Correlation of CX3CL1 and CX3CR1 Levels with Response to Infliximab Therapy in Patients with Rheumatoid Arthritis

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ABSTRACT. Objective. To examine the relation between serum chemokine levels and patient responsiveness to infliximab, and the influence of infliximab administration on serum chemokine levels.

Methods. Serum levels of the chemokines CX3CL1, CXCL8, CCL3, and CXCL10 were quantified prior to (at baseline) and after 30 weeks of treatment with infliximab in 20 patients using enzyme-linked immunosorbent assays. Disease status was assessed using the Disease Activity Score (DAS28). The response to infliximab was classified according to the European League Against Rheumatism (EULAR) response criteria.

Results. By 30 weeks, infliximab produced a significant overall reduction in DAS28 among the 20 patients with RA, although only 12 achieved a good to moderate response based on EULAR response criteria. A significant reduction in CX3CL1 was seen in the responsive group, although infliximab treatment had no significant effect on the serum levels of the other 3 chemokines. Comparison of patients with lower (< 2000 pg/ml) and higher (\geq 2000 pg/ml) basal CX3CL1 levels revealed that DAS28, erythrocyte sedimentation rate, C-reactive protein, and CX3CL1 levels were all significantly diminished by infliximab in RA patients with lower basal CX3CL1 levels, but not in those with higher basal levels. In addition, cell-surface expression of CX3CR1 protein in peripheral blood CD8+CD3+ T cells and mRNA expression of CX3CR1 in lymphocytes were both significantly downregulated after infliximab treatment in the responsive group.

Conclusion. Our results suggest that the CX3CL1-CX3CR1 system in patients with active RA may be sensitive to anti-tumor necrosis factor- α therapy, and confirm that CX3CL1 plays a crucial role in the pathogenesis of RA. (J Rheumatol First Release April 15 2009; doi:10.3899/jrheum.081074)

Key Indexing Terms: TUMOR NECROSIS FACTOR CHEMOKINE INHIBITOR RHEUMATOID ARTHRITIS

INFLIXIMAB

Rheumatoid arthritis (RA) is a systemic autoimmune disease characterized by inflammation of the synovial joints, infiltration by blood-derived cells, and eventual destruction of cartilage and bone, as well as various extraarticular signs and symptoms¹. The tumor necrosis factor- α (TNF- α) blocker infliximab is a chimeric IgG monoclonal antibody (mAb) that neutralizes both soluble and membrane-bound TNF in humans². Several clinical trials have provided compelling evidence of the safety and clinical efficacy of infliximab for both the induction and maintenance of remission in RA. But although infliximab often induces rapid clinical

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K. Wakabayashi, MD; T. Isozaki, MD, PhD; N. Yajima, MD, PhD; Y. Miwa, MD, PhD; T. Kasama, MD, PhD, Division of Rheumatology, Department of Internal Medicine, Showa University School of Medicine. Address reprint requests to Dr. T. Kasama, Division of Rheumatology, Department of Internal Medicine, Showa University School of Medicine, Tokyo, 142-8666, Japan. E-mail: tkasama@med.showa-u.ac.jp Accepted for publication January 15, 2009. improvement in patients with RA, 20%–40% of patients treated with infliximab show little or no clinical response^{3,4}. Further, these patients may experience severe adverse effects, and the cost of infliximab therapy is very high. Consequently, identification of early predictors of the response to infliximab would likely have important clinical and economic benefits for the patients with RA.

Chemokines are a family of small proteins that regulate cell migration into sites of inflammation⁵. An important advance in our understanding of the development and progression of RA was the recognition that chemokines expressed in the synovium of RA joints are key mediators of cell migration into the joints, and are thus key contributors to the pathogenesis of RA synovitis⁶⁻¹⁰. In that context, our aim in this study was to examine the relation between serum chemokine levels and patient responsiveness to infliximab, as well as the effect of infliximab administration on serum chemokine levels.

MATERIALS AND METHODS

Patients. Twenty patients with active RA and who fulfilled the 1987 American College of Rheumatology (ACR) criteria¹¹ were enrolled in our

study between March 2005 and December 2007. All patients had active RA that, according to ACR guidelines¹², failed to respond to treatment with methotrexate (MTX) or other disease modifying antirheumatic drugs (DMARD). No biological agents had been used in these patients, but glucocorticoids (< 10 mg/day of prednisolone) and nonsteroidal antiinflammatory drugs had to have been given at a stable dose for at least 4 weeks prior to enrollment and during the course of treatment in the study. Patients received intravenous infliximab at a dose of 3 mg/kg at baseline and at 2, 6, and 8-week intervals. Disease activity and its clinical improvement were assessed using Disease Activity Score (DAS28) [erythrocyte sedimentation rate (ESR) 4] with European League Against Rheumatism (EULAR) response criteria^{13,14}. Patients with moderate to good responses to infliximab therapy were defined as the responsive group, while patients showing no response were defined as the unresponsive group. Serum and peripheral blood cells were collected immediately prior to (baseline) and 30 weeks after starting infliximab therapy. Serum rheumatoid factor (RF), matrix metalloproteinase (MMP-3), C-reactive protein (CRP) levels, and ESR were determined using a latex photometric immunoassay and the Westergren method, respectively. Anti-cyclic citrullinated protein antibody (anti-CCP Ab) was measured using commercial double ligand ELISA kit (Diastat Anti-CCP; MBL, Tokyo, Japan).

All experiments were carried out in accord with protocols approved by the Human Subjects Research Committee at our institution, and informed consent was obtained from all patients and volunteers.

Determination of serum chemokine levels. Serum levels of the chemokines CXCL8 [interleukin 8 (IL-8)], CXCL10 (interferon-inducible protein 10), CCL3 (macrophage inflammatory protein-1 α), and CX3CL1 (fractalkine) were quantified using commercial ELISA kits according to the manufacturer's instructions (R&D Systems, Minneapolis, MN, USA).

Flow cytometry. Flow cytometric analysis of CX3CR1 expression on peripheral blood mononuclear cells (PBMC) was carried out as described¹⁵. PBMC from heparinized venous blood collected from patients and healthy volunteers were labeled first with anti-CD3-FITC, anti-CD4-PE, anti-CD8-PE, anti-CD14 (monocyte)-FITC (all from BD PharMingen, San Diego, CA, USA), or rabbit anti-CX3CR1 Ab (ProSci Inc., Poway, CA, USA) and then with a secondary Ab (biotin-conjugated anti-rabbit IgG) and tertiary reagent (CyChrome-conjugated streptavidin; BD PharMingen). The fluorescence intensity and percentages of positive cells were measured on a 3-color FACScan flow cytometer (Becton Dickinson, Mountain View, CA, USA).

Isolation of total RNA and real-time polymerase chain reaction (PCR). The monocyte fraction was isolated from PBMC using a monocyte isolation kit II (Miltenyi Biotec, Auburn, CA, USA), which included a cocktail of biotin-conjugated mAb against CD3, CD7, CD16, CD19, CD56, CD123, and glycophorin A. The purity of the CD14 expression was assessed with flow cytometry (> 95%). The remaining monocyte-depleted cell fraction was used as peripheral blood lymphocytes. Isolation of total cellular RNA from the lymphocyte and monocyte fractions and real-time PCR were all performed as described¹⁶. One-microgram samples of total RNA were reverse transcribed into cDNA using TaqMan RT reagents (Applied Biosystems, Foster City, CA, USA), after which real-time PCR was conducted using an ABI Prism® 7900 sequence detection system (Applied Biosystems). Detection of CX3CR1 and ribosomal RNA expression was accomplished using appropriate Assays-on-DemandTM primers and probes (Applied Biosystems). To make quantitative comparisons between different samples, a dilution series of pooled cDNA from unstimulated PBMC from healthy volunteers was used as internal standards for CX3CR1 and was loaded in each analysis. Maximum levels were assigned a value of 100 units.

Statistical analysis. Data were expressed as means \pm standard error of the mean. The differences between groups were evaluated using the Mann-Whitney U-test. Followup data were evaluated using Wilcoxon's test. The relationship between serum chemokine levels and the RA disease activity and indicated measures was evaluated using the Spearman rank correlation. Values of p < 0.05 were considered significant.

RESULTS

Kinetic study of serum chemokine levels in patients treated with infliximab. The patient characteristics are summarized in Table 1. At the start of therapy, the mean age of the patients was 55.4 years, disease duration was 5.9 years, and the baseline DAS28 was 5.79. Further, 19 patients were positive for anti-CCP Ab and RF. At baseline, there were no significant correlations between RA disease activity (DAS28) and RF, MMP-3, or anti-CCP Ab levels. There was also no significant correlation between the serum levels of any of the chemokines tested and either RA disease activity or serum measures including RF, MMP-3, and anti-CCP Ab levels (data not shown). Thirty weeks after the start of infliximab treatment, there was a significant overall reduction of RA disease activity (DAS28) from 5.79 \pm 0.29 to 4.45 \pm 0.27 (p < 0.05) among the patients. In addition, there was a significant reduction in serum CX3CL1 levels (baseline, 1926.7 ± 351.5 pg/ml vs 30 wks, 1533.3 ± 402.6 pg/ml; p < 0.05; Figure 1). Infliximab treatment had no significant effect on the serum levels of the other 3 chemokines tested (CXCL8, CXCL10, and CCL3), although levels of CXCL8 tended to be higher after infliximab treatment. Moreover, no significant reduction in serum levels of RF, MMP-3, and anti-CCP Ab levels were observed after 30 weeks of infliximab treatment (data not shown).

We next analyzed the relation between changes in serum chemokine levels and clinical responses to infliximab therapy. At 30 weeks, 12 patients with RA showed a moderate to good response to infliximab based on EULAR response criteria, while 8 showed no clinical improvement. There were no significant differences between the responsive and unresponsive groups with respect to mean age, disease duration, doses of either MTX or prednisolone, serum anti-CCP Ab levels, or RA disease activity (DAS28; Table 1). Basal CX3CL1 and CXCL8 levels tended to be lower in the responsive group (CX3CL1, 1494.1 ± 266.3 pg/ml; CXCL8, 57.8 ± 9.9) than in the unresponsive group (CX3CL1, 2575.6 \pm 755.7 pg/ml; CXCL8, 301.1 \pm 254.2), but the difference was not significant. On the other hand, there was a significant reduction in serum CX3CL1 levels in the responsive group $(1494.1 \pm 266.3 \text{ vs } 793.3 \pm 219.8 \text{ pg/ml}; \text{ p} < 0.005)$, but not in the unresponsive group (Figure 2A). No significant changes were seen in the levels of CCL3, CXCL10, or CXCL8 in either group (Figures 2B, 2C, and 2D).

Because the results summarized above suggest that serum CX3CL1 levels may be related to RA disease activity and/or responsiveness of infliximab, we assessed the responsiveness of patients with RA to infliximab after dividing them into 2 groups based on whether their serum CX3CL1 levels were higher or lower than 2000 pg/ml. That criterion was selected because mean basal serum CX3CL1 level was 1926.7 \pm 351.5 pg/ml for the entire study population. Comparison of patients with lower (< 2000 pg/ml, n = 13) and higher (\geq 2000 pg/ml, n = 7) basal CX3CL1 levels

Table 1. Patient characteristics.

Patients (male/female)	All, 20 (3/17)	Responders, 12 (2/10)	Nonresponders, 8 (1/7)	р
Disease duration, yrs	$5.90 \pm 1.95 (1-31)$	$3.08 \pm 0.5 (1-6)$	$10.1 \pm 4.58 (1-31)$	NS
Dosages of				
MTX, mg/wk	7.83 ± 0.34 (6–12.5)	7.50 ± 0.26 (6–8)	8.31 ± 0.75 (6–12.5)	NS
Prednisolone, mg/day	6.44 ± 0.44 (5–10)	$6.89 \pm 0.69 (5-10)$	$5.86 \pm 0.45 \ (5-7.5)$	NS
ESR, mm/h	67.8 ± 8.18 (8–131)	66.7 ± 9.48 (27–131)	$69.5 \pm 15.5 \ (8-125)$	NS
CRP, mg/dl	$3.74 \pm 1.05 \ (0-19.9)$	$3.75 \pm 1.60 \ (0-19.9)$	3.71 ± 1.18 (0.2–8.4)	NS
ГЈС	$9.1 \pm 1.5 (3-21)$	$9.75 \pm 1.70 \ (4-17)$	8.1 ± 2.8 (3–21)	NS
SJC	$4.2 \pm 1.0 (2-15)$	$7.2 \pm 1.1 \ (2-15)$	5.9 ± 1.4 (2–12)	NS
DAS28, ESR-4	$5.79 \pm 0.29 (3.59 - 7.61)$	$6.02 \pm 0.31 (3.95 - 7.61)$	$5.45 \pm 0.55 (3.59 - 7.59)$	NS
RF, IU/ml	$159.0 \pm 46.0 \ (0-841.9)$	$177.0 \pm 68.2 (28.9 - 841.9)$	$128.3 \pm 49.0 \ (0-373.6)$	NS
MMP-3, ng/ml	92.1 ± 54.7 (63.8–792)	385.1 ± 82.9 (92.1–792)	254.0 ± 58.1 (63.8–494)	NS
Anti-CCP Ab, U/ml	617.8 ± 303.5 (0-5950)	920.3 ± 502.1 (28-5950)	$164.0 \pm 53.1 \ (0-444)$	NS

MTX: methotrexate; ESR: erythrocyte sedimentation rate; CRP: C-reactive protein; TJC: tender joint count; SJC: swollen joint count; DAS28: Disease Activity Score 28; RF: rheumatoid factor; MMP-3: matrix metalloproteinase-3; anti-CCP Ab: anti-cyclic citrullinated protein antibody; NS: not significant.

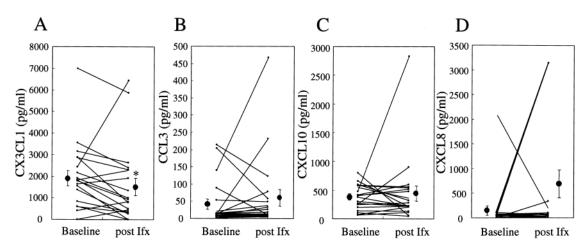


Figure 1. Changes in serum chemokine levels after infliximab administration. Paired serum samples from 20 patients with RA were collected at baseline and 30 wks after starting infliximab treatment. A: CX3CL1, B: CCL3, C: CXCL10, D: CXCL8. Each line represents an individual patient. Significant reduction of serum CX3CL1 was seen after infliximab treatment (*p< 0.05).

revealed no significant difference in DAS28, serum CRP levels, anti-CCP Ab levels, daily dosage of prednisolone, or weekly dosage of MTX (data not shown). DAS28, ESR, CRP, and CX3CL1 levels were all significantly reduced (DAS28 declined from 6.08 \pm 0.32 to 4.43 \pm 0.30, p < 0.0001; ESR, from 70.6 ± 8.9 to 48.2 ± 7.2, p < 0.01; CRP, from 4.2 \pm 1.5 to 1.2 \pm 0.4, p < 0.005; CX3CL1, from 1111.2 ± 200.7 to 682.7 ± 198.5 pg/ml, p < 0.05) in infliximab-treated RA patients with lower basal CX3CL1 levels (< 2000 pg/ml; Figure 3). By contrast, there were no significant reductions in DAS28, CRP, or CX3CL1 levels in patients with higher basal CX3CL1 levels ($\geq 2000 \text{ pg/ml}$; Figure 3). In addition, there was no significant reduction in serum levels of RF, MMP-3, and anti-CCP Ab levels after 30 weeks of infliximab treatment in patients with either higher or lower basal levels of CX3CL1 (data not shown).

Increased cell-surface expression of CX3CR1 protein and CX3CR1 mRNA in patients with RA and their reduction by infliximab therapy. CX3CR1, a specific CX3CL1 receptor, is also crucially involved in the dysregulation of CX3CL1 expression under conditions of inflammation. To examine the phenotype of cells expressing CX3CR1 protein and mRNA, we used flow cytometry and real-time PCR to analyze their expression in peripheral blood-specific cell populations from 6 patients with RA randomly selected at the start of infliximab therapy and from 8 healthy controls. Mean fluorescence intensity (MFI) reflecting CX3CR1 expression was significantly (p < 0.05) more pronounced on CD8+CD3+ T cells from patients with active RA than on those from healthy controls (Figures 4A and 4B). No such difference was seen with CD4+CD3+ T cells or macrophages. Among the 6 patients with RA, 3 showed a

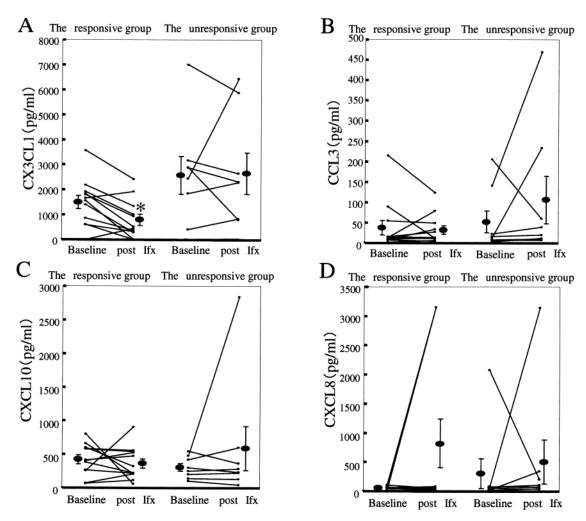


Figure 2. Serum chemokine levels between the responsive (n = 12) and unresponsive group (n = 8). Serum chemokines levels were measured at baseline and 30 wks after start of infliximab therapy. Clinical responses to infliximab were based on EULAR response criteria. A: CX3CL1, B: CCL3, C: CXCL10, D: CXCL8. Each line represents an individual patient. Significant reduction of serum CX3CL1 in the responsive group was seen after infliximab treatment (*p < 0.005).

moderate to good response 30 weeks after the start of infliximab therapy, while the other 3 showed little or no response. Notably, the MFI signals of CX3CR1 surface expression on CD8+CD3+ T cells were significantly reduced after infliximab treatment in the responsive group (Figure 4C), but not in the unresponsive group (Figure 4D). In contrast with CD8+CD3+ T cells, there was no significant alteration of CX3CR1 expression on either CD4+CD3+ T cells or macrophages (data not shown).

Consistent with the reduced expression of CX3CR1 protein in those responsive to infliximab, basal expression of CX3CR1 mRNA tended to be lower in peripheral blood lymphocytes from responsive patients with RA (203.0 \pm 22.7 units) than in those from the unresponsive group (489.7 \pm 138.4 units), although the difference was not statistically insignificant. In addition, expression of CX3CR1 mRNA was significantly reduced after infliximab treatment in lymphocytes from responsive patients (n = 3), but not from unresponsive ones (Figure 5).

DISCUSSION

CX3CL1, CXCL8, CXCL10, and CCL3 all play crucial roles in the pathogenesis of RA^{6,9,10}. We have shown that whereas serum levels of CXCL8, CXCL10, and CCL3 are unaffected by infliximab therapy, CX3CL1 levels decline in patients showing a clinical response to infliximab treatment. Moreover, patients with active RA who did not show a clinical response to infliximab showed higher basal CX3CL1 levels than those who did.

Expression of a variety of cytokines and chemokines, including IL- $6^{17,18}$, IL- 10^{19} , IL- 15^{20} , IL- 18^{21} , CXCL 16^{22} , and GRO α (CXCL1)²³ is reportedly blocked or diminished after infliximab therapy. Notably, all of these molecules are known to be induced in cells upon stimulation with

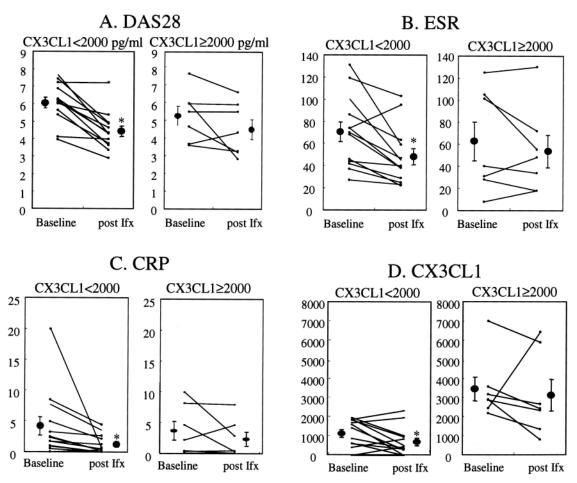


Figure 3. Effect of infliximab treatment on RA disease activity (DAS28), ESR, and serum CRP and CX3CL1 levels in RA patients with lower and higher basal CX3CL1 levels. Responsiveness to infliximab was statistically examined after dividing patients into 2 groups based on whether their basal CX3CL1 levels were higher or lower than 2000 pg /ml. DAS28 (A), ESR (B, mm/h), CRP (C, mg/dl), and CX3CL1 (D, pg/ml) levels were all significantly reduced (DAS28, p < 0.0001; ESR, p < 0.01; CRP, p < 0.005; CX3CL1, p < 0.05) in patients with lower basal CX3CL1 levels.

TNF- α^{24} , which suggests that inhibition by infliximab of TNF activity and its downstream cascade effectively suppresses production of these inflammatory cytokines and chemokines in patients with RA. Thus neutralization of endogenous TNF- α in patients with RA may underlie the clinical response to infliximab²⁵.

The major cellular source of CX3CL1 in the periphery is the endothelium, and endothelium-derived CX3CL1 likely plays a pivotal role in such pathological conditions as vascular inflammation, glomerulonephritis, and systemic vasculitis²⁶⁻²⁹. In an earlier study, we showed that impaired endothelial cells within vasculitis lesions express CX3CL1³⁰. In addition, it was also recently shown that infliximab treatment reverses endothelial dysfunction in patients with RA³¹, and that otherwise elevated levels of soluble adhesion molecules (intercellular adhesion molecule-1, vascular cell adhesion molecule-1, and E-selectin) and vascular endothelial growth factor were significantly reduced by administration of infliximab to patients with active RA^{19,32,33}. Thus infliximab may ameliorate endothelial cell activation, leading to downregulation of CX3CL1 expression.

Kageyama, et al reported that infliximab had no significant effect on serum CX3CL1 or CXCL10 levels in patients with RA²². We also found that infliximab had no significant effect on serum CXCL10 levels but, as discussed, we observed a significant reduction in CX3CL1 levels in some patients. Although the explanation for that difference in the response to infliximab is not clear, we did find that patients with basal serum CX3CL1 levels ≥ 2000 pg/ml were unresponsive to infliximab. Only when serum CX3CL1 levels were < 2000 pg/ml did infliximab have a beneficial effect in patients with active RA. Consistent with our findings are recent reports that RA disease activity is positively correlated with CX3CL1 levels in serum and synovial fluid, that expression of CX3CL1 is upregulated in inflamed synovial RA lesions, and that joint inflammation in an experimental arthritis model is reduced by neutralization of

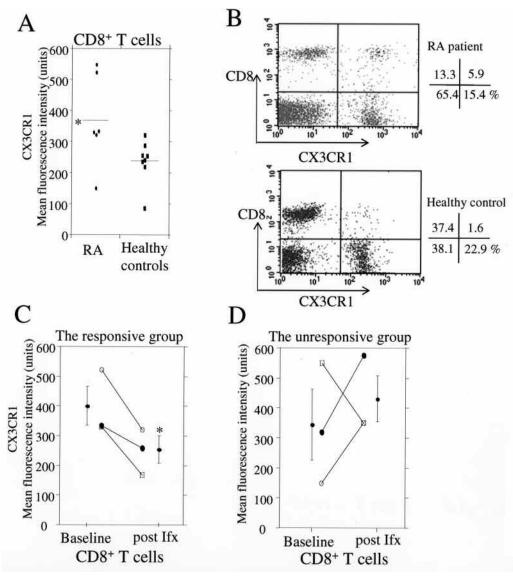


Figure 4. Cell-surface expression of CX3CR1 protein in PBMC from RA patients and its reduction after infliximab treatment. PBMC were obtained from 6 patients with RA and 8 healthy controls and labeled with anti-CD3+, anti-CD4+, anti-CD4+, anti-CD14+, and anti-CX3CR1 Ab. CX3CR1 expression on gated cells was then assayed by 3-color flow cytometry. A: Expression (mean fluorescence intensity; MFI) of CX3CR1 on CD8+CD3+ T cells from patients with active RA was significantly higher than in healthy controls. B: Representative dot plot graph of CX3CR1 expression on CD8+CD3+ T cells from patients and healthy controls. C and D: Expression of CX3CR1 on CD8+CD3+ T cells from 6 patients with RA [3 responsive (C) and 3 unresponsive (D) to infliximab therapy] randomly selected at the start of infliximab therapy. Data were expressed as MFI (units). Significant reductions in cell-surface CX3CR1 expression was seen in responsive group; p < 0.05 vs baseline.

CX3CL1^{30,34-37}. TNF antagonists such as infliximab, etanercept, and adalimumab have only limited efficacy against pathological conditions related to systemic vasculitis^{38,39}. We observed previously that serum CX3CL1 levels are significantly higher in patients with rheumatoid vasculitis or RA with extraarticular manifestations than in RA patients without vasculitis or other systemic manifestations³⁰. We therefore suggest that patients with RA showing higher levels of CX3CL1 in this study may have latent vascular inflammation or vasculitis, resulting in a limited response against infliximab. These findings suggest that basal serum CX3CL1 levels may correlate with the responsiveness of patients with RA to infliximab therapy.

Expression of CX3CL1 and its specific receptor, CX3CR1, on CD8+ T lymphocytes was suppressed following TNF- α neutralization with infliximab in the responsive group. This may reflect diminished interaction between the endothelium and synovium, resulting in reduced infiltration

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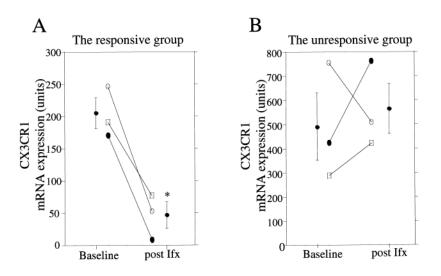


Figure 5. Expression of CX3CR1 mRNA in peripheral blood lymphocytes from patients with RA. Expression of CX3CR1 mRNA in peripheral blood lymphocytes from 6 patients [3 responsive (A) and 3 unresponsive (B) to infliximab therapy] randomly selected at the start of infliximab therapy. Data expressed in arbitrary units. Significant reductions in CX3CR1 mRNA were seen in responsive group; p < 0.05 vs baseline.

of leukocytes (CD8+ T cells) into the rheumatoid synovium, which in turn likely reflects diminished CX3CL1/CX3-CR1-mediated chemotactic effects on monocytes, natural killer cells, and T lymphocytes. In addition, the CX3-CL1/CX3CR1 complex can also act as an adhesion molecule able to promote the firm adhesion of a subset of leukocytes to endothelium under conditions of physiological flow and of rheumatoid synovitis^{34,40-42}.

It was recently shown that levels of IgM-RF antibodies are sometimes reduced following infliximab treatment, and that the progressive reduction of IgM-RF levels in patients treated with etanercept, another TNF antagonist, may have some prognostic importance⁴³. Braun-Moscovici, *et al* showed that basal levels of anti-CCP antibody serve as a predictor of infliximab therapy¹⁸. In contrast, we could not detect any significant correlations between clinical response to infliximab therapy and serum levels of either RF or anti-CCP Ab. Although we did not examine the prognostic value of these antibodies in this study, further investigation of serum CX3CL1 in combination with other serum measures in larger numbers of patients should enable more precise prediction of the efficacy of anti-TNF therapy.

Our results suggest that the CX3CL1-CX3CR1 system in patients with active RA may be sensitive to anti-TNF- α therapy and confirm that CX3CL1 plays a crucial role in the pathogenesis of RA, although further investigations in a larger sample of RA patients with anti-TNF- α therapy are required to understand the clinical significance of CX3-CL1-CX3CR1. They also suggest that basal serum CX3CL1 levels correlate with the responsiveness of patients with RA to infliximab therapy.

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REFERENCES

- 1. Firestein GS. Evolving concepts of rheumatoid arthritis. Nature 2003;423:356-61.
- Maini RN, Taylor PC. Anti-cytokine therapy for rheumatoid arthritis. Annu Rev Med 2000;51:207-29.
- Feldmann M, Maini RN. Anti-TNF alpha therapy of rheumatoid arthritis: what have we learned? Annu Rev Immunol 2001;19:163-96.
- Scott DL, Kingsley GH. Tumor necrosis factor inhibitors for rheumatoid arthritis. N Engl J Med 2006;355:704-12.
- Gerard C, Rollins BJ. Chemokines and disease. Nature Immunol 2001;2:108-15.
- Kunkel SL, Lukacs NW, Kasama T, Strieter RM. The role of chemokines in inflammatory joint disease. J Leukocyte Biol 1996;58:6-12.
- Jenkins JK, Hardy KJ, McMurray RW. The pathogenesis of rheumatoid arthritis: a guide to therapy. Am J Med Sci 2002;323:171-80.
- 8. Haringman JJ, Ludikhuize J, Tak PP. Chemokines in joint disease: the key to inflammation? Ann Rheum Dis 2004;63:1186-94.
- Chen X, Oppenheim JJ, Howard OM. Chemokines and chemokine receptors as novel therapeutic targets in rheumatoid arthritis (RA): inhibitory effects of traditional Chinese medicinal components. Cell Mol Immunol 2004;1:336-42.
- 10. Szekanecz Z, Szucs G, Szanto S, Koch AE. Chemokines in rheumatic diseases. Curr Drug Targets 2006;7:91-102.
- 11. Arnett FC, Edworthy SM, Bloch DA, et al. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. Arthritis Rheum 1988;31:315-24.
- 12. Guidelines for the management of rheumatoid arthritis: 2002 Update. Arthritis Rheum 2002;46:328-46.
- 13. van der Heijde DM, van 't Hof MA, van Riel PL, et al. Judging disease activity in clinical practice in rheumatoid arthritis: first step

Odai, et al: Infliximab in RA

in the development of a disease activity score. Ann Rheum Dis 1990;49:916-20.

- 14. van Gestel AM, Prevoo ML, van 't Hof MA, van Rijswijk MH, van de Putte LB, van Riel PL. Development and validation of the European League Against Rheumatism response criteria for rheumatoid arthritis. Comparison with the preliminary American College of Rheumatology and the World Health Organization/International League Against Rheumatism Criteria. Arthritis Rheum 1996;39:34-40.
- 15. Yajima N, Kasama T, Isozaki T, et al. Elevated levels of soluble fractalkine in active systemic lupus erythematosus: Potential involvement in neuropsychiatric manifestations. Arthritis Rheum 2005;52:1670-5.
- Kasama T, Isozaki T, Odai T, et al. Expression of angiopoietin-1 in osteoblasts and its inhibition by tumor necrosis factor-alpha and interferon-gamma. Transl Res 2007;149:265-73.
- Charles P, Elliott MJ, Davis D, et al. Regulation of cytokines, cytokine inhibitors, and acute-phase proteins following anti-TNF-alpha therapy in rheumatoid arthritis. J Immunol 1999;163:1521-8.
- Braun-Moscovici Y, Markovits D, Zinder O, et al. Anti-cyclic citrullinated protein antibodies as a predictor of response to anti-tumor necrosis factor-alpha therapy in patients with rheumatoid arthritis. J Rheumatol 2006;33:497-500.
- Macias I, Garcia-Perez S, Ruiz-Tudela M, Medina F, Chozas N, Giron-Gonzalez JA. Modification of pro- and antiinflammatory cytokines and vascular-related molecules by tumor necrosis factor-α blockade in patients with rheumatoid arthritis. J Rheumatol 2005;32:2102-8.
- Kageyama Y, Takahashi M, Torikai E, et al. Treatment with anti-TNF-alpha antibody infliximab reduces serum IL-15 levels in patients with rheumatoid arthritis. Clin Rheumatol 2007;26:505-9.
- 21. Pittoni V, Bombardieri M, Spinelli FR, et al. Anti-tumour necrosis factor (TNF) alpha treatment of rheumatoid arthritis (infliximab) selectively downregulates the production of interleukin (IL) 18 but not of IL12 and IL13. Ann Rheum Dis 2002;61:723-5.
- Kageyama Y, Torikai E, Nagano A. Anti-tumor necrosis factor-alpha antibody treatment reduces serum CXCL16 levels in patients with rheumatoid arthritis. Rheumatol Int 2007;27:467-72.
- 23. Torikai E, Kageyama Y, Suzuki M, Ichikawa T, Nagano A. The effect of infliximab on chemokines in patients with rheumatoid arthritis. Clin Rheumatol 2007;26:1088-93.
- 24. McDermott MF. TNF and TNFR biology in health and disease. Cell Mol Biol 2001;47:619-35.
- Edrees AF, Misra SN, Abdou NI. Anti-tumor necrosis factor (TNF) therapy in rheumatoid arthritis: correlation of TNF-alpha serum level with clinical response and benefit from changing dose or frequency of infliximab infusions. Clin Exp Rheumatol 2005;23:469-74.
- 26. Chapman GA, Moores KE, Gohil J, et al. The role of fractalkine in the recruitment of monocytes to the endothelium. Eur J Pharmacol 2000;392:189-95.
- 27. Furuichi K, Wada T, Iwata Y, et al. Upregulation of fractalkine in human crescentic glomerulonephritis. Nephron 2001;87:314-20.
- Cockwell P, Chakravorty SJ, Girdlestone J, Savage CO. Fractalkine expression in human renal inflammation. J Pathol 2002;196:85-90.

- Bjerkeli V, Damas JK, Fevang B, Holter JC, Aukrust P, Froland SS. Increased expression of fractalkine (CX3CL1) and its receptor, CX3CR1, in Wegener's granulomatosis: possible role in vascular inflammation. Rheumatology 2007;46:1422-7.
- Matsunawa M, Isozaki T, Odai T, et al. Increased serum levels of soluble fractalkine (CX3CL1) correlate with disease activity in rheumatoid vasculitis. Arthritis Rheum 2006;54:3408-16.
- Cardillo C, Schinzari F, Mores N, et al. Intravascular tumor necrosis factor alpha blockade reverses endothelial dysfunction in rheumatoid arthritis. Clin Pharmacol Ther 2006;80:275-81.
- 32. Klimiuk PA, Sierakowski S, Domyslawska I, Fiedorczyk M, Chwiecko J. Reduction of soluble adhesion molecules (sICAM-1, sVCAM-1, and sE-selectin) and vascular endothelial growth factor levels in serum of rheumatoid arthritis patients following multiple intravenous infusions of infliximab. Arch Immunol Ther Exp (Warsz) 2004;52:36-42.
- Strunk J, Bundke E, Lange U. Anti-TNF-alpha antibody infliximab and glucocorticoids reduce serum vascular endothelial growth factor levels in patients with rheumatoid arthritis: a pilot study. Rheumatol Int 2006;26:252-6.
- 34. Ruth JH, Volin MV, Haines GK 3rd, et al. Fractalkine, a novel chemokine in rheumatoid arthritis and in rat adjuvant-induced arthritis. Arthritis Rheum 2001;44:1568-81.
- Volin MV, Woods JM, Amin MA, Connors MA, Harlow LA, Koch AE. Fractalkine: a novel angiogenic chemokine in rheumatoid arthritis. Am J Pathol 2001;159:1521-30.
- Blaschke S, Koziolek M, Schwarz A, et al. Proinflammatory role of fractalkine (CX3CL1) in rheumatoid arthritis. J Rheumatol 2003;30:1918-27.
- Nanki T, Urasaki Y, Imai T, et al. Inhibition of fractalkine ameliorates murine collagen-induced arthritis. J Immunol 2004;173:7010-6.
- Aries PM, Lamprecht P, Gross WL. Biological therapies: new treatment options for ANCA-associated vasculitis? Exp Opin Biol Ther 2007;7:521-33.
- Lin J, Ziring D, Desai S, et al. TNF-alpha blockade in human diseases: an overview of efficacy and safety. Clin Immunol 2008;126:13-30.
- Imai T, Hieshima K, Haskell C, et al. Identification and molecular characterization of fractalkine receptor CX3CR1, which mediates both leukocyte migration and adhesion. Cell 1997;91:521-30.
- Nanki T, Imai T, Nagasaka K, et al. Migration of CX3CR1-positive T cells producing type 1 cytokines and cytotoxic molecules into the synovium of patients with rheumatoid arthritis. Arthritis Rheum 2002;46:2878-83.
- 42. Volin MV, Huynh N, Klosowska K, Chong KK, Woods JM. Fractalkine is a novel chemoattractant for rheumatoid arthritis fibroblast-like synoviocyte signaling through MAP kinases and Akt. Arthritis Rheum 2007;56:2512-22.
- 43. Vis M, Bos WH, Wolbink G, et al. IgM-rheumatoid factor, anti-cyclic citrullinated peptide, and anti-citrullinated human fibrinogen antibodies decrease during treatment with the tumor necrosis factor blocker infliximab in patients with rheumatoid arthritis. J Rheumatol 2008;35:425-8.