

The Functional Polymorphism 844 A>G in Fc_αRI (CD89) Does Not Contribute to Systemic Sclerosis or Rheumatoid Arthritis Susceptibility

JASPER C.A. BROEN, MARIEKE J.H. COENEN, BLANCA RUEDA, TORSTEN WITTE, LEONID PADYUKOV, LARS KLAESKOG, ROGER HESSELSTRAND, DIRK M. WUTTGE, CARMEN SIMEON, NORBERTO ORTEGO-CENTENO, MIGUEL A. GONZÁLEZ-GAY, ANNA PROS, NICHOLAS HUNZELMAN, GABRIELA RIEMEKASTEN, ALEXANDER KREUTER, MADELON VONK, RAFAELLA SCORZA, LORENZO BERETTA, PAULO AIRÒ, PIET L.C.M. van RIEL, ROBERT KIMBERLY, JAVIER MARTIN, JEFFREY EDBERG, and TIMOTHY R.D.J. RADSTAKE

ABSTRACT. Objective. To investigate the role of the Fc_αRI 844 A>G functional polymorphism in the genetic predisposition to rheumatoid arthritis (RA) and systemic sclerosis (SSc) susceptibility.

Methods. The study population was composed of 1401 patients with SSc, 642 patients with RA, and 1317 healthy controls. The Fc_αRI (CD89) single-nucleotide polymorphism rs16986050 was genotyped by pyrosequencing.

Results. We observed no significant deviation of the genotype and allele frequencies in RA and SSc compared to controls. A metaanalysis and a recessive and dominant model yielded similar negative results.

Conclusion. Our data show that the Fc_αRI 844 A>G polymorphism is not associated with SSc or RA susceptibility. (First Release Dec 15 2010; J Rheumatol 2011;38:446–9; doi:10.3899/jrheum.100427)

Key Indexing Terms:

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From the Radboud University Nijmegen Medical Center, Departments of Rheumatology and Human Genetics, Nijmegen, The Netherlands; Instituto de Parasitología y Biomedicina López-Neyra, CSIC, Granada; Servicio de Medicina Interna, Hospital Clínico Universitario, Granada; Servicio de Medicina Interna, Hospital Valle de Hebron, Barcelona; Hospital del Mar, Barcelona; and Servicio de Reumatología, Hospital Xeral-Calde, Lugo, Spain; Department of Medicine, Clinic for Immunology and Rheumatology, Hannover Medical School, Hannover; Department of Dermatology, University of Cologne, Cologne; Department of Rheumatology and Clinical Immunology, Charité University Hospital, and German Rheumatism Research Centre, Berlin; and Department of Rheumatology, Ruhr University of Bochum, Bochum, Germany; Referral Center for Systemic Autoimmune Diseases, Fondazione IRCCS Ospedale, Milan; and Servizio di Reumatologia ed Immunologia Clinica Spedali Civili, Brescia, Italy; Division of Clinical Immunology and Rheumatology, Department of Medicine, Division of Rheumatology at Karolinska University Hospital, Stockholm; and Department of Rheumatology, Lund University Hospital, Lund, Sweden; and Department of Medicine, University of Alabama at Birmingham, Birmingham, Alabama, USA.

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J.C.A. Broen, MSc, Department of Rheumatology, Radboud University Nijmegen Medical Center; M.J.H. Coenen, PhD, Department of Human Genetics, Radboud University Nijmegen Medical Center; B. Rueda, PhD, Instituto de Parasitología y Biomedicina López-Neyra, CSIC; T. Witte, MD, PhD, Department of Medicine, Clinic for Immunology and

Rheumatology, Hannover Medical School; L. Padyukov, PhD, Department of Medicine, Division of Rheumatology, Karolinska University Hospital; L. Klareskog, MD, PhD, Department of Medicine, Division of Rheumatology, Karolinska University Hospital; R. Hesselstrand, MD; D.M. Wuttge, MD, Department of Rheumatology, Lund University Hospital; C. Simeon, MD, Servicio de Medicina Interna, Hospital Valle de Hebron; N. Ortego-Centeno, MD, for the Spanish Systemic Sclerosis group, Servicio de Medicina Interna, Hospital Clínico Universitario; M.A. González-Gay, MD, Servicio de Reumatología, Hospital Xeral-Calde; A. Pros, MD, Hospital del Mar; N. Hunzelman, MD, PhD, Department of Dermatology, University of Cologne; G. Riemekasten, MD, PhD, Department of Rheumatology and Clinical Immunology, Charité University Hospital, and German Rheumatism Research Centre; A. Kreuter, MD, PhD, Department of Rheumatology, Ruhr University of Bochum; M. Vonk, MD, PhD, Radboud University Nijmegen Medical Center, Department of Rheumatology; R. Scorza, MD, PhD; L. Beretta, MD, PhD, Referral Center for Systemic Autoimmune Diseases, Fondazione IRCCS Ospedale; P. Airò, MD, PhD, Servizio di Reumatologia ed Immunologia Clinica Spedali Civili; P.L.C.M. van Riel, MD, PhD, Department of Rheumatology, Radboud University Nijmegen Medical Center; R. Kimberly, MD, Division of Clinical Immunology and Rheumatology, Department of Medicine, University of Alabama at Birmingham; J. Martin, MD, PhD, Instituto de Parasitología y Biomedicina López-Neyra, CSIC; J. Edberg, PhD, Division of Clinical Immunology and Rheumatology, Department of Medicine, University of Alabama at Birmingham; T.R.D.J. Radstake, MD, PhD, Department of Rheumatology, Radboud University Nijmegen Medical Center.

Address correspondence to Dr. T.R.D.J. Radstake, Department of Rheumatology, Radboud University Nijmegen Medical Centre, Geert Grooteplein 8, 6500 HB Nijmegen, The Netherlands. E-mail: Tradstake73@gmail.com

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Fc receptors (FcR) play a pivotal role in linking humoral and cellular components of immunity by bringing about the recognition of antigens bound to immunoglobulins (Ig). There is a large body of evidence describing genetic variations in FcR that were found to be associated with a wide range of autoimmune pathologies, such as systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA)¹. Although disease-specific IgA is present in autoimmune disease, its effect remains the least scrutinized. IgA exhibits its immunoregulatory functions mainly by triggering cellular effector functions through the Fc- α receptor (Fc α R) on the cell surface. Fc α RI (CD89) is the functionally most important IgA receptor and is expressed on various cells of the immune system². Serum IgA regulates secretion of interleukin 1 β (IL-1 β), IL-10, tumor necrosis factor- α (TNF- α), IL-6, and IL-1RA. To mediate these effects, the FcR-associated signal-transducing transmembrane γ -chain (FCGR) needs to be present in complex with Fc α RI³. Intriguingly, a polymorphism (844 A>G, rs16986050) in the coding region of Fc α RI leads to an amino acid change of serine 248 to glycine (S248G), which stimulates IL-6 production and induces cytokine release in the absence of the FCGR chain⁴. This variant was found to be enriched in 2 populations with SLE compared to healthy controls⁵. In other autoimmune diseases, specific IgA antibodies are also present. For instance, antitopoisomerase IgA and rheumatoid factor (RF) IgA and anti-citrullinated protein (ACPA) IgA are present in systemic sclerosis (SSc) and RA^{6,7}. Although the exact role of IgA in these diseases remains to be elucidated, we hypothesized that an increased inflammatory response upon disease-specific IgA binding caused by this polymorphism could contribute to these diseases. For this reason we investigated the frequency of this variant in patients with SSc and RA.

MATERIALS AND METHODS

Study population. The study population was composed of 1401 patients with SSc, 642 Dutch patients with RA, and 1317 healthy controls derived from blood donors. The subjects were matched demographically and by age and sex. Because SSc is a rare disease, we composed a cohort of 5

case-control sets of Europeans (Table 1). All patients fulfilled the American College of Rheumatology (ACR) 1980 classification criteria for SSc⁸. The local ethics committee from each center approved the study. Patients and controls provided written informed consent. All patients included in our study were classified as having limited cutaneous SSc (lcSSc) or diffuse cutaneous SSc (dcSSc), using the criteria proposed by LeRoy, *et al*⁹. Further information on clinical phenotype was recorded as well. Autoantibody testing was performed in every center separately using either ELISA or immunofluorescence microscopy (Table 1). Our study included RA patients who met the ACR 1987 revised criteria for RA¹⁰ (Table 2).

Genotyping. The Fc α RI 844 A>G variant was genotyped by pyrosequencing, congruent with our previous reports. Pyrosequencing reactions were performed according to the manufacturer's instructions on a PSQ-HS96A system (Biotage, Uppsala, Sweden)^{5,11}.

Statistical analysis. Significance levels were calculated with Fisher's exact test. P values < 0.05 were considered significant after Bonferroni adjustment. Homogeneity of OR was assessed with Breslow-Day statistics. Pooled OR were calculated under a fixed-effects model (Mantel-Haenszel). In the SSc population (n = 1401), our study reaches a power of 80% to detect an OR of 1.21. The estimation of the power for the RA population (n = 645) is 80% to detect an OR of 1.31.

RESULTS

After genotyping, no divergence in Hardy-Weinberg equilibrium was observed. We observed no significant deviation in genotype and allele frequencies of the Fc α RI 844 A>G polymorphism in Dutch patients with RA compared to Dutch controls and when comparing patients who were ACPA-positive or negative and patients with RA who were RF-positive or negative (Table 3). No significant hetero-

Table 2. Clinical characteristics of the study patients with rheumatoid arthritis (n = 642).

Characteristic	
Age, yrs (SD)	65 (13)
Women, %	66.5
RF-positive, %	78
Anti-CCP-positive, %*	65

* Status available for 155 patients. RF: rheumatoid factor; anti-CCP: anti-cyclic citrullinated peptide.

Table 1. Clinical characteristics of the study patients with systemic sclerosis.

Population	Netherlands	Spain	Germany	Sweden	Italy
No.	143	231	422	161	444
Age, yrs (SD)	56 (13)	55 (13)	57 (12)	53 (15)	54 (13)
Disease duration, mo (SD)	132 (87)	143 (90)	112 (109)	83 (73)	146 (138)
Women, %	73.2	83.8	83.2	78.8	89.0
Limited phenotype, %	74.8	73.7	61.2	72.0	74.4
ANA-positive, %	73.2	85.4	83.2	75.1	81.6
Anti-topo I-positive, %	22	NA	24.8	16.7	25.2
ACA-positive, %	26.8	46.7	41.3	26.1	47.1
Controls					
Age, yrs (SD)	45 (12)	56 (14)	48 (9)	52 (11)	52 (14)
Women, %	69.3	85.1	82.4	80.2	85.0

ANA: antinuclear antibodies; anti-topo I: antitopoisomerase I antibodies; ACA: anticentromere antibodies.

geneity was detected between the 5 European populations, justifying a metaanalysis. Initially, we observed a divergence in genotype distribution in the Swedish ($p = 0.034$) and Spanish ($p = 0.019$) populations with SSc. The same was observed for lcSSc susceptibility in both the Swedish ($p = 0.016$) and Spanish ($p = 0.022$) populations with SSc. The $Fc_{\alpha}RI$ 844G allele was associated with an increased risk of SSc, lcSSc, and anticentromere antibody (ACA)-positive SSc in the Swedish cohort (respectively, $p = 0.019$, 0.030 , and 0.032). The 844G allele was found less frequently in the Italian ACA-positive patients with SSc ($p = 0.042$; Table 3). However, after correction for multiple testing, no result remained significant. In addition, a metaanalysis and recessive and dominant models yielded similar negative results (Table 3 and data not shown).

DISCUSSION

We show that a common polymorphism in the coding region of $Fc_{\alpha}RI$ is not associated with RA or SSc susceptibility in 2 large cohorts. Considering the power of our study to detect significant deviations in allele frequencies of the $Fc_{\alpha}RI$ 844G variant between cases and controls, it is unlikely that the lack of association is due to a type 2 error. This indicates that the $Fc_{\alpha}RI$ 844 A>G polymorphism does not play a role in the susceptibility to RA and SSc and does not influence clinical phenotype. This is in contrast to the previous association of the $Fc_{\alpha}RI$ 844 A>G polymorphism with SLE susceptibility⁵. A number of polymorphisms have been found to influence susceptibility to RA as well as SSc and SLE^{12,13}. These polymorphisms therefore form merely a genetic foundation for autoimmunity in general. Intriguingly, the poly-

Table 3. Genotype frequencies of the $Fc_{\alpha}RI$ 844 A>G polymorphism in 5 European systemic sclerosis populations and a Dutch rheumatoid arthritis population.

Origin	Subtype	Total, n	AA, %	AG, %	GG, %	P vs Control	A, %	G, %	p vs Control
Italy	SSc	444	0.76	0.22	0.02	0.484	0.87	0.13	0.277
	lcSSc	327	0.75	0.24	0.01	0.253	0.87	0.13	0.211
	dcSSc	112	0.74	0.21	0.05	0.409	0.85	0.15	0.903
	ACA+	144	0.80	0.19	0.01	0.123	0.90	0.10	0.042
	Anti-topo+	146	0.71	0.27	0.01	0.631	0.85	0.15	0.923
	Control	362	0.73	0.25	0.02		0.85	0.15	
Sweden	SSc	161	0.58	0.30	0.12	0.034	0.73	0.27	0.019
	lcSSc	116	0.60	0.26	0.14	0.016	0.73	0.27	0.030
	dcSSc	45	0.53	0.40	0.07	0.252	0.73	0.27	0.107
	ACA+	44	0.55	0.32	0.13	0.055	0.70	0.30	0.039
	Anti-topo+	25	0.64	0.28	0.08	0.712	0.78	0.22	0.591
	Control	165	0.67	0.29	0.04		0.81	0.19	
Spain	SSc	231	0.66	0.26	0.08	0.019	0.79	0.21	0.639
	lcSSc	157	0.66	0.25	0.09	0.022	0.79	0.21	0.794
	dcSSc	56	0.62	0.29	0.09	0.274	0.76	0.24	0.900
	ACA+	89	0.66	0.28	0.06	0.388	0.80	0.20	0.525
	Anti-topo+	NA	NA	NA	NA		NA	NA	
	Control	250	0.60	0.36	0.04		0.78	0.22	
Germany	SSc	422	0.64	0.31	0.05	0.564	0.79	0.21	0.680
	lcSSc	261	0.68	0.29	0.03	0.532	0.82	0.18	0.612
	dcSSc	160	0.63	0.32	0.05	0.642	0.79	0.21	0.722
	ACA+	78	0.69	0.25	0.06	0.936	0.81	0.19	0.825
	Anti-topo+	50	0.58	0.34	0.08	0.616	0.75	0.25	0.507
	Control	266	0.66	0.28	0.06		0.80	0.20	
Netherlands	SSc	143	0.69	0.28	0.03	0.434	0.83	0.17	0.769
	lcSSc	98	0.72	0.24	0.04	0.983	0.84	0.16	0.999
	dcSSc	33	0.61	0.39	0.00	0.09	0.80	0.20	0.484
	ACA+	34	0.76	0.24	0.00	0.455	0.88	0.12	0.383
	Anti-topo+	33	0.70	0.27	0.03	0.860	0.83	0.17	0.862
	RA	642	0.71	0.26	0.03	0.593	0.84	0.16	0.582
Total Mantel-Haenszel	Control	274	0.72	0.24	0.04		0.84	0.16	
	SSc	1401	0.68	0.27	0.05	0.662	0.82	0.18	0.675
	lcSSc	959	0.68	0.26	0.06	0.842	0.81	0.19	0.825
	dcSSc	406	0.63	0.32	0.05	0.102	0.79	0.21	0.192
	ACA+	389	0.72	0.24	0.04	0.278	0.84	0.16	0.287
	Anti-topo+	254	0.68	0.28	0.04	0.394	0.82	0.18	0.547
Control	1317	0.68	0.28	0.04		0.82	0.18		

SSc: systemic sclerosis; lcSSc: limited cutaneous systemic sclerosis; dcSSc: diffuse cutaneous systemic sclerosis; RA: rheumatoid arthritis; ACA: anticentromere antibodies; anti-topo: antitopoisomerase antibodies; NA: not analyzed.

morphism we investigated seems to be specific for SLE. An explanation for this can perhaps be found in the properties of IgA in these 3 conditions. In contrast to RA and SSc, mean total IgA has been found to be significantly elevated in SLE compared to controls, and IgA-mediated inflammation may thus play a proportionally larger role in SLE^{14,15}. IgA ACPA antibodies are present in 29% of the overall patients with RA and in 47% of the patients with RA who have IgG anti-CCP antibodies. In SSc, IgA anti-topoisomerase antibodies have been described in 26.6% in a study containing 45 patients^{15,16}. However, this implies that in a subgroup of patients with SSc or RA who have high IgA antibody titers, which has been described in patients with RA who smoke, an effect of this variant may still be present¹⁵. These data were not available from the cohorts described in our study. In addition, autoantibody measurements have been performed in separate centers with either ELISA or immunofluorescence microscopy. Caution is warranted when comparing outcomes from these techniques¹⁷. Future investigations into the role of IgA and the Fc_αRI gene might therefore benefit from taking this observation into account and focusing on subgroups with high IgA titers. Altogether, we could not demonstrate a role for the Fc_αRI 844 A>G variant in SSc and RA.

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