg/day/1.73 m<sup>2</sup>) higher in normoexcretors with gout than in underexcretors, despite a mean serum urate level 1.0 mg/dl lower in the former than in the latter (Table 2). The fractional excretion of uric acid (Cur/Ccr×100) in normoexcretors was a mean of 46% higher than in underexcretors (p < 0.001). Clinical characteristics were not significantly different between the 2 groups with gout except for age and renal function. Patients with uric acid underexcretion were on average 6.4 years older than normoexcretors, although the duration of the disease evolution was similar for the 2 groups (slightly above 12 yrs). The glomerular filtration rate was on average 10 ml/min/1.73 m<sup>2</sup> lower in underexcretors than in normoexcretors (Table 2), with 20 patients in the former group showing a 24-h creatinine clearance below 60 ml/min/1.73 m<sup>2</sup> (range 45–59 ml/min/1.73 m<sup>2</sup>). Mean serum urate levels and 24-h urinary uric acid excretion were not significantly different in patients with creatinine clearance levels below or above 60 ml/min/1.73 m<sup>2</sup> (8.5 mg/dl and  $402 \text{ mg/day}/1.73 \text{ m}^2 \text{ vs } 8.0 \text{ mg/dl}$  and  $542 \text{ mg/day}/1.73 \text{ m}^2$ , respectively).

SNP distribution in control subjects and patients with gout. All of the genotypic frequencies for the 5 study SNP complied with the Hardy-Weinberg equilibrium (p > 0.05) in control subjects<sup>25</sup>. A strong association was found in all study subjects between the presence of allele T of rs11231825 (*URAT1*), allele G of rs16890979 (*GLUT9*), and allele A of rs2231142 (*ABCG2*) and serum urate concentra-

Table 1. Primer sequences (5'-3') and probes used for amplification of SNP regions in 5 human genes that encode tubular urate transporters.

SNP	Gene	Localization/ reference	Amplicon Size	Primers	Probes			
rs11231825	SLC22A12/URATI	Chromosome 11	218	CCCTAGAGGTCACCAGACCA	CGTGTGTGACTCTCACGCTCT-FL			
		NM_144585.2		GGGGGTACTCACCTGTCTGA	LC640-AAGCCCATGGCCCAGTCCA-PH			
rs16890979	SLC2A9/GLUT 9	Chromosome 4	182	GACAATCACGGTGACCAC	CGGCTCTCAGCCAGGACCTCCTCT-FL			
		NM_001001290.	1	GCTGAAAGTCCATGTTGTC	L670-CCTCTTGGGAAACGTCTGC-PH			
rs17300741	SLC22A11/OAT4	Chromosome 11	277	CCAAGATCGCACCACTACA	TGGTGAAGCAGCCAACTC-FL			
		NM_018484.2		AGAATAAAGCTGAGTCCCAATG	LC640-AGGGACACCCATGCCCAGG-PH			
rs1165196	SLC17A1/NPT1	Chromosome 6	234	TTAATGAATTTCTCTCCAGGTCAG	AAAACGTAAAACTACCAATGGAAAT-FL			
		NM_005074.3		AATGTACACAGAGTCTTTCGTGA	LC640-CCCAGACTGGAAGCGACTTAAGT-PH			
rs2231142	BCRP/ABCG2	Chromosome 4	128	GTCTCATTAAAATGCTATTTGCCT	GCTGAGAACTTTAAGTTTTCTCTCACC-FL			
		NM_00482.2		GAATGACCCTGTTAATCCG	LC610-TCAGAGTGCCCATCACAACATCATCCT-PH			

SNP: single-nucleotide polymorphism.

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2014. All rights reserved.

Torres, et al: A genetic study in gout

	Patients with Gout, n = 104	Underexcretors, n = 88	Normoexcretors, n = 16	р	
Age, yrs	58.7 ± 8.9	59.8 ± 9.1	$53.4 \pm 6.8$	0.009	
Men, n (%)	100 (95)	84 (95)	16 (100)	0.998	
Comorbidities <sup>a</sup> , n (%)					
Smokers	12 (11)	11 (12)	1 (6)	0.687	
Alcohol <sup>b</sup>	36 (34)	33 (37)	3 (19)	0.252	
Clinical atherosclerosis <sup>c</sup>	12 (11)	12 (12)	0 (0)	0.321	
Arterial hypertension	78 (74)	69 (78)	9 (56)	0.116	
Type 2 diabetes mellitus	24 (23)	22 (24)	2 (13)	0.353	
Hyperlipidemia	62 (58)	55 (62)	7 (44)	0.269	
Overweight	22 (21)	19 (21)	3 (20)	0.999	
Obesity	73 (70)	62 (70)	11 (69)	0.999	
Metabolic syndrome	39 (37)	35 (39)	4 (25)	0.403	
Gout characteristics	0) (0))	00 (0))	. (20)	01102	
Age at diagnosis, yrs	$47.0 \pm 12.0$	$48.0 \pm 11.9$	$41.0 \pm 8.7$	0.027	
Gout disease, yrs	$12.0 \pm 9.0$	$12.1 \pm 8.9$	$12.5 \pm 9.1$	0.869	
Acute gout flares, per year	$1.0 \pm 1.0$	$1.0 \pm 1.0$	$0.5 \pm 0.4$	0.152	
Renal lithiasis, n (%)	9 (9)	6 (7)	3 (19)	0.137	
Upper limb involvement, n (%)	7 (7)	5 (6)	2 (13)	0.289	
Lower limb involvement, n (%)	101 (96)	87 (98)	14 (87)	0.109	
First MTP joint involvement, n (%)	69 (66)	58 (65)	11 (69)	0.781	
Polyarticular involvement, n (%)	7 (7)	6 (7)	1 (6)	0.997	
Physical examination	, (,)	0(1)	1 (0)	0.777	
Weight, kg	$83.8 \pm 8.6$	$83.3 \pm 8.7$	$83.9 \pm 6.8$	0.794	
Body mass index, $kg/m^2$	$29.2 \pm 2.9$	$29.2 \pm 2.9$	$28.9 \pm 2.8$	0.703	
Tophi, n (%)	2 (2)	2 (2)	0 (0)	0.999	
Systolic blood pressure, mmHg	$139 \pm 12.3$	$140.0 \pm 12.2$	$132.5 \pm 9.1$	0.021	
Diastolic blood pressure, mmHg	$84 \pm 6.6$	$84.4 \pm 7.0$	$81.9 \pm 3.7$	0.168	
Analysis	012010	0.111 = 7.10	010 = 011	01100	
Baseline serum urate, mg/dl	$8.8 \pm 0.9$	$9.0 \pm 0.9$	$7.5 \pm 0.85$	0.001	
Serum urate, purine-free diet, mg/dl	$8.1 \pm 1.0$	$8.3 \pm 1.1$	$7.0 \pm 0.58$	0.001	
Uric acid excretion, $mg/day/1.73 m^2$	$509 \pm 147$	$455 \pm 111$	$799 \pm 184$	0.001	
Cur/Ccr, %	$5.35 \pm 1.12$	$4.98 \pm 0.84$	$7.26 \pm 1.48$	0.001	
Total cholesterol, mg/dl	$200 \pm 32$	$199 \pm 32$	$200 \pm 24$	0.906	
Triglycerides, mg/dl	$172 \pm 78$	$175 \pm 71$	$165 \pm 76$	0.574	
Cholesterol HDL, mg/dl	$46 \pm 8$	$47 \pm 8$	$43 \pm 5$	0.130	
Cholesterol LDL, mg/dl	$127 \pm 25$	$126 \pm 25$	$130 \pm 22$	0.550	
Fasting glucose, mg/dl	$127 \pm 25$ $102 \pm 15$	$120 \pm 25$ $104 \pm 15$	$96 \pm 15$	0.052	
HbA1c, %	$5.6 \pm 0.3$	$5.6 \pm 0.3$	$5.5 \pm 0.3$	0.225	
Serum creatinine, mg/dl	$1.2 \pm 0.2$	$1.2 \pm 0.2$	$1.1 \pm 0.1$	0.004	
$GFR, ml/min/1.73 m^2$	$68.4 \pm 13.1$	$67.2 \pm 12.9$	$77.4 \pm 8.6$	0.003	
Microalbuminuria, mg/g creatinine	$20.9 \pm 19.9$	$20 \pm 20$	$6 \pm 3$	< 0.00	

*Table 2*. Clinical characteristics of patients with primary gout stratified into normal (normoexcretors) and diminished (underexcretors) urinary uric acid excretion related to their serum urate level. Data are mean  $\pm$  SD unless otherwise indicated.

<sup>a</sup> According to the definitions established by the European Society of Hypertension<sup>20</sup>. <sup>b</sup> Alcohol,  $\ge 2$  drinks per day or  $\ge 200$  g/week. <sup>c</sup> Clinical atherosclerosis includes the following: stroke, coronary artery disease, peripheral arterial disease, and renal insufficiency with a GFR < 60 m/min/1.73 m<sup>2</sup>. MTP: metatarsophalangeal; Cur/Ccr: fractional excretion of uric acid; HDL: high-density lipoprotein; LDL: low-density lipoprotein; GFR: glomerular filtration rate.

tions (Table 3). In contrast, no significant association was found between the presences of allele A of rs17300741 (*OAT4*) and allele C of rs1165196 (*NPT1*) and serum urate levels.

The allele distributions of all the SNP analyzed were significantly different when comparing the controls with all patients with gout as a group (Table 4). Two reabsorption transporter genetic loci, *URAT1* rs11231825 and *GLUT9* 

rs16890979, were positively associated with underexcretors with gout (OR 1.62, 95% CI 1.00–2.63, p = 0.04 and OR 3.78, 95% CI 1.09–13.12, p = 0.005, respectively). However, no reabsorption transporter genetic loci were associated with normoexcretors with gout. One secretion transporter SNP (rs2231142 of *ABCG2*) showed a positive association with gout both in underexcretors (OR 2.91, 95% CI 1.40–6.08, p = 0.005) and in normoexcretors (OR 28.95,

Personal non-commercial use only. The Journal of Rheumatology Copyright © 2014. All rights reserved.

The Journal of Rheumatology 2014; 41:9; doi:10.3899/jrheum.140126

*Table 3*. Associations of serum urate concentrations (mean  $\pm$  SD) with SNP in the study population [healthy controls (n = 300) and patients with primary gout (n = 101)]. The number of study participants is provided in parentheses. p = Bonferroni test. Numbers in bold face are significant data.

SNP Identified (Allele 1/Allele 2); Gene	Yes, Examined Allele*	No, Examined Allele	р
rs11231825 (C/T); URAT1 examined allele T	5.34 ± 2.23 (187)	4.79 ± 2.10 (213)	0.0121
rs16890979 (G/A); <i>SLC2A9</i> examined allele A	4.89 ± 2.02 (151)	$5.20 \pm 2.27$ (250)	0.1749
rs16890979 (G/A); <i>SLC2A9</i> examined allele G	5.13 ± 2.21 (386)	3.87 ± 0.88 (15)	0.0290
rs17300741(G/A); <i>OAT4</i> examined allele A	5.14 ± 2.23 (296)	$4.71 \pm 1.91$ (97)	0.0873
rs1165196 (T/C); <i>NPT1</i> examined allele C	$4.96 \pm 2.09$ (266)	5.39 ± 2.35 (132)	0.0630
rs2231142 (C/A); <i>ABCG2</i> examined allele A	$6.09 \pm 2.56$ (54)	4.92 ± 2.08 (347)	0.0003

\* Both heterozygous and homozygous. SNP: single-nucleotide polymorphism.

*Table 4*. Allele distribution of the analyzed SNP in the different groups (control, gout, UE and NE patients with gout). Table expresses percentage of the examined allele in heterozygosis and homozygosis; parentheses indicate values in each different group; n = no. subjects with the examined allele in heterozygosis or homozygosis versus the total number of subjects in the group (n/total). OR and 95% CI were calculated to assess the relationship between SNP and the diagnosis of primary gout and gout subgroups (UE and NE). A multivariate logistic regression analysis was used to compare the allele distributions of the risk allele of each SNP in the different subject groups. Numbers in bold face are significant data.

	Control	Gout	UE	NE	Control vs Gout		Control vs UE			Control vs NE			
SNP (allele), gene	% (n/total)	% (n/total)	% (n/total)	% (n/total)	OR	CI	р	OR	CI	р	OR	CI	р
rs11231825 (T),	50.1	62.4	62.3	62.5	1.631	1.038-2.561	0.0326	1.628	1.005-2.637	0.0482	1.635	0.599-4.465	0.3408
URAT1	(150/299)	(63/101)	(53/85)	(10/16)									
rs16890979 (G),	95.0	100	100	100	3.995	1.219-13.091	0.0029	3.788	1.093-13.129	0.0057	3.011	0.283-31.932	0.2062
SLC2A9	(285/300)	(101/101)	(85/85)	(16/16)									
rs17300741 (A),	72.5	84.0	82.5	92.8	1.850	1.081-3.165	0.0200	1.687	0.957-2.974	0.0725	2.822	0.839–9.485	0.0583
OAT4	(217/299)	(79/94)	(66/80)	(13/14)									
rs1165196 (C),	70.0	57.4	62.4	31.3	0.567	0.351-0.915	0.0217	0.701	0.418-1.177	0.1807	0.169	0.057-0.496	0.0031
NPT1	(208/299)	(58/101)	(53/85)	(5/16)									
rs2231142 (A),	9.7	24.8	21.2	43.8	3.637	1.880-7.036	0.0003	2.918	1.400-6.085	0.0052	28.954	5.962-140.619	0.0002
ABCG2	(29/300)	(25/101)	(18/85)	(7/16)									

SNP: single-nucleotide polymorphism; UE: underexcretors; NE: normoexcretors.

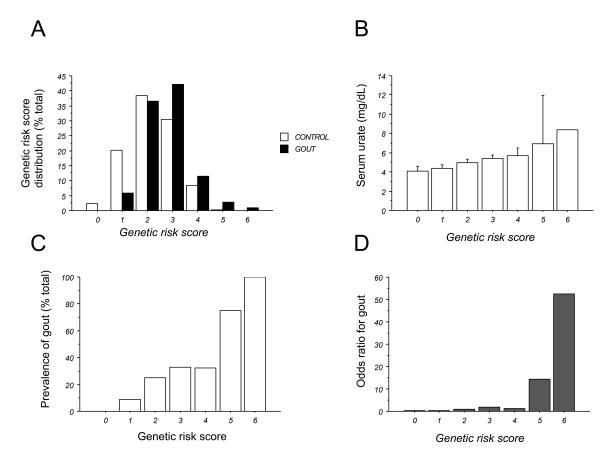
95% CI 5.96–140.6, p = 0.0002), although the OR for normoexcretors was 10 times higher. The other studied secretion transporter SNP, rs1165196 *NPT1*, showed a negative association only in normoexcretors (OR 0.16, 95% CI 0.05–0.49, p = 0.003).

*Genetic risk score*. An individual genetic risk score was generated by counting the number of alleles of those genes associated with serum urate levels in our population (rs11231825 allele T, rs16890979 allele G, and rs2231142 allele A; range 0 to 6 points according to the number of risk alleles). The proportion of subjects across the genetic risk score showed a markedly different distribution, with control subjects and patients with gout skewed to the lower and upper risk scores, respectively (Figure 2A). Mean serum urate concentrations in the entire population increased

linearly with the number of risk alleles (chi-square = 20.035, p = 0.0027; Figure 2B). In addition, the prevalence of gout also increased linearly with the number of risk alleles (Figure 2C). The OR for the diagnosis of primary gout, adjusted for age and sex, increased significantly for those patients with  $\geq$  3 risk alleles (Figure 2D). For subjects with no risk alleles, the crude prevalence of gout was 0.257 and increased to 52.478 for those with 6 risk alleles (208-fold increase).

#### DISCUSSION

Our study shows that patients with primary gout have a significantly increased prevalence of certain alleles for some SNP related to tubular urate transporter genes, which might help explain their uric acid metabolic disorder. Moreover,



*Figure 2*. Association of a genetic risk score, constructed with the number of hyperuricemia risk alleles, and the prevalence of gout and serum urate concentrations. (A) Proportion of control subjects and patients with primary gout across the genetic risk score. (B) Mean serum urate (mg/dl) levels and their 95% CI for each genetic risk score. (C) Crude prevalence of gout for each genetic risk score. (D) OR of gout for each genetic risk score.

tubular urate transporter SNP differentiate patients with gout with normal and decreased urinary uric acid excretion. The association found between 3 SNP with serum urate concentrations allowed us to construct an individual hyperuricemia-genetic risk score. We have shown that this additive genetic score had a strong and graded association with primary gout.

To the best of our knowledge, SNP analyses of genes that encode tubular urate transporters have not been reported in patients with gout classified according to their uric acid excretion rate related to their serum uric acid levels. In our study, according to the uric acid nomogram, only 15.4% of patients with gout were classified as normoexcretors. The mean daily uric acid excretion was remarkably higher in normoexcretors with gout than in underexcretors, despite a mean serum urate level 1.0 mg/dl lower in the former than in the latter (Table 2).

Patients with gout showed different allele distributions of the 5 SNP analyzed than did control subjects. However, the diagnosis of underexcretor was only positively associated with the presence of the T allele of *URAT1* rs11231825, the G allele of *GLUT9* rs16890979, and the A allele of *ABCG2* rs2231142, whereas no reabsorption transporter genetic loci were associated with normoexcretors with gout.

The SNP on secretion transporter ABCG2 showed a positive association with gout both in underexcretors and in normoexcretors, although the OR for normoexcretors was 10 times higher. Given that ABCG2 mediates tubular urate secretion<sup>23</sup>, it is likely that the dysfunction of this molecule most probably reduces uric acid excretion (Figure 1). The finding of dysfunctional variants of the urate secretion transporter ABCG2 among underexcretors with gout and the increased prevalence of the 2 tubular urate reabsorption transporter SNP studied herein may help explain the longlasting uric acid underexcretion reported in most patients with primary gout<sup>2,3</sup>. However, what explains the dysfunction in the urate secretion transporter ABCG2 in patients with normal uric acid excretion rates? Dysfunctional variants of ABCG2 have been reported as major causes of gout and hyperuricemia<sup>23,27</sup>. Recently, in a study of 644 men with hyperuricemia, the rate of uric acid excretion was found to be inversely related to the ABCG2

function<sup>28</sup>. ABCG2 is expressed not only in the kidneys<sup>29</sup>, but also in the apical membrane of several tissues, including the liver and intestines<sup>30</sup>. In those organs, ABCG2 may contribute to urate secretion<sup>31</sup>, provided that one-third of urate excretion in humans depends on gut excretion<sup>31,32,33</sup>. It is conceivable that the dysfunction of ABCG2 affects all organs in which it is expressed. The finding of an increased uric acid excretion rate in ABCG2-knockout mice<sup>34</sup> led to the proposition that ABCG2 dysfunction may, by decreasing extrarenal urate excretion ("extrarenal urate underexcretion"), increase the body urate pool, which would enhance urinary uric acid excretion<sup>34</sup>. This proposal has led to a new classification known as "overproduction" or "renal overload" hyperuricemia<sup>34</sup>, which may explain the mechanism of urate overproduction in a substantial proportion of patients with gout (15% in our study) who have no evidence of increased uric acid synthesis attributable to an enzyme defect.

The decreased prevalence of allele C of rs1165196 (NPT1) among patients with gout suggests that this allele is a protective, rather than a risk, allele that modulates the secretory expression of NPT1, reducing the likelihood of hyperuricemia and gout. The result of a significantly decreased prevalence of allele C of rs1165196 (NPT1) among normoexcretors with gout, but not among underexcretors with gout, may be interpreted in light of the apparent paradoxical result of ABCG2. SNP rs1165196 is a nonsynonymous polymorphism that causes a threonine 269 to isoleucine amino acid change in NPT1 protein. The NPT1 I269 protein (T allele) has been shown to transport about one-third less than the T269 NPT1 protein (C allele). At least 1 SLC17 member has been found to localize in the intestinal brush border membrane<sup>35</sup>; thus, it is possible to speculate that similar to ABCG2, NPT1 dysfunction may decrease extrarenal urate excretion. This fact could facilitate a new classification of patients with gout into 2 groups: renal urate underexcretors and extrarenal urate underexcretors (formerly normoexcretors).

The finding of a significant correlation between the genetic risk score and both serum urate concentrations and the prevalence of gout can best be interpreted as an indication that certain SNP markedly determine the kidneys' handling of urate. This suggests that knowledge of patients' genotypes could help identify individuals at risk of gout, such as those with metabolic syndrome or cardiovascular diseases<sup>36</sup>, long before the onset of clinical features, and may help guide clinical decisions, particularly when considering drugs known to increase serum urate levels<sup>37</sup>. One limitation of our study is that the results are related to 5 genes, among 9 urate transporter genes described to date $^{32}$ . However, to our knowledge, this is the largest study in which 5 urate transporter gene SNP were studied in a wellcharacterized cohort with gout according to their uric acid excretion rate. The reduced number of normoexcretors with

gout (n = 16, 15%) is in agreement with the proportion of those patients reported in most series<sup>2,3,4,38</sup>. Despite the reduced numbers, we found significant results that were markedly different from those of underexcretors with gout. A study with a similar number of participants from 3 ancestral groups recently concluded that genetic variations in *GLUT9* influence uric acid metabolism in European white subjects, but not in Maori and Pacific ethnic groups<sup>39</sup>. Another limitation of our study is that we assigned the same value in the genetic risk score to each allele. However, this methodology has been previously reported in other studies on gout<sup>6,26</sup>, and when the association between cholesterol polymorphisms and cardiovascular event risk has been examined<sup>40</sup>.

Our study shows that patients with primary gout, and normal or decreased uric acid excretion rates, have different prevalence rates for certain SNP related to tubular urate transporters. These results may contribute to a better understanding of the mechanism of hyperuricemia in primary gout.

### ACKNOWLEDGMENT

We thank the physicians who participated in the Grupo MAPA-Madrid for referring their patients to the Metabolic Unit. We are indebted to Carolina Velasco García, the research manager of the Metabolic Vascular Unit, and its nursing staff (Inés Narillos and Arantxa Sánchez) for their excellent patient care and followup; to Rosario Madero, PhD, for the statistical analysis; and to Almudena Ligos Díaz for her assistance in preparing the manuscript.

## REFERENCES

- 1. Terkeltaub R. Update on gout: new therapeutic strategies and options. Nat Rev Rheumatol 2010;6:30-8.
- Simkin PA. Uric acid excretion in patients with gout. Arthritis Rheum 1979;22:98–9.
- García Puig J, Mateos Antón F, López Jiménez M, Conthe Gutiérrez P. Renal handling of uric acid in gout: impaired tubular transport of urate not dependent on serum urate levels. Metabolism 1986;35:1147-53.
- 4. Anzai N, Jutabha P, Kimura T, Fukutomi T. Urate transport: relationship with serum urate disorder. Curr Rheumatol Rev 2011;7:123–31.
- Stark K, Reinhard W, Neureuther K, Wiedmann S, Sedlacek K, Baessler A, et al. Association of common polymorphisms in GLUT9 gene with gout but not with coronary artery disease in a large case-control study. PLoS One 2008;3:e1948.
- Dehghan A, Köttgen A, Yang Q, Hwang SJ, Kao WL, Rivadeneira F, et al. Association of three genetic loci with uric acid concentration and risk of gout: a genome-wide association study. Lancet 2008;372:1953-61.
- Kolz M, Johnson T, Sanna S, Teumer A, Vitart V, Perola M, et al. Meta-analysis of 28,141 individuals identifies common variants within five new loci that influence uric acid concentrations. PLoS Genet 2009;5:1-10.
- Hollis-Moffatt JE, Xu X, Dalbeth N, Merriman ME, Topless R, Waddell C, et al. Role of the urate transporter SLC2A9 gene in susceptibility to gout in New Zealand Maori, Pacific Island, and Caucasian case-control sample sets. Arthritis Rheum 2009; 60:3485-92.
- 9. Yang Q, Köttgen A, Dehghan A, Smith AV, Glazer NL, Chen MH,

et al. Multiple genetic loci influence serum urate levels and their relationship with gout and cardiovascular disease risk factors. Circ Cardiovasc Genet 2010;3:523-30.

- Tu HP, Chen CJ, Tovosia S, Ko AM, Lee CH, Ou TT, et al. Associations of a nonsynonymous variant in SCL2A9 with gouty arthritis and uric acid levels in the Han Chinese and Solomon Islanders. Ann Rheum Dis 2010;69:887-90.
- Urano W, Taniguchi A, Anzai N, Inoue E, Kanai Y, Yamanaka M, et al. Sodium-dependent phosphate cotransporter type 1 sequence polymorphisms in male patients with gout. Ann Rheum Dis 2010;69:1232-4.
- Köttgen A, Albrecht E, Teumer A, Vitart V, Krumsiek J, Hundertmark C, et al. Genome-wide association analyses identify 18 new loci associated with serum urate concentrations. Nat Genet 2013;45:145-54.
- Matsuo H, Chiba T, Nagamori S, Nakayama A, Domoto H, Phetdee K, et al. Mutations in glucose transporter 9 gene SLC2A9 cause renal hypouricemia. Am J Hum Genet 2008;83:744-51.
- Dinour D, Gray NK, Campbell S, Shu X, Sawyer L, Richardson W, et al. Homozygous SLC2A9 mutations cause severe renal hypouricemia. J Am Soc Nephrol 2010;21:64–72.
- Sebesta I, Stiburkova B, Bartl J, Ichida K, Hosoyamada M, Taylor J, et al. Diagnostic tests for primary renal hypouricemia. Nucleosides Nucleotides Nucleic Acids 2011;30:1112-6.
- Hollis-Moffatt JE, Phipps-Green AJ, Chapman B, Jones GT, van Rij A, Gow PJ, et al. The renal urate transporter SLC17A1 locus: confirmation of association with gout. Arthritis Res Ther 2012;14:R92.
- Shin J, Kim Y, Kong M, Lee C. Genetic architecture for susceptibility to gout in the KARE cohort study. J Hum Genet 2012;57:379-84.
- Wallace SL, Robinson H, Masi AT, Decker JL, McCarty DJ, Yü T-F. Preliminary criteria for the classification of the acute arthritis of primary gout. Arthritis Rheum 1977;20:895-900.
- Puig JG, Torres RJ, Mateos FA, Ramos TH, Arcas JM, Buno AS, et al. The spectrum of hypoxanthine-guanine phosphoribosyltransferase (HPRT) deficiency. Clinical experience based on 22 patients from 18 Spanish families. Medicine 2001;80:102-12.
- Torres RJ, Mateos FA, Puig JG, Becker MA. A simplified method for the determination of phosphoribosylpyrophosphate synthetase activity in hemolysates. Clin Chim Acta 1994;224:55-63.
- Puig JG, Torres RJ, de Miguel E, Sánchez A, Bailén R, Banegas JR. Uric acid excretion in normal subjects: A nomogram to assess the mechanisms underlying purine metabolic disorders. Metabolism 2012;61:512-8.
- 22. Mancia G, De Backer G, Dominiczak A, Cifkova R, Fagard R, Germano G, et al, ESH-ESC Task Force on the Management of Arterial Hypertension. 2007 ESH-ESC practice guidelines for the management of arterial hypertension: ESH-ESC task force on the management of arterial hypertension. J Hypertens 2007;25:1751-62.
- Woodward OM, Köttgen A, Coresh J, Boerwinkle E, Guggino WB, Köttgen M. Identification of a urate transporter, ABCG2, with a common functional polymorphism causing gout. Proc Natl Acad Sci USA 2009;106:10338-42.
- 24. Graessler J, Graessler A, Unger S, Kopprasch S, Tausche AK, Kuhlisch E, et al. Association of the human urate transporter 1 with reduced renal uric acid excretion and hyperuricemia in a German caucasian population. Arthritis Rheum 2006;54:292-300.

- 25. Rodriguez S, Gaunt TR, Day IN. Hardy-Weinberg equilibrium testing of biological ascertainment for Mendelian randomization studies. Am J Epidemiol 2009;169:505-14.
- Gunjaca G, Boban M, Pehlić M, Zemunik T, Budimir D, Kolcić I, et al. Predictive value of 8 genetic loci for serum uric acid concentration. Croat Med J 2010;51:23-31.
- 27. Matsuo H, Takada T, Ichida K, Nakamura T, Nakayama A, Ikebuchi Y, et al. Common defects of ABCG2, a high-capacity urate exporter, cause gout: a function-based genetic analysis in a Japanese population. Sci Transl Med 2009;1:5ra11.
- Hosomi A, Nakanishi T, Fujita T, Tamai I. Extra-renal elimination of uric acid via intestinal efflux transporter BCRP/ABCG2 decreased extra-renal urate excretion is a common cause of hyperuricemia. PLoS One 2012;7:e30456.
- 29. Huls M, Brown CD, Windass AS, Sayer R, van den Heuvel JJ, Heemskerk S, et al. The breast cancer resistance protein transporter ABCG2 is expressed in the human kidney proximal tubule apical membrane. Kidney Int 2008;73:220-5.
- Maliepaard M, Scheffer GL, Faneyte IF, van Gastelen MA, Pijnenborg AC, Schinkel AH, et al. Subcellular localization and distribution of the breast cancer resistance protein transporter in normal human tissues. Cancer Res 2001;61:3458-64.
- Sorensen LB. Role of the intestinal tract in the elimination of uric acid. Arthritis Rheum 1965;8:694-706.
- Anzai N, Jutabha P, Amonpatumrat-Takahashi S, Sakurai H. Recent advances in renal urate transport: characterization of candidate transporters indicated by genome-wide association studies. Clin Exp Nephrol 2012;16:89-95.
- Lipkowitz MS. Regulation of uric acid excretion by the kidney. Curr Rheumatol Rep 2012;14:179-88.
- Ichida K, Matsuo H, Takada T, Nakayama A, Murakami K, Shimizu T, et al. Decreased extra-renal urate excretion is a common cause of hyperuricemia. Nature Com 2012;3:764.
- 35. Togawa N, Miyaji T, Izawa S, Omote H, Moriyama Y. A Na+-phosphatecotransporter homologue (SLC17A4 protein) is an intestinal organic anion exporter. Am J Physiol Cell Physiol 2012;302:C1652-60
- López Jiménez M, Vigil Medina L, Condés Moreno E, García Carretero R, Fernández Mejias C, Ruiz Galiana J. [Uricemia and metabolic syndrome in patients with hypertension]. [Article in Spanish] Rev Clin Esp 2012;212:425-31.
- Armario P, Oliveras A, de la Sierra A. [Resistant hypertension]. [Article in Spanish] Rev Clin Esp 2013;213:388-93.
- Gutman AB, Yu T-F. Renal function in gout. With a commentary on the renal regulation of urate excretion, and the role of the kidney in the pathogenesis of gout. Am J Med 1957;23:600-22.
- Dalbeth N, House ME, Gamble GD, Horne A, Pool B, Purvis L, et al. Population-specific influence of SLC2A9 genotype on the acute hyperuricaemic response to a fructose load. Ann Rheum Dis 2013;72:1868-73.
- Kathiresan S, Melander O, Anevski D, Guiducci C, Burtt NP, Roos C, et al. Polymorphisms associated with cholesterol and risk of cardiovascular events. N Engl J Med 2008;58:1240-9.